Biomarkers in dementia: clinical utility and new directions


ABSTRACT

Imaging, cerebrospinal fluid (CSF) and blood-based biomarkers have the potential to improve the accuracy by which specific causes of dementia can be diagnosed in vivo, provide insights into the underlying pathophysiology, and may be used as inclusion criteria and outcome measures for clinical trials. While a number of imaging and CSF biomarkers are currently used for each of these purposes, this is an evolving field, with numerous potential biomarkers in varying stages of research and development. We review the currently available biomarkers for the three most common forms of neurodegenerative dementia, and give an overview of research techniques that may in due course make their way into the clinic.

INTRODUCTION

A biomarker is a characteristic that can be objectively measured and evaluated as an indicator of normal biological or pathogenic processes or pharmacological responses to a therapeutic intervention.1 An ideal biomarker is reproducible, stable over time, widely available and reflects directly the relevant disease process.2 For the dementias, biomarkers may be used to distinguish different aspects of the underlying pathology; detect presymptomatic pathological changes; predict decline or conversion between clinical disease states; and/or monitor disease progression and response to treatment. In this review we provide an overview of currently available biomarkers, with a particular focus on the three commonest neurodegenerative dementias: Alzheimer’s disease (AD), frontotemporal dementia (FTD) and dementia with Lewy bodies (DLB).

BIOMARKERS OF DEMENTIA CURRENTLY IN CLINICAL USE

Biomarkers in dementias can be divided into imaging modalities, which are already widely used in clinical and research settings; and cerebrospinal fluid (CSF) measures whose clinical use varies widely between countries and centres. Currently, no blood-based or urine-based biomarkers are available for routine clinical use.

Imaging

Structural brain imaging

Structural brain imaging (CT or MRI) is recommended in all patients being investigated for dementia, according to UK, European and US guidelines.3 Moving away from excluding ‘surgical’ causes (eg, mass lesions) of cognitive impairment, structural imaging (and MRI in particular) can usefully assess vascular damage, white matter signal changes with a wide range of causes,4 and spongiform and gliotic changes as seen in prion disease. The pattern of regional brain loss (atrophy) reflecting neuronal loss has positive predictive value for different dementias5 and is incorporated into diagnostic criteria for several dementia syndromes (discussed below). Atrophy can be assessed using simple visual rating scales or more complex quantitative manual or automated techniques. Serial imaging—particularly with MRI, which provides superior grey/white contrast without radiation exposure—is widely used as a safety and outcome measure for clinical trials, with rates of atrophy considered surrogate markers of neurodegeneration.

Fluid biomarkers

Cerebrospinal fluid

In the context of dementia, CSF examination has traditionally been used to exclude infection, malignancy and neuroinflammation, as reflected by guidelines recommending CSF examination in the context of differential diagnosis.
individuals with cognitive impairment under the age of 55, individuals with rapid disease course, ‘unusual’ dementia syndromes or those who are immunosuppressed. In degenerative forms of dementia, the cell count is not usually raised, and there is no evidence for CNS-specific immune responses. Where abnormalities are present, these should prompt consideration of unusual forms of dementia, including infectious and inflammatory conditions. In cases of rapidly progressive dementia, the presence of a positive 14-3-3 protein, elevated S100B, elevated total-tau to phospho-tau ratio, and recently the use of real-time quaking-induced conversion (RT-QUIC) technology have positive predictive value for prion disease. CSF analysis using a variety of immunochemical techniques allows a range of neuronal-specific or neuronal-enriched proteins to be measured (see below). The use of neuronal-enriched CSF markers β-amyloid and tau in the routine evaluation of patients with dementia varies considerably between countries and between clinicians; these markers have been incorporated into new AD diagnostic criteria (see below, and table 2), and are increasingly included as inclusion/outcome measures for clinical trials. There is considerable variability in the methods used to collect and analyse CSF, leading to initiatives to harmonise preanalytical handling and standardise laboratory practices (see table 1). Developing normal reference ranges and reliable cut points for CSF, leading to initiatives to harmonise preanalytical factors

### BIOMARKERS OF AD

AD is the commonest cause of dementia particularly in older individuals, and is characterised neuropathologically by amyloid plaques and tau containing neurofibrillary tangles. Other pathological changes include the presence of activated microglia around amyloid plaques, and amyloid angiopathy and microhaemorrhages in some individuals with AD. Most cases of AD are sporadic, with autosomal dominant mutations in either the presenilin 1 (PSEN1), presenilin 2 (PSEN2) or amyloid precursor protein (APP) genes accounting for <1% of cases. In sporadic AD more than 20 genetic risk factors have been identified, implicating cholesterol transport, innate immunity and endosomal vesicle recycling in pathogenesis. Progressive impairment of episodic memory is the commonest clinical presentation of AD; rarer focal presentations include posterior cortical atrophy, logopenic aphasia (LPA) and a dysexecutive presentation. In clinical practice there is often significant overlap between these syndromes and a merging of symptoms as the disease progresses. The original clinical criteria for AD did not acknowledge this phenotypic diversity, and by requiring that an individual have dementia and multidomain cognitive impairment, precluded the early symptomatic and presymptomatic stages of AD. Newer criteria place an emphasis on using biomarkers to provide an earlier and more specific diagnosis (table 2). For research purposes only, this may extend to the prodromal phase of the disease, which may begin a decade or more before cognitive impairment occurs.

### Table 1 Optimal practice for CSF collection and processing

<table>
<thead>
<tr>
<th>Confounding variable</th>
<th>Ideal situation</th>
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<tbody>
<tr>
<td><strong>Preamanlytical factors</strong></td>
<td></td>
</tr>
<tr>
<td>Time of collection</td>
<td>8:00–12 noon to avoid potential for diurnal variation in CSF biomarkers</td>
</tr>
<tr>
<td>LP needle</td>
<td>Needle gauge or design not known to influence measured biomarker concentration but gauge is related to risk of post-LP headache. Smallest size practical to use in diagnostic LP is 22G. Atraumatic needles are associated with reduced post-LP headache but increased failure rate</td>
</tr>
<tr>
<td>Use of lumbar catheters/manometers</td>
<td>Aβ1-42 may adhere—to be avoided, if possible</td>
</tr>
<tr>
<td>Collection vessel</td>
<td>Polypropylene tube recommended. Aβ1-42 and other proteins adhere to polystyrene and glass significantly reducing measured concentrations. Tube brand may also influence measured biomarker concentrations</td>
</tr>
<tr>
<td>Fasting</td>
<td>Not required</td>
</tr>
<tr>
<td><strong>Blood contamination/blood–brain barrier dysfunction</strong></td>
<td>Blood contamination of up to 5000 erythrocytes/μL cells does not alter measured biomarker concentration, but blood–brain barrier dysfunction equivalent to CSF/serum albumin ratio &gt;11 results in reduced measured Aβ1-42 concentration and should be interpreted with care</td>
</tr>
<tr>
<td><strong>Optimal volume</strong></td>
<td>In addition to CSF collected for routine clinical examination (eg, cell count, oligoclonal bands, cytology, etc) 15 mL can safely be collected without increased risk of post LP headache</td>
</tr>
<tr>
<td><strong>Centrifugation</strong></td>
<td>Within 30 min of LP at 3000 rpm for 10 min to remove cells and other debris</td>
</tr>
<tr>
<td><strong>Aliquot storage volume</strong></td>
<td>Samples that are frozen prior to analysis should be stored in aliquots of a standardised volume which fits the tube size well. Specifically, one should strive for as high volume to surface ratio as possible (well-filled tubes). Volume to surface ratio and number of tube transfers influence measured Aβ1-42 concentration, probably due to protein adsorption</td>
</tr>
<tr>
<td><strong>Freeze thawing</strong></td>
<td>One or less freeze thaw cycles is recommended. Measured Aβ1-42 concentration drops by 20% after 3 freeze thaw cycles. Aβ1-42 and t concentrations are stable at temperatures of −80°C</td>
</tr>
<tr>
<td><strong>Choice of immunoassay/platform</strong></td>
<td>Consistency required; variability in commercially available ELISA-based assays, calibration peptides and platforms mean interlaboratory and interassay consistency is poor</td>
</tr>
</tbody>
</table>

CSF, cerebrospinal fluid; LB, lumbar puncture.
**Table 2  Biomarkers currently used in diagnostic criteria**

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>AD</strong></td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Biomarkers of amyloid pathology: Low CSF Aβ1-42 on CSF examination</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Positive amyloid PET scan</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Biomarkers of neuronal injury: Elevated CSF tau and phospho-tau;</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Hypometabolism on FDG-PET; Disproportionate atrophy of medial, basal and lateral temporal lobe, and medial parietal cortex on structural MRI</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>McKhann criteria^16 require evidence of amyloid pathology and neuronal injury to support a diagnosis of highly probable AD (biomarker evidence only recommended in individuals who do not meet the core clinical criteria for probable AD dementia). Dubois criteria (IWG2) require specific clinical features of AD (typical or atypical) plus evidence of in vivo AD pathology. Evidence of in vivo AD pathology: low CSF Aβ1-42 together with increased total-tau or phospho-tau or positive amyloid PET or proven mutation in PSEN1, PSEN2, or APP or other proven genes (including Down’s syndrome trisomy 21).</td>
<td></td>
</tr>
<tr>
<td><strong>Frontotemporal dementia</strong></td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>bvFTD</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Frontal and/or anterior temporal lobe atrophy on MRI or CT</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Progressive non-fluent aphasian</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Predominant left frontoinsular atrophy on MRI</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Predominant left frontoinsular hypoperfusion or hypometabolism on PET or SPECT</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Semantic dementia</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Predominant anterior temporal lobe atrophy</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Predominant anterior temporal hypoperfusion or hypometabolism on PET or SPECT</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td><strong>Dementia with Lewy bodies</strong></td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Relative preservation of medial temporal lobe structures on CT/MRI</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Generalised low uptake on SPECT/PET perfusion scan with reduced occipital activity</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Abnormal (low uptake) MIBG myocardial scintigraphy</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
<tr>
<td>Abnormal uptake on PET/SPECT (eg, 123 I-FP C15-DaTSCAN)</td>
<td>Either evidence of low CSF Aβ1-42 or positive amyloid PET scan required for diagnosis of amyloid brain deposition^1^</td>
</tr>
</tbody>
</table>

AD, Alzheimer’s disease; APP, amyloid precursor protein; bvFTD, behavioural variant frontotemporal dementia; CSF, cerebrospinal fluid; FDG, 18-F-fluorodeoxyglucose; MIBG, metaiodobenzylguanidine; p-tau, tau phosphorylated at 181; PET, positron emission tomography; SPECT, single photon emission tomography; t-tau, total tau.

**Currently used biomarkers and their utility**

**Structural imaging**

In AD, the typical imaging appearance is of global brain atrophy with early disproportionate symmetrical involvement of medial temporal lobe structures including the hippocampi. The presence of symmetrical medial temporal atrophy can differentiate AD from ageing with a sensitivity and specificity of around 80–85%; and in a single centre pathologically proven study, established AD from DLB and vascular cognitive impairment with a sensitivity and specificity of 91% and 91%, respectively. The presence of medial temporal lobe atrophy per se is unable to differentiate AD from FTLD in most instances, although patterns of (medial and lateral) temporal lobe atrophy may be very helpful in identifying specific FTLD subgroups (see below). Medial temporal lobe atrophy can predict which individuals will develop clinical AD from a mild cognitive impairment (MCI) state with a sensitivity and specificity of 73% and 81%, respectively. Progressive atrophy of the parietal/occipital lobes is supportive of AD and in particular in distinguishing AD from FTLD; incorporating visual ratings of posterior atrophy can improve the distinction of AD from other causes of dementia. Rates of whole brain and hippocampal atrophy, calculated from serial volumetric MRI are sensitive markers of progression of neurodegeneration and are increasingly used as outcome measures in trials of potentially disease modifying therapies in AD.

**Functional imaging**

The typical AD pattern on FDG-PET or HMPAO-SPECT imaging is bilateral hypometabolism and hypoperfusion in the temporal and parietal cortices; the sensitivity and specificity in diagnosing AD versus other neurodegenerative diseases has been reported as 79% and 88%, respectively. These changes may predate brain atrophy or cognitive symptoms, and correlate with disease severity. Amyloid PET is a sensitive and specific means of imaging brain amyloid in individuals in vivo, and has been shown to correlate closely with autopsy measures of fibrillar amyloid load, and has considerable potential value in ruling in/out AD pathology as the cause of cognitive decline in a patient with cognitive impairment. Several amyloid PET agents are now licensed for clinical use, and amyloid imaging is now included in criteria for the diagnosis of AD. Importantly however as up to a third of elderly non-demented individuals may have a positive amyloid scan, the significance of which is as yet unclear, the clinical utility of amyloid PET imaging in older individuals is still to be determined and in particular it is not recommended in cognitively healthy individuals outside of research studies. Additionally, the presence of amyloid pathology does not always equate to a diagnosis of AD, as for instance a proportion of patients with DLB will also have amyloid deposition.

Current guidelines (eg, by the UK Royal College of Radiologists and Physicians) advocate the use of amyloid PET in highly selected individuals with cognitive impairment after evaluation by a dementia expert where the presence or absence of amyloid pathology is expected to increase diagnostic certainty and influence management. At the time of writing, availability and cost of amyloid PET imaging has limited its use in clinical practice.

**Cerebrospinal fluid**

**β-Amyloid**

CSF levels of Aβ1-42, thought to be one of the key pathological forms of Aβ in brain tissue, are reduced in AD, with the degree...
of reduction correlating with brain amyloid plaque load. Reduction of CSF Aβ1-42 occurs years before symptom onset, and has good positive predictive value for conversion from MCI to clinical AD; accordingly CSF Aβ1-42 is now included in new diagnostic criteria for MCI due to AD. In clinical practice, a normal CSF Aβ1-42 in a demented individual should prompt re-evaluation of a diagnosis of AD. Conversely however, a low CSF Aβ1-42, does not always reflect brain amyloid deposition, being seen in other conditions including multiple sclerosis, and more commonly due to CSF being collected, stored or processed incorrectly. Other forms of β-amyloid, notably Aβ1-40 can be measured in CSF and may better reflect both total brain Aβ burden than Aβ1-42. While some studies have suggested that addition of CSF Aβ1-40 may improve differential diagnosis in certain circumstances this has not yet entered routine clinical practice.

**Tau and Phospho-tau 181**

CSF levels of t-tau and tau phosphorylated at 181 (p-tau), are both increased in AD. Like Aβ1-42, t-tau and p-tau are usually measured using ELISA-based assays. There are also multiplexed assays for the three analytes. T-tau is increased after stroke, in inflammatory conditions and in other neurodegenerative diseases—most notably in Creutzfeldt-Jakob disease where levels are often orders of magnitude higher than in AD; p-tau elevation is thought to have high specificity for AD. Stability and reproducibility of t-tau and p-tau levels is good, and levels remain stable over periods of up to 6 months, suggesting that these biomarkers may be capable of detecting small biochemical changes induced by treatment. There are at present no data to support the use of tau/p-tau assays in peripheral blood, due to tau concentrations being below the lower limit of detection for most assays in the blood.

**Combining CSF biomarkers**

In 2006 a longitudinal study of 137 individuals followed for 4–6 years demonstrated that the combination of low CSF Aβ1-42, and elevated t-tau and p-tau could distinguish individuals with MCI/incipient AD from those without with 95% sensitivity and 87% specificity. These findings have since been replicated in several other studies. On a research basis, the combination of low Aβ42, elevated tau and p-tau has also been used to predict future cognitive decline in healthy older individuals. In clinical practice, the combination of low CSF Aβ1-42 and elevated tau (or p-tau) to Aβ1-42 ratio is often used to support the diagnosis of AD, with one recent study suggesting that tau:Aβ42 ratio is the most robust single biomarker combination.

In practice, combining several of these different biomarkers, each of which provides different insights into the underlying disease process, may increase diagnostic certainty (see Case Study figure 1).

**Emerging and future biomarkers**

**Advanced MRI**

Current hypotheses predict that amyloid deposition, tau mediated neuronal dysfunction, neuroinflammation and synaptic loss precede the development of structural brain changes, that is, atrophy by several years, which in turn predicts cognitive impairment. There is therefore considerable interest in determining imaging biomarkers to detect and quantify AD-related network disruption after the emergence of molecular pathology but before irreversible neuronal loss. Techniques including diffusion tensor imaging (DTI) and rsfMRI probing white matter tracts and functional integrity of neuronal networks, respectively, show promising results in group studies, but are only just being applied at the individual level. Using rsfMRI, presymptomatic individuals at increased risk of developing AD have shown altered resting connectivity in a distributed temporoparietofrontal network (the so-called default mode network) and altered task-related...
activations involving components of this network that may precede structural brain damage by up to many years. This suggests a potential future application in trial settings where rsfMRI might be used to assess the early impact of candidate interventions on brain function. Arterial spin labelling MRI has the potential to demonstrate cerebral blood flow pattern, which may provide a non-invasive means of obtaining similar information to FDG-PET.

Tau PET imaging

Very recently, a number of tau PET tracers have been developed. At the time of writing, tau imaging has been performed only in small number of individuals with AD, and relatively little data is available. Once mature, tau PET imaging may prove valuable both for differential diagnosis, for prognostication, and as an outcome measure for clinical trials.

Fluid biomarkers

Biomarkers of amyloid processing

The dominant hypothesis for AD pathogenesis focuses on the production of toxic, amyloidogenic forms of Aβ. The APP can be cleaved either by β-secretase and γ-secretase cleavage, producing amyloidogenic forms of Aβ, including Aβ1-42; or via other pathways to non-amyloidogenic forms. The β-site APP-cleaving enzyme 1 (BACE1), the major β-secretase in the brain, can be measured in the CSF. While some studies have determined higher levels of CSF BACE1 in AD, results have generally been inconsistent. Some studies have determined higher levels of CSF BACE1 in AD, but as yet their use in clinical practice is limited due to the difficulties of measuring their low concentration in CSF.

Biomarkers of neuroinflammation

A number of pathways related to neuroinflammation and more specifically microglial activation have been implicated in AD pathogenesis. The astrocytic and oligodendrocytic protein S100B is elevated in a range of conditions—most notably prion diseases, but possibly also in mild/moderate AD where it may correlate with rate of brain atrophy. The glycoprotein YKL-40 produced by astrocytes or activated microglia, plays a number of poorly defined roles in CNS inflammation, and is increased in CSF and serum in AD and FTD. F2-Isoprostanes, markers of membrane lipid peroxidation and inflammation, have been shown to predict conversion from MCI to AD. Other potential measures are shown in supplementary table.

Biomarkers for subcortical axonal degeneration and synaptic dysfunction

Elevated CSF neurofilament light protein (NFL) levels indicate involvement of predominantly large-calibre myelinated axons, and are elevated in a range of disorders including stroke, inflammatory disorders, vascular cognitive impairment and FTD (see below), but typically not ‘pure’ AD. A range of putative markers of synaptic function, and notably neurogranin show promise, but as yet their use in clinical practice is limited due to the difficulties of measuring their low concentration in CSF.

Blood-based and urine-based biomarkers

Despite much research, perhaps for the reasons discussed above, there are currently no established blood-based biomarkers for AD. Recent approaches using techniques where several biomarkers are analysed simultaneously have identified promising biomarkers, but the results have unfortunately been hard to replicate. Over 20 such studies, using different clinical cohorts and different methodology, now exist, but the majority of potential biomarkers, including a set of 10 lipids published in a recent study have only been detected in single studies. A small number of proteins have been identified as promising potential biomarkers across more than one study, these will however need to be tested in larger cohorts with prospective follow-up to determine their clinical utility.

BIOMARKERS OF FTD

The term FTD refers to a group of neurodegenerative disorders characterised by atrophy of the frontal and temporal lobes. Prevalence studies suggest that FTD is the second commonest cause of young onset dementia after AD. The two main clinical syndromes of FTD, are behavioural variant FTD (bvFTD), where there is deterioration in social function and personality; and primary progressive aphasia (PPA) where there is an insidious decline in language skills. PPA is further divided into several subtypes including semantic dementia (SD), progressive non-fluent aphasia (PNFA), logopenic aphasia (LPA) (typically an AD variant) and progressive apraxia of speech, based on the pattern of language breakdown. FTD and motor neuron disease show considerable overlap. Approximately a third to a half of patients with FTD—and particularly those with bvFTD—have a family history with an autosomal dominant pattern of inheritance. The three most common genes associated with FTD are hexanucleotide expansions in the C9orf72 gene (also linked to motor neuron disease phenotypes), mutations in the microtubule associated protein tau (MAPT) gene, and loss of function mutations in the progranulin (GRN) gene; a number of rarer genetic causes are also recognised.

FTD occurring on a sporadic and an autosomal dominant basis is associated with a range of different underlying pathologies, based on the predominant protein accumulation. These include tau (4-repeat—progressive supranuclear palsy (PSP) or corticobasal degeneration (CBD) type, 3-repeat—Pick’s disease type, and mixed 3-repeat and 4-repeat forms), Tau-DNA binding protein (TDP)-43 (types A–D) and fused in sarcoma protein (FUS). In bvFTD any of the pathological variants can be found, with tau and TDP-43 pathologies representing the majority in an approximately 50:50 split. SD and the behavioural/prosopagnostic presentations of SD (associated with more prominent right temporal lobe atrophy) are strongly associated with TDP type C pathology. The pathology of the other language forms is more complex, and includes AD (particularly in LPA), tauopathy (with PSP and CBD pathology commonly seen in patients with apraxia of speech), and TDP-43 proteinopathies. The co-occurrence of FTD and MND is strongly suggestive of underlying TDP-43 (typically type B) pathology.

Currently used biomarkers and their utility

Structural brain imaging

While by no means 100% concordant, pattern of brain atrophy can provide important clues, not only to clinical phenotype but also for the underlying pathology and in some cases genetic basis of disease. Based on clinical phenotype, patients with bvFTD often show atrophy of mesial frontal, orbitofrontal and
Cognitive neurology

anterior insula cortices. As the disease progresses there is involvement of the other frontal neocortical grey matter regions, the striatum, hippocampi, posterior insulae and parietal lobes. Subcortical and insular portions of the dominant hemisphere.62 As the disease progresses this degeneration extends caudally into the posterior temporal lobes and rostrally into the posterior, inferior frontal lobes. The structural imaging findings in PNFA are very heterogeneous and scans are often remarkably similar with the close clinicopathological correlation with SD.65

When classified on the basis of pathology, 3-repeat tau Pick’s disease is often associated with atrophy involving the prefrontal cortex with severe dorsolateral frontal atrophy, which can be markedly asymmetric. Patients with PSP and CBD can often present with an FTD-like syndrome, in which case certain features on structural imaging may be particularly helpful: patients with PSP classically have atrophy of the rostral midbrain, leading to the so-called Hummingbird sign on imaging. This is however not always present in dementia dominant cases. In PSP and CBD atrophy may affect the posterior frontal lobes, while in CBD the atrophy will often involve the parietal lobes and can be very asymmetric. However, in pathologically ascertained series, the atrophy in ‘cognitive’ CBD is often surprisingly nonspecific and symmetrical.

Certain patterns of atrophy have been associated, although by no means invariably, with the different TDP-43 subtypes. Thus TDP-A is often associated with asymmetric atrophy involving the frontal, temporal and parietal lobes; TDP-B is associated with frontal atrophy, in keeping with the association of this pathology with the clinical syndrome of FTD/MND; and TDP-C with asymmetric anterior temporal lobe atrophy, consistent with the close clinicopathological correlation with SD. FUS pathology is associated with frontal atrophy often with striking atrophy of the caudate nuclei.

From a genetic perspective, individuals with tau (MAPT) mutations, often have very symmetrical anterior and inferior medial temporal lobe atrophy, with involvement of the orbitofrontal cortices. In GRN mutations atrophy is typically highly asymmetric involving the temporal, inferior frontal and parietal lobes. No clear and consistent pattern of atrophy has yet emerged in C9orf72 mutation cases: atrophy is often symmetrical, but not always; and while often relatively mild, can on occasions be very widespread. It can involve the frontal and temporal lobes, and thalami, but may extend to the occipital lobe and cerebellum, where TDP-43 or p62-positive inclusions are frequently found at histology.

Functional imaging

FDG-PET typically shows frontal and temporal lobe hypometabolism in FTD. Demonstrating hypometabolism may be particularly valuable in cases with behavioural change with normal structural imaging, where a non-degenerative FTD-mimic is considered. Where the differential diagnosis is between AD and FTD, amyloid PET imaging may be very useful in ruling in/out the presence of amyloid pathology; this may prove particularly valuable in the progressive aphasias given their considerable clinical and pathological heterogeneity and also in corticobasal syndrome where it is difficult to distinguish CBD from AD as the underlying cause.

Cerebrospinal fluid

In the absence of specific CSF biomarkers for TDP-43, or to distinguish the different tauopathies, currently the most important clinical utility of CSF biomarkers in FTD is to distinguish underlying AD from other FTD pathologies. In particular, if CSF is appropriately handled and measured, reduction of Aβ1-42 level would not be expected in cases with tau or TDP-43 proteinopathies. Several studies have shown that CSF t-tau levels are lower in FTD than those seen in AD, but higher than that seen in controls, with t-tau levels correlating with neuropsychological, neuroimaging and prognosis in patients with FTD; however in many cases with FTD, CSF t-tau levels can be normal. As discussed above, p-tau elevation is typically seen in AD rather than other neurodegenerative diseases, with one study of FTD finding that a reduced p-tau to t-tau ratio predicts TDP-43 pathology in FTD.

Emerging and future biomarkers

Imaging

As with AD, DTI and rsfMRI have considerable potential to detect presymptomatic and disease-specific network breakdown in the various forms of FTD. Specific patterns of DTI breakdown have been associated with the clinical presentations in FTD and on a group level DTI may be helpful in discriminating between tau and TDP-43 proteinopathies, the latter showing greater white matter damage. There is considerable evidence again at least on a group level that breakdown in both functional connectivity in FTD involves different functional networks compared with AD, for example, targeting the so-called salience (environmentally directed) as opposed to default mode (internalised thought) network. Within the FTD spectrum, different diseases may have characteristic, molecularly determined network signatures or ‘neoxopathies’. It is however not clear if and how these techniques can be applied on an individual basis.

As with AD, ASL may provide a non-invasive alternative to FDG-PET, providing an MRI-based measure of cerebral perfusion. It is not yet clear to what extent available tau PET ligands bind different subtypes of tau pathology or how reliably they can distinguish tau, AD and TDP-43 pathologies, or whether this technique will prove a useful means of reliably distinguishing the various FTD pathologies in vivo. To date, no specific TDP-43-based ligands are available.

Fluid biomarkers

Tar-DNA binding protein-43

Increased TDP-43 levels have been found in CSF in FTD and MND cases. Kasai et al found increased levels in MND particularly early in disease progression, suggesting that CSF TDP-43 may be an early marker of TDP-43 proteinopathies. Patients with C9orf72 expanded repeats or GRN mutations, where TDP-43 pathology can reliably be predicted in vivo, have been shown to have increased plasma and CSF levels of phosphorylated TDP-43 compared with other patients with FTD and normal controls; this same study found plasma levels of total TDP-43 to be decreased in mutation carriers, possibly due to alterations in the ratio of phosphorylated to total TDP-43 in favour of the former.

**Progranulin**

Serum progranulin levels show considerable promise as a biomarker of underlying progranulin mutations. Null mutations have been associated with a fourfold reduction in plasma progranulin levels compared to controls, with missense mutations resulting in a smaller reduction. Studies of large, mixed, FTD populations are required to determine the sensitivity and specificity of serum progranulin as a predictor of mutations and of the underlying pathology.

**Neurofilament**

Two recent studies have found that elevated CSF NFL levels correlate with FTD disease severity. The highest levels were in tau negative cases and SD, where TDP-43 is the predominant pathology. Different levels, and perhaps forms of neurofilament may therefore find utility in differentiating the underlying pathology in cases of FTD, in distinguishing AD from non-AD pathology, and for tracking disease in patients with confirmed disease; however, as NFL is elevated in several other conditions—and notably vascular disease—its utility in isolated unselected cases is less clear.

**Biomarkers of neuroinflammation**

As with AD, there is considerable interest in inflammation in FTD. A recent study showed increased serum TNF-α in two different conditions strongly associated with TDP-43 pathology, that is, SD and GRN mutation carriers. A number of inflammatory biomarkers have been reported to be elevated in FTD. One study has suggested that IL-23 may be specific for FTD associated with tau, while IL-17 may be specific for FTD associated with TDP-43 pathology; this however requires further confirmation.

**BIOMARKERS OF DLB**

DLB, the second most common cause of neurodegenerative dementia after AD, is characterised by a progressive deterioration in cognition, with core features including fluctuating cognition and variations in alertness; recurrent visual hallucinations; and features of motor parkinsonism. Neuropsychological testing typically reveals deficits in attention, executive function and visuospatial ability. The core pathology of DLB is the presence of Lewy bodies in brainstem, limbic regions or cortex comprising α-synuclein. DLB is widely thought to be on a continuum with Parkinson’s disease dementia, and at autopsy is commonly associated with other pathologies including AD and vascular disease.

**Currently used biomarkers and their utility**

**Structural brain imaging**

Typically, DLB is associated with preserved volume of the medial temporal lobes relative to that seen in AD. The extent to which hippocampal sparing as a marker of DLB as opposed to AD can be applied in individual cases is limited, with some studies finding that MRI is not useful in differentiating DLB from AD.

**Functional imaging**

Current clinical diagnostic criteria include PET/SPECT (eg, 123 I-FP CIT—DaTSCAN) measures of dopaminergic loss in the basal ganglia as a suggestive feature of DLB. DaTSCAN imaging can be used to differentiate DLB from AD and some forms of FTD with reasonable diagnostic accuracy (sensitivity around 80% with 90% specificity), but is less useful in differentiating DLB from atypical parkinsonian conditions which can also show nigrostriatal abnormalities. FDG-PET occipital hypoperfusion may have utility in distinguishing DLB from AD, although changes can be seen in patients with posterior variants of AD, and in CBD. Several studies have found decreased metaiodobenzylguanidine myocardial scintigraphy reflecting impaired sympathetic nervous system function in DLB compared with AD.

**Emerging and future biomarkers**

**Imaging**

Amyloid PET imaging is likely to have limited utility in distinguishing AD from DLB as both may have similar cortical amyloid load, with a recent review of 12 studies finding amyloid PET positivity in 57% of clinically diagnosed DLB cases. However, it may have a role in differentiating DLB from other parkinsonian conditions. There are as yet no α-synuclein PET ligands available.

**CSF**

Levels of CSF tau are very variable in DLB, typically being lower than in AD although in rapidly progressive cases they can be very elevated. Studies examining Aβ1-42 levels have found similar levels between DLB and AD. Some studies have reported reduced levels of CSF α-synuclein in DLB, while others have not. An oxidised form of Aβ1-40 has been proposed as a potential DLB marker. However, this oxidation may appear in CSF postsampling and the results need replication before any conclusions can be drawn on the diagnostic usefulness of this Aβ isoform.

**CONCLUSION**

In the correct clinical context, imaging and CSF biomarkers can provide in vivo evidence for the various pathological processes underpinning the different causes of dementia, which in turn be used to improve diagnostic sensitivity and specificity, as inclusion and exclusion criteria and outcome measures in clinical trials, and to provide insights into pathogenesis. Biomarkers are now being incorporated into new diagnostic criteria (see table 2), and are being used—at least on a research basis—to allow for presymptomatic or paucity-symptomatic diagnosis. While the specific biomarkers vary across different dementias, there are common themes including their potential ability to allow presymptomatic diagnosis, track disease progression by the sequence of changes in biomarkers, and reflect underlying pathology. Many of the biomarkers have shown utility on a group level, but how they can be applied on an individual level and in clinical practice requires further investigation. This should not however diminish their importance especially in the setting of clinical trials, for patient stratification for entry and as outcome measures. No one biomarker is diagnostic of any one condition in its own right, each has limitations, and interpretation should always be done in the appropriate clinical context. Future studies will benefit from ever more sensitive and automated techniques, which will hopefully make blood-based and/or urine-based biomarkers a reality. Critical to the development and validation of any biomarker is standardisation of sample/scan collection, analysis and interpretation; and ultimately post-mortem confirmation of diagnosis.

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UK.


Biomarkers in dementia: clinical utility and new directions

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