Cherry-red spot myoclonus syndrome and α-neuraminidase deficiency: neurophysiological, pharmacological and biochemical study in an adult

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SUMMARY A 22 year old patient with non-familial progressive myoclonus, macular cherry-red spot, moderate cerebellar syndrome and normal intelligence is described. The myoclonus began at the age of 18 years. Focal myoclonus could easily be elicited by voluntary and passive movements, and by touch and electrical stimulation of median nerve. Somatosensory evoked potentials showed a high voltage early component. Jerk-locked averaging of the EEG preceding action myoclonus detected an otherwise hidden, time-related, EEG spike. The myoclonus responded partially but clearly to L-5 hydroxytryptophan plus carbidopa treatment. Biochemical study showed an α-neuraminidase deficiency in cultured fibroblasts: the decrease in this enzyme activity was compared to that found in a patient affected by mucolipidosis III.

A slowly progressing neurological syndrome that combines action sensitive and stimulus-sensitive myoclonus with cherry-red spots at the macula in patients with α-neuraminidase deficiency has been described recently.1-3 In these patients, there were no dementia or bone deformities, and generalised seizures that occurred in two patients were easily controlled by antiepileptic therapy. According to Rapin et al this peculiar syndrome should be kept distinct from other more malignant storage disorders and progressive myoclonic epilepsies and put in a new class of diseases characterised by accumulation of sialilated glycopeptides and glycolipids in tissues, owing to a deficiency of neuraminidase. Lowden and O’Brien4 have defined as Sialidosis type I the Cherry-red Spot Myoclonus Syndrome (CRS-MS) with normal intelligence and α-neuraminidase deficiency.

The present report deals with electrophysiological, pharmacological and biochemical findings in a case of CRS-MS observed recently.

Case report

The patient was an Italian male aged 21 years. There was nothing significant in the family history. The patient was cyanotic at birth but his psycho-motor development had been normal. At age 13 years he had an episode of acute polyarthritis diagnosed as rheumatic fever. At age 18 years the patient began to complain of walking clumsily and a few months later he had a short tonic-clonic generalised seizure. Neurological examination performed at that time showed mild dysmetria and brisk tendon reflexes.

In the following months his difficulties in standing and walking rapidly became worse with the appearance of muscular jerks that affected all four limbs and were exacerbated by voluntary movement. A progressive irregular course eventually led to a condition in which the patient was no longer able to care for himself. When he was admitted to hospital in December 1978 (at the age of 21 years) neurological examination showed diffuse hypotonia, brisk deep reflexes and bilateral Babinski signs. A mild ataxia was evident on finger-nose and heel-knee tests, although evaluation was made difficult by the superimposed myoclonic jerks. The myoclonic jerks were absent at rest but precipitated by any attempt to change posture or even by a simple request to move. The focal character of the myoclonus was evident only at the beginning of the voluntary movement owing to its marked tendency to spread rapidly and end up in repeated massive bilateral...
Cherry-red spot myoclonus syndrome

jerks. In addition to the action and intention myoclonus, there was also reflex myoclonus that could be triggered by passive movements and tactile stimulation. Examination of the ocular fundus showed a normal optic disk and a cherry-red spot in the macular region, more striking in the right eye (fig 1). The results of audiometric examination were normal. The patient was still able to communicate satisfactorily with other people, talking very slowly and using limited mimic movements. He showed no sign of mental impairment.

The CT scan was normal. Blood examination (red and white cell count, erythrocyte sedimentation rate, electrolytes, nitrogen, glucose and bilirubin levels, SGOT, SGPT, alkaline phosphatase, serum protein electrophoresis, serum immunodiffusion) was normal. In the cerebrospinal fluid cell count, total protein, glucose, chloride, and protein electrophoresis and immunodiffusion were normal; 5-hydroxyindoleacetic acid level was 35.57 ng/ml (controls mean value 31.37 ± 4.43). Blood and urinary amino acids were normal. Assays of arylsulphatase A and β-galactosidase in the leucocyte also were normal.

Electrophysiological findings

The EEG and EMG were recorded from surface electrodes on a 12 channel OTE Biomedica polygraph. Scalp electrodes were placed according to the 10–20 international system. In addition selected recording samples were stored on FM magnetic tape for averaging of EEG and EMG activity, time-locked to stimuli and to myoclonic jerks, for detection of non-obvious EEG correlates of myoclonus, the EEG activity preceding the myoclonic discharge was analysed by jerk-locked averaging employing a 100 μs analogue delay-line circuit. An electronic window (Vescovini 477-25–28) was employed to trigger the program at the onset of myoclonic potentials. One hundred and twenty EEG samples of 250 ms each were usually averaged.

Somatosensory evoked potentials (SEPs) and reflex myoclonus were elicited by median nerve stimulation by paired surface electrodes placed 3 cm apart on the wrist. Square wave pulses of 1 ms duration were delivered at rate of 0.5–1/s, the intensity was adjusted to induce the least detectable motor response of the opponens muscle. The SEP values computed for 200 bipolar EEG samples were recorded from central and parietal (C3–P3) regions; the analysis time was 250 ms. Evoked EMG activity (that is reflex myoclonus) was displayed on the oscilloscope and fed into the computer. Sixty responses were usually averaged (analysis time 250 ms).

In the resting condition, the electrophysiological recording did not show anything of particular interest. Voluntary and passive movements and tactile stimulation by light touch or tap precipitated vigorous focal myoclonic jerks consisting of repetitive biphasic or polyphasic EMG potential lasting 15–30 ms, occurring five to 15 times per second, which spread rapidly to all the muscles examined eventually leading to a recurrent massive myoclonus (fig 2). Both agonist and antagonist muscles usually were involved. Concomitant EEG bursts of 16–20 Hz activity were almost always detectable in central and parietal regions of both sides, in spite of the superimposed muscle artifacts.

Case 15

![Fundus oculi (right eye): note macular cherry-red spot (courtesy of Dr Delle Grottaglie).](image)

Fig 1

![EEG-EMG recording of action myoclonus evoked by voluntary movement of lower limbs. Bottom: EMG recording of action myoclonus from paired muscles. Note that the biceps could lead or follow the quadriceps according to whether it was precipitated by voluntary movement of upper limbs (upper traces) or of lower limbs (lower traces).](image)

Fig 2
The characteristics of the EEG correlate were demonstrated better by jerk-locked averaging which revealed that each jerk was time-related to an EEG spike lasting about 30 ms and preceding the muscle potential in the contralateral biceps by 17 ms (Fig 3A).

Reflex myoclonus was regularly elicited by electrical stimulation of the median nerve as shown in Fig 3B. The EMG recording in this case showed a more complex wave form than that of action myoclonus and with longer duration (40–60 ms). A quite constant pattern of latency values ranging from 35 ms (for proximal muscles of the ipsilateral upper limb) to 53 ms (for ipsilateral quadriceps) was observed, as demonstrated by the averaging of 60 responses (Fig 4). Note in the same figure that the contralateral SEP evoked by the same median nerve stimulation shows an high voltage (35 uV) early component with 21.5 ms latency preceding by 15 ms the EMG potential recorded in the biceps muscle.

Pharmacological tests and therapy. Pharmacological tests with L-5-hydroxytryptophan (L-5HTP) and clonazepam (CZP) were performed according to a protocol previously set up in our department.5 L-5HTP (Roche: Ro 3–5940/601) was administered by intravenous (iv) infusion of 100 mg in 250 ml of isotonic saline solution (infusion time 60 s) after pretreatment with carbidopa (Merck Sharpe and Dohme) 100 mg given by mouth.

Thirty minutes after the start of the infusion a clearcut reduction of the action myoclonus (Fig 5) and the reflex myoclonus was observed resulting in marked functional improvement. This beneficial effect lasted four hours. No side effects were observed. A similar but more transient effect was induced by a single 1 mg dose of CZP given intravenously.

A chronic trial was then started with increasing oral doses of L-5HTP plus carbidopa, up to 600 and 125 mg respectively. These two drugs were simply added to the preceding therapy, which was maintained unchanged. During 12 months of follow-up, the beneficial effects of L-5HTP on action-sensitive and stimulus-sensitive myoclonus were confirmed. The ensuing improvement in motor performance increased the degree of self-sufficiency of the patient in some daily activities such as dressing, washing and eating. So far there has been no sign of loss of efficacy. Neither clinical nor laboratory side effects have been observed.

Biochemical findings. Fibroblasts from normal controls, a patient with mucolipidosis III (ML III) and the CRS–MS patient were grown in Eagle’s MEM, supplemented with 10% foetal calf serum as previously described.6 Cells were analysed for lysosomal enzymes and a-neuraminidase activities after five to 15 subcultures, seven days after they became confluent.

Lysosomal enzymes in cultured cells and in culture medium were analysed as follows: (a) cells were washed twice with tris-buffered saline (TBS), pH 7.4 and trypsinised with 0.25% trypsin–1% mM EDTA, pH 7.4. The cell pellets were washed twice with TBS. Lysosomal enzymes activities in sonicated cells were then analysed as previously described; (b) fibroblasts were washed twice with Eagle’s MEM without foetal calf serum and then incubated overnight with MEM in order to remove serum proteins entrapped in the cell network. The medium was removed and the fibroblasts incubated for 24 hrs at 37°C in MEM: the

*Fig 3: A: Jerk-locked averaging of the EEG and EMG during action myoclonus showing an EEG spike preceding the muscular potential by 17 msec. B: Reflex myoclonus evoked by stimulation of right median nerve at the wrist.*

*Fig 4: Averaging of reflex myoclonus evoked in different muscles by stimulation of the right median nerve at the wrist is compared with contralateral EEG response evoked by the same stimulation (EMG averaging: 60 samples; EEG averaging: 200 samples, bipolar recording from C3–P3 leads).*
media from three plates for each cell line were pooled: lysosomal enzymes secreted by cultured fibroblasts were precipitated with 80% (NH₄)₂SO₄ centrifuged at 3000 rpm at 4°C in the presence of 0.43 mg/ml albumin as a carrier. The precipitates were suspended in 1 ml of glass redistilled water, dialysed for 24 hr against 500 ml of 0.5 M acetate buffer pH 5.5, in 0.15 M NaCl, with two changes of dialysis solution, and then analysed for lysosomal enzymes.

Neuraminidase activity in sonicated fibroblasts was determined under optimal conditions with α-N-acetyl-neuraminylactose (α2-3) as substrate as recently described. Table 1 shows the activities of α-neuraminidase in the cells of controls, of the ML III patient and of the CRS-MS patient: α-neuraminidase was markedly reduced, to about 5% of the mean control values in the cells of the two patients.

Table 2 shows the activities of five lysosomal hydrolases in cultured fibroblasts and in the culture media from the controls and the two patients. A marked reduction of the activity of the five hydrolases was observed in the cells of the ML III patient, with corresponding increases in enzyme activity in his culture medium, except for that of β-galactosidase; this latter enzyme and the other four hydrolases measured in the cells and in the culture medium of CRS-MS patient showed activities comparable to those of controls.

**Table 1 Activity of α-neuraminidase in cultured fibroblasts from the two patients and the controls**

<table>
<thead>
<tr>
<th></th>
<th>α-neuraminidase activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls (15)</td>
<td>0.300 ± 0.006</td>
</tr>
<tr>
<td>Patient CRS-MS</td>
<td>0.020</td>
</tr>
<tr>
<td>Patient ML III</td>
<td>0.012</td>
</tr>
<tr>
<td>(Mucolipidosis III)</td>
<td></td>
</tr>
</tbody>
</table>

Activity is expressed as nanomoles α-N-acetyl-neuraminilactose (α2-3) hydrolysed/mg/protein/hr ± SE. In brackets the number of control cell lines in duplicate.

**Table 2 Activity of lysosomal hydrolases in cultured cells and in culture medium of cells from the patients and the controls**

<table>
<thead>
<tr>
<th></th>
<th>Controls Medium</th>
<th>CRS-MS patient Cells Medium</th>
<th>ML III patient Cells Medium</th>
</tr>
</thead>
<tbody>
<tr>
<td>arylsulphates A</td>
<td>1240 ± 436 (12)</td>
<td>16.5 ± 2.6 (3)</td>
<td>1832 20.6</td>
</tr>
<tr>
<td>β-hexosaminidase</td>
<td>8921 ± 2610 (12)</td>
<td>87 ± 21 (3)</td>
<td>11704 102</td>
</tr>
<tr>
<td>α-mannosidase</td>
<td>259 ± 64 (6)</td>
<td>0.93 ± 0.16 (3)</td>
<td>239 0.20</td>
</tr>
<tr>
<td>α-fucosidase</td>
<td>217 ± 72 (6)</td>
<td>2.5 ± 0.4 (3)</td>
<td>229 1.7</td>
</tr>
<tr>
<td>β-galactosidase</td>
<td>724 ± 180 (12)</td>
<td>1.29 ± 0.26 (3)</td>
<td>1227 0.75</td>
</tr>
</tbody>
</table>

*Activities are expressed as nanomoles substrates hydrolysed/mg protein/hr ± S.D. In brackets the number of lines examined in duplicate.
Comment

The clinical picture of this patient is strikingly similar to that reported by Rapin et al. Common to our patient and to the three previously reported cases are: (1) progressive action and stimulus-sensitive myoclonus with late onset, (2) cherry-red spot at the macula, (3) deficiency of α-neuraminidase, (4) no evidence of dementia. Our patient shares with the two sisters reported by Rapin et al an Italian origin.

According to Engel et al, the electrophysiological findings in this syndrome include: (1) low voltage fast EEG background activity; (2) paroxysmal bursts of bilateral 10–20 Hz positive spikes, time-locked but not topographically related to the myoclonus; (3) changes in SEP amplitude and waveform. Similar but not identical features were present in our patient who also showed a low voltage fast EEG background with paroxysmal bursts which can be better defined as “central fast rhythm” according to Kelly et al rather than as 10–20 positive spikes.

By the jerk-locked averaging technique, however, it was possible to detect a time-locked EEG spike preceding each myoclonic jerk. According to Shibasaki et al, this myoclonus-related cortical spike may be due to the same mechanisms underlying the giant primary SEP (also present in our patient). Whether or not these two phenomena can be related to a cortical reflex type of myoclonus is not specifically relevant to the present discussion: the question of cortical-versus-subcortical generators of myoclonus will be discussed in more detail in another paper now in preparation, based on a large number of patients with various kinds of myoclonus.

An interesting point is the effect of L 5HTP on action-sensitive and stimulus-sensitive myoclonus in our patient, both in an acute and a chronic trial. Beneficial effects of serotonin precursors have been demonstrated far less certainly in non-postanoxic myoclonus syndromes than in postanoxic ones. It has been suggested that low CSF concentrations of 5HIAA (which suggests an impairment in serotonin metabolism) may be used to predict the efficacy of serotonin precursors and that this criterion could be used to detect an L 5HTP sensitive population among patients with myoclonic syndromes. This would not hold true for our patient or for some other patients with myoclonic syndromes we have studied. According to the preliminary results of our study the only common characteristic shared by all patients responding to L 5HTP was the prominence of action and intention-myoclonus.

It should be noted, however, that such myoclonus also was present in the three patients with a cherry-red spot reported by Rapin et al to be unresponsive to L 5HTP plus carbipoda. α-neuraminidase deficiency in cultured fibroblasts from patients with mucolipidoses II and III, has been reported. In these patients, α-neuraminidase deficiency accompanied marked deficiencies in several lysosomal hydrolases in cultured cells, with increases in the corresponding activities in the cell culture media or serum. In cultured fibroblasts from patients with mucolipidosis I, a disease characterised by bone dysplasia, Hurler-life face, macular cherry-red spot, myoclonic jerks, progressive dementia and no inclusion bodies in cultured cells, the marked deficiency in α-neuraminidase is not associated with other known enzyme deficiencies. Neuraminidase deficiency has been regarded as a primary defect in this condition and the disease named “Sialidosis.”

The spectrum of α-neuraminidase deficiency was broadened by the reports of Rapin et al and O'Brien, when they described their three patients with progressive incapacitating myoclonus, macular cherry-red spots and α-neuraminidase deficiency in their cultured cells. These patients differed from those with ML I in the adult onset of the signs and symptoms, the lack of bone dysplasia and the normal intelligence. They share with the ML I patient macular cherry-red spots, myoclonus and α-neuraminidase deficiency. Moreover, additional patients with clinical features distinct from those of the patients with mucolipidoses I, II, III and the cherry-red spot myoclonus syndrome, but with α-neuraminidase deficiency, have been reported: an eight-month-old girl with normal mental development and dysostosis multiplex and an adult patient with skeletal dysplasia, cerebellar ataxia, cherry-red spot and myoclonus; conversely, patients with a phenotypic condition similar to mucolipidosis I and to the CRS-MS have been described in whom the biochemical defect was however β-galactosidase deficiency: in some patients β-galactosidase deficiency was associated and probably secondary to α-neuraminidase deficiency. It is clear, therefore, that similar phenotypic conditions can be associated with different biochemical abnormalities and that a given enzyme deficiency (for example, α-neuraminidase deficiency) can be associated with different clinical phenotypes.

Our patient was clinically and biochemically closely similar to the patients described by Rapin et al and falls into the type I Sialidosis.
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The degree of α-neuraminidase deficiency in cultured cells from our patient, as measured with α-2-3 neuraminylactose, was not distinguishable from that found in cultured cells of a patient with ML III (table 2). The two patients could not be distinguished on the basis of α-neuraminidase activity in their cultured cells. However, the two diseases are clinically and biochemically different; cultured cells from the CRS-MS patient showed both intracellular and extracellular activities of lysosomal enzymes comparable to those of controls, while cells from ML III patient showed the known decrease in intracellular and increase in extracellular lysosomal enzyme activities (table 2).

α-neuraminidase deficiency is a secondary defect in I-cell disease, but it is considered a primary defect in both mucolipidosis I and the cherry-red spot myoclonus syndrome. We believe, however, that a more detailed characterisation of α-neuraminidase activity using natural substrates is needed in order to link well-defined clinical conditions with biochemical abnormalities.

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