Nerve conduction during Wallerian degeneration in the baboon

R. W. GILLIATT AND R. J. HJORST

From The Institute of Neurology, Queen Square, London

SUMMARY Conduction in the lateral popliteal nerve of the baboon was studied during the course of Wallerian degeneration. Six nerves were examined. In each case the muscle response to nerve stimulation and the ascending nerve action potential were recorded daily until the nerve became inexcitable. The muscle response to nerve stimulation disappeared after four to five days, but ascending nerve action potentials could be recorded for a further two to three days. There was no change in maximal motor conduction velocity or in distal latency until the muscle response to nerve stimulation was severely reduced in amplitude. At this stage there was a consistent increase in distal latency, sometimes associated with a mild reduction in maximal motor velocity in the leg. There was no change in the velocity of ascending nerve action potentials. Histological studies confirmed the presence of degeneration in the terminal parts of the intramuscular nerve fibres at a time when the proximal parts of the same fibres were relatively normal.

Conduction in peripheral nerves during Wallerian degeneration has been studied in small animals such as rats and guinea-pigs (Gutmann and Holubár, 1950, 1952; Kaeser and Lambert, 1962) and in cats and dogs (Rosenblueth and Dempsey, 1939; Erlanger and Schoepfle, 1946) but surprisingly little information is available for primates. In eight patients with traumatic nerve transections Landau (1953) stimulated the peripheral portion of the cut nerves and recorded the time after injury at which the muscle response to nerve stimulation disappeared. No electrophysiological records of response amplitude or latency were made. Gilliatt and Taylor (1959) studied the response of the facial muscles to nerve stimulation in three patients after surgical division of the nerve for the relief of hemifacial spasm. Response amplitude and latency were recorded but, in view of the hemifacial spasm, these nerves cannot be regarded as entirely normal before section; indeed one of them had been crushed in a previous surgical attempt to relieve the spasm.

In sub-human primates even fewer observations are available. Landau included observations of one macaque in his paper on human nerve lesions. The only other relevant observation we have found in the literature is that of Heinbecker, Bishop, and O'Leary (1932) on excised nerves from a macaque two days after section.

Recent work on toxic neuropathy in the baboon has renewed interest in the time-course of Wallerian degeneration. In the present experiments conduction in the lateral popliteal nerve of the baboon has been studied on successive days after nerve section and the results correlated with the anatomical changes.

METHODS

The experiments were carried out on sexually mature female baboons (Papio papio). The lateral popliteal nerve was divided under aseptic conditions in the lower thigh. The site of division was subsequently checked at necropsy; in each case it was 5 to 10 cm proximal to the site of stimulation at the neck of the fibula. Anaesthesia for nerve section and for electrophysiological studies was provided by intramuscular phencyclidine hydrochloride and promazine, supplemented by intravenous pentobarbitone. This form of anaesthesia did not give rise to side-effects when repeated daily in individual animals for six to nine days.

For the measurement of maximal motor conduction velocity, supramaximal shocks were delivered to the lateral popliteal nerve at the neck of the fibula, and to the anterior tibial nerve on the dorsum of the ankle, muscle action potentials being recorded from extensor digitorum brevis through belly-tendon electrodes. Stimulating and recording electrodes were stainless steel needles placed subcutaneously as described by Hopkins and Gilliatt (1971). After
the muscle records had been made, ascending action potentials were recorded from the lateral popliteal nerve at the neck of the fibula with stimulation of the anterior tibial nerve at the ankle. Details of the technique are described by Hopkins and Gilliatt (1971). The intramuscular temperature in the tibialis anterior muscle was recorded at the beginning and end of each session and varied between 35° and 38°C.

For histological studies, specimens of muscle and nerve were taken shortly after the animals were killed, and were fixed to cards. Flemming’s solution was used to fix nerve for subsequent staining with Kultschitsky’s haematoxylin (Gutmann and Sanders, 1943) but for all other stains the fixative was 10% neutral formalin. Specimens of the nerve were stained with osmium tetroxide in preparation for the examination of single teased fibres (Thomas, 1955), while others were embedded in paraffin and stained by the Holmes silver method, combined with cresyl violet and luxol fast blue (McDonald, 1963). Some specimens of muscle were embedded in gelatine and thick (50–100 µ) sections were cut which were stained by Koelle’s cholinesterase method for end-plates (Gomori, 1952). In order to show the intramuscular nerve fibres in the same sections, additional myelin staining with Sudan black B (Cavanagh, Passingham, and Vogt, 1964) or axon staining with a modified Bielschowsky silver technique was performed before mounting in a water-based mountant.

RESULTS

Electrophysiological observations before and after nerve section were made on six nerves in four animals. In each case recordings were made daily until conduction failed completely.

Tracings of muscle action potentials from one animal are shown in Fig. 1. It can be seen that the amplitude of the muscle action potentials had fallen by only 21% after two days but thereafter the change was more rapid, the amplitude being reduced to approximately 3% of the control.

**TABLE 1**

<table>
<thead>
<tr>
<th></th>
<th>Pre-op</th>
<th>Days after nerve section</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Number of nerves</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>Mean response amplitude with knee stimulation (mV)</td>
<td>15.5</td>
<td>14.2</td>
</tr>
<tr>
<td>Mean response amplitude with ankle stimulation (mV)</td>
<td>15.4</td>
<td>13.9</td>
</tr>
<tr>
<td>Ratio knee:ankle</td>
<td>1.01</td>
<td>1.02</td>
</tr>
</tbody>
</table>

**FIG. 1.** Nerve B2R. Muscle action potentials evoked by nerve stimulation on successive days after nerve section in the thigh. The position of stimulating and recording electrodes is shown below.
value after four days. No muscle response to nerve stimulation was obtained after this time. Results for all six nerves are shown in Fig. 2 from which it can be seen that a muscle response was obtainable after five days in one case but not in the others. No muscle response was obtained after six days in any animal.

The values for muscle action potential amplitude shown in Fig. 2 are those obtained by nerve stimulation at the ankle. In Table 1 a comparison is made with the amplitude of the muscle response obtained by stimulation at the knee. It can be seen that after nerve section the reduction in muscle action potential amplitude was similar for knee and ankle stimulation. From this it may be assumed that the failure of conduction was
Distal latency occurring at a point distal to the stimulating electrode at the ankle.

The changes in maximal motor conduction velocity and distal latency after nerve section are shown in Fig. 3. It can be seen that velocity and distal latency remained unchanged until response amplitude was greatly reduced. With values for response amplitude of 2 mV or less, maximal velocity was sometimes reduced and distal latency was increased in every case, the highest value (3.8 msec) being seen in nerve 4L, four days after section. On this occasion, response amplitude was only 0.1 mV and conduction velocity was 44 m/sec. Initial values for response amplitude, conduction velocity, and distal latency in this nerve were 18 mV, 58 m/sec, and 1.1 msec respectively.

The effects of nerve section on ascending action potentials are illustrated by the tracings shown in Fig. 4, and results for the six nerves are shown graphically in Fig. 2. It can be seen that ascending nerve action potentials (which include antidromic impulses in motor fibres) persisted longer after nerve section than the muscle response amplitude was only 0.1 mV and conduction velocity was 44 m/sec. Initial values for response amplitude, conduction velocity, and distal latency in this nerve were 18 mV, 58 m/sec, and 1.1 msec respectively.

The effects of nerve section on ascending action potentials are illustrated by the tracings shown in Fig. 4, and results for the six nerves are shown graphically in Fig. 2. It can be seen that ascending nerve action potentials (which include antidromic impulses in motor fibres) persisted longer after nerve section than the muscle

**TABLE 2**

<table>
<thead>
<tr>
<th>Days after nerve section</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of observations</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>6</td>
<td>5</td>
<td>4</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>Mean velocity and S.E.</td>
<td>(98)</td>
<td>(101)</td>
<td>(102)</td>
<td>(97)</td>
<td>(97)</td>
<td>(94)</td>
<td>(98)</td>
<td>(98)</td>
</tr>
</tbody>
</table>

(mean: 3.5 m/sec; s.d.: 1.2 m/sec).

Motor velocity

amplitude of muscle response (mV)

FIG. 3. Maximal motor velocity and distal latency on successive days after nerve section (six nerves). Values are related to the amplitude of the muscle response in each case.
degeneration. In some of the animals a low voltage deflection was seen preceding the main potential change. This early component was originally described by Hopkins (1968) and considered to be due to impulses in a group of low threshold, high-velocity, afferent fibres. In the present study, this initial component made latency measurement difficult. Before nerve section its amplitude was only 1–3 μV, and when this was further reduced during Wallerian degeneration, accurate measurement became impossible. However, velocity calculated from the latency measured to the foot of the main deflection of the ascending action potential showed no significant change after nerve section in the six nerves examined (Table 2).

For comparison with the electrophysiological results, nerves were taken for histology from two additional animals three and six days after section. Control nerves were also examined. Three days after section the nerve trunk in the leg showed only minor changes. Longitudinal sections stained by the Holmes silver method and counterstained by luxol fast blue and cresyl violet were normal save for occasional vacuoles in the myelin. Individual teased fibres showed slight irregularity of the contour of the myelin, particularly in the paranodal regions, but the

![Image](https://i.imgur.com/339.png)

**Fig. 4. Nerve B4R. Ascending action potentials recorded from the lateral popliteal nerve at the knee after section of the nerve in the thigh. Twenty-five faint traces superimposed in each record.**

responses to motor nerve stimulation. The significance of this is discussed in a subsequent section. It can also be seen from the tracings in Fig. 4 that there was no change in the latency of the ascending action potentials during Wallerian

<table>
<thead>
<tr>
<th>Species</th>
<th>Nerve</th>
<th>Time to failure (hr)</th>
<th>Authors</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nerve trunk</td>
<td>Rabbit peroneal (lat. popliteal)</td>
<td>71–78  (1950)</td>
<td>Gutmann and Holubár</td>
</tr>
<tr>
<td></td>
<td>Rat peroneal (lat. popliteal)</td>
<td>79–81  (1950)</td>
<td>Gutmann and Holubár</td>
</tr>
<tr>
<td></td>
<td>Guinea-pig peroneal (lat. popliteal)</td>
<td>72–82  (1950)</td>
<td>Gutmann and Holubár</td>
</tr>
<tr>
<td></td>
<td>Cat sciatic</td>
<td>72–101 (1950)</td>
<td>Rosenblueth and Dempsey</td>
</tr>
<tr>
<td></td>
<td>Dog phrenic</td>
<td>96 (1946)</td>
<td>Erlanger and Schloepfle</td>
</tr>
<tr>
<td></td>
<td>Baboon lateral popliteal</td>
<td>120–216 (1946)</td>
<td>Present series</td>
</tr>
<tr>
<td>Nerve muscle</td>
<td>Rabbit peroneal (lat. popliteal)</td>
<td>30–32 (1952)</td>
<td>Gutmann and Holubár</td>
</tr>
<tr>
<td></td>
<td>Rat sciatic</td>
<td>24–36 (1962)</td>
<td>Miledi and Slater</td>
</tr>
<tr>
<td></td>
<td>Guinea-pig sciatic</td>
<td>40–45 (1962)</td>
<td>Kaeser and Lambert</td>
</tr>
<tr>
<td></td>
<td>Cat sciatic</td>
<td>69–79 (1962)</td>
<td>Lissak, Dempsey, and</td>
</tr>
<tr>
<td></td>
<td>Man median, ulnar*</td>
<td>85–128 (1953)</td>
<td>Rosenblueth (1939)</td>
</tr>
<tr>
<td></td>
<td>Man facial</td>
<td>120–192 (1959)</td>
<td>Gilliatt and Taylor</td>
</tr>
<tr>
<td></td>
<td>Baboon lateral popliteal</td>
<td>96–144 (1959)</td>
<td>Present series</td>
</tr>
</tbody>
</table>

* From observation of muscle twitch—no electrical recording.
nodes of Ranvier were not widened (cf. Causey and Palmer, 1952). In contrast to these relatively normal appearances, some of the intramuscular nerve bundles already showed fragmentation of myelin which increased as the end-plate region was approached (Fig. 5). In many cases the terminal portions of the fibres had disappeared, although the proximal parts were still visible. Six days after nerve section, loss of continuity of axons could be seen in the leg as well as in the intramuscular nerve bundles, the changes still being more marked distally. By this time the terminal portions of the intramuscular nerves had disappeared.

**DISCUSSION**

Previous writers have emphasized that failure of neuromuscular transmission precedes loss of conduction in the nerve trunk during Wallerian degeneration (Titeca, 1935; Lissák, Dempsey, and Rosenblueth, 1939; Birks, Katz, and Miledi, 1960). Histological studies have shown early changes in terminal nerve fibres (Gutmann and Holubář, 1952) and in the end-plates themselves (Miledi and Slater, 1970). These observations are supported by our own findings. For example, histological examination three days after section showed advanced changes in the terminal branches of intramuscular nerve fibres, whereas the same fibres in the leg showed only occasional abnormalities. In the nerves examined electrophysiologically, ascending action potentials were present after the muscle response to nerve stimulation had disappeared. This latter finding is unlikely to be due to the longer survival of sensory fibres than motor fibres, since others have shown that conduction failure tends to occur earlier in the former than in the latter (Gutmann and Holubář, 1949).

A slight reduction in conduction velocity during Wallerian degeneration has been described by previous writers (Gutmann and Holubář, 1950; Kaeser and Lambert, 1962). In
the present experiments no change in maximal motor velocity or in distal latency occurred until most of the nerve fibres had ceased to conduct impulses. At this stage distal latency had increased in all nerves and conduction velocity was mildly reduced in two. These results do not necessarily imply a velocity change in individual fibres. They could be explained by the survival of normal fibres with a relatively low velocity or a long intramuscular course. In the case of ascending nerve action potentials, the present results provide no evidence of a velocity change preceding conduction failure.

The time to conduction failure varies considerably in different species. In Table 3 our own results are compared with those of previous workers. From this it appears that the time to final failure of conduction in the lateral popliteal nerve of the baboon is substantially longer than that found in the nerve trunks of lower mammals. Neuromuscular transmission in the baboon also persists for longer than is the case in lower mammals. There is a suggestion that this time might be longer still in man (Gilliat and Taylor, 1959) and further information on this point would be of considerable interest.

This work was supported by a grant from the Medical Research Council.

REFERENCES


Nerve conduction during Wallerian degeneration in the baboon

R. W. Gilliatt and R. J. Hjorth

J Neurol Neurosurg Psychiatry 1972 35: 335-341
doi: 10.1136/jnnp.35.3.335

Updated information and services can be found at:
http://jnnp.bmj.com/content/35/3/335

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/