Short report

Serum and cerebrospinal fluid antibodies against myelin basic protein and their IgG subclass distribution in multiple sclerosis

ANTONIO GARCÍA-MERINO,* MATS AA PERSSON,† JAN ERNERUDH,‡ JUAN J. DÍAZ-GIL,§ TOMAS OLSSON

From the Departments of Neurology* and Clinical Immunology,† Karolinska Institutet, Huddinge University Hospital, Stockholm, Department of Neurology,‡ University Hospital, Linköping, Sweden, and Department of Experimental Biochemistry,§ Clínica Puerta de Hierro, Madrid, Spain

SUMMARY IgG class antibodies reactive with myelin basic protein (MBP) were determined by enzyme-linked immunosorbent assay (ELISA) in serum and cerebrospinal fluid (CSF) of 37 patients with multiple sclerosis and a control group of 32 patients with tension headache or psychoneurosis. Using standardised amounts of IgG from CSF and serum in ELISA, significantly higher mean antibody levels were found in CSF as well as in serum from the patients with multiple sclerosis. Ten (27%) of the multiple sclerosis CSF samples and 15 (41%) of the multiple sclerosis sera revealed anti MBP antibody levels exceeding 2 SD of the control group. Seven patients (19%) showed exclusive or higher levels of anti MBP antibodies in CSF, suggesting synthesis within the central nervous system. Analysis by ELISA for IgG subclasses of anti MBP antibodies revealed that they were restricted to IgG 1 in four patients and IgG 3 in one.

Immunoglobulin aberrations especially of the cerebrospinal fluid (CSF) are important characteristics of multiple sclerosis.1,2 The biological significance of the CSF IgG and its antigenic specificity are largely unknown.

Since the identification of myelin basic protein (MBP) as the antigen causing experimental allergic encephalomyelitis (EAE),3 much interest has been devoted to this protein, mainly regarding its possible pathogenetic role as a target for the immune attack in multiple sclerosis. With regard to antibodies directed against MBP, results have been conflicting. While some authors reported low levels of anti MBP antibodies in multiple sclerosis patients,4 other authors report negative results.5 The main reasons for these discrepancies include differences in methodology and selection of patients.

Most previous studies of human anti MBP antibodies have employed various radioimmunoassays and protein blotting techniques. Enzyme-linked immunosorbent assay (ELISA) may be better than radioimmunoassays, because it is more convenient and simple, and involves more stable reagents.6 The present study was undertaken to analyse whether ELISA could delineate multiple sclerosis patients with antibodies directed against MBP in their sera and CSF. A further purpose was to select positive samples for definition of IgG subclass distribution of the anti MBP antibodies, by a second ELISA. Such subclass distribution data would be important for further understanding of the biological role and production of the anti MBP antibodies.

Material and methods

Patients. Forty seven paired samples of CSF and serum taken simultaneously from 37 patients with definite (31) or probable (6) multiple sclerosis, diagnosed according to the criteria given by Rose et al.,7 were analysed. A control group of paired serum and CSF samples from 32 patients with tension headache or psychoneurosis was included. The two groups had a similar age distribution.
Routine CSF and serum studies. 15 ml of CSF was obtained by lumbar puncture, examined for cell quantities by phase contrast microscopy and centrifuged at 200 g for 10 minutes. The erythrocyte count in CSF did not exceed 106 × 106 cells/l. Determinations of albumin, IgG and IgA were carried out by immunoprecipitation nephelometry on unconcentrated CSF and on serum obtained in parallel. The CSF IgG index was calculated.8 The upper limit in our laboratory is 0-7. As indicator of the blood-brain barrier status, the CSF albumin/serum albumin ratio was determined. Oligoclonal IgG bands were demonstrated on unconcentrated CSF samples and diluted sera, according to described methods.9

Myelin basic protein. Normal appearing white matter was obtained within 4 hours after death from a patient who died of non-neurological disease. MBP was purified by ion-exchange chromatography10 and gel filtration of G-50 Superfine (Pharmacia Fine Chemicals, Uppsala, Sweden). The preparation of MBP was found to be pure on SDS polyacrylamide gel electrophoresis.

ELISA for detection of antibodies against myelin basic protein. Methods for detection of anti MBP antibodies in experimental animals11–13 were modified. To ELISA plates (Dynatech M 129B; Dynatech, Zug, Switzerland) coated over night with MBP, 5 µg/ml in 0-01 M phosphate buffer, pH 6-0, and saturated with bovine serum albumin (BSA, product No. A-4503, Sigma Chemical Co, St. Louis, MO, USA) 10 mg/ml and histones (Type II-AS, Sigma) 1 mg/ml, were added in sequence 200 µl/well aliquots of: (1) samples, in duplicate, adjusted to constant IgG concentrations of 15 µg/ml, and incubated 2 h at 37°C; (2) rabbit anti-human IgG, gamma-chains specific (Dakopatts, Copenhagen, Denmark), diluted 1:4000 and kept over night at room temperature; (3) alkaline phosphatase conjugated goat anti-rabbit IgG (Miles-Yeda Ltd, Israel), diluted 1:2500, and incubated for 4 h at room temperature. Samples and antisera were diluted in 0-01 M phosphate buffer pH 7-0, containing 1% BSA, 1-2% NaCl, 0-5% Tween 80, and 0-1% histones. After addition of substrate (disodium p-nitro-phenylphosphate from Sigma, 1 mg/ml in 10% diethanolamine buffer, pH 9-8) the reaction was read after 90 minutes at 405 nm in a Tilter Multiscan (Eflab Oy, Helsinki, Finland). Background value obtained by adding only diluting buffer instead of sample to some coated wells, was calculated in each plate and subtracted from all the readings. To avoid variations in results, these were expressed as arbitrary units related to a multiple sclerosis CSF sample, which was found to have higher optical density values than the mean of control CSF samples in preliminary experiments, and run in each plate.

ELISA for determination of anti MBP IgG antibody subclasses. Monoclonal antibodies to IgG subclasses14 were purchased from Seward Laboratories, Bedford, Buckinghamshire, UK. The following monoclonals were used: anti-IgG1 BAm 15, anti-IgG2 BAm 10, anti-IgG3 BAm 8, and anti-IgG4 BAm 16. They were purified from ascitic fluid as described.15 Rabbit anti-mouse IgG antisera was supplied by Dakopatts A/S (product No. Z 109), and absorbed to human proteins before use. Alkaline phosphatase conjugated goat anti-rabbit IgG was purchased from Sigma (product No. F 8025). The method for IgG subclass determination has been described elsewhere.15

Results

Routine studies of CSF and serum revealed that all except three of the 37 patients with multiple sclerosis had increased CSF IgG index, and all but one had oligoclonal IgG bands. All 32 patients in the control group were negative in these respects.

ELISA for detection of antibodies reactive with MBP revealed a significantly higher (p < 0-01; Mann-Whitney's U contrast test) mean value in the CSF of multiple sclerosis patients compared with controls (Fig. 1). Ten of the 37 multiple sclerosis patients (27%) had values in CSF exceeding 2 SD of the controls. In sera, the mean value of anti MBP antibodies was also higher for multiple sclerosis patients than controls (p < 0-01) (fig). Values in 15 of 37 multiple sclerosis sera (40%) exceeded 2 SD of the control sera.

A comparison of CSF and serum values of the same individuals exceeding 2 SD of controls, revealed eight patients having anti MBP antibodies both in
serum and CSF, five of them with higher values in CSF than serum (arbitrary units in CSF/arb
arbitrary units in serum > 1-0), and three with higher values in serum than in CSF (arbitrary
units in CSF < 1-0). Two patients had anti MBP antibodies in CSF only, and seven patients had such antibodies in serum only. This means that we obtained evidence for production of anti MBP antibodies within the CNS in seven (19%) of our multiple sclerosis patients.

Ten patients with multiple sclerosis in whom anti MBP antibody values in CSF and/or serum exceeded 2 SD of controls were selected for IgG subclass analysis. In seven of them both CSF and serum were studied, in only one only CSF, and in the remaining two only serum. Using optical densities, read at 405 nm over 0-100 after background subtraction, to delineate subclass specific antibodies a ELISA revealed the IgG subclass distribution of the anti MBP antibodies in five patients. Three of them had increased anti MBP levels in both serum and CSF, one had increased levels in serum only, and one in CSF only. In four of the patients, the antibodies were of the IgG 1 subclass only, in the remaining one of the IgG 3 only. In this last mentioned patient the subclass distribution was identical in three samples taken 6 months apart (table).

Discussion

The present study corroborates previous findings of anti MBP antibodies of IgG class in CSF and serum of a substantial number of multiple sclerosis patients, compared with controls without evidence of inflammatory disease of the CNS. Nevertheless there is a large proportion of patients with multiple sclerosis who in various analyses do not show detectable amounts of anti MBP antibodies. From these data the biological significance of anti MBP antibodies cannot be evaluated; whether they are a part of a primary, pathogenetically important immune response or represent secondary responses of unknown significance remains unanswered.

Firstly, MBP is a strongly positively charged molecule and by this it is very prone to bind IgG nonspecifically. This makes it difficult to estimate amounts of anti MBP specific antibodies. In the present study some of these difficulties have been overcome by applying constant amounts of serum IgG and CSF IgG, and by the use of histones which are known to inhibit non-specific binding when applied at high concentrations. These difficulties have also the implication that lower amounts of antibodies may be “hidden” in a large non-specific background, and the proportion of patients having anti MBP antibodies may well be higher. Secondly, antibodies against MBP or other structural nervous tissue antigens may be produced in vivo but absorbed to the target and, therefore, not be detected as free immunoglobulins. In fact, we have recently demonstrated in animals with chronic relapsing encephalomyelitis that production of antibodies against myelin or MBP is easily detected in vitro culture supernatants of CNS lymphocytes but not in brain extracts. Thirdly, whether anti MBP antibodies belonging to other immunoglobulin classes may appear in multiple sclerosis remains to be elucidated.

The site of production of anti MBP antibodies is controversial; intra- or extrathecal or both. It has been claimed that antibodies appeared in CSF only after passive transudation from serum. In the present study parallel estimations of anti MBP antibodies was made both in serum and CSF samples. Higher anti MBP antibody values in CSF compared with serum would indicate an intrathecal production site and was found in seven of our 37 patients (19%). When values were similar in CSF and serum or higher in serum, the site of production may not be accurately

<table>
<thead>
<tr>
<th>Table</th>
<th>IgG subclass distribution of anti MBP antibodies in CSF and serum from five patients with multiple sclerosis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patient</td>
<td>Type of sample</td>
</tr>
<tr>
<td>1</td>
<td>CSF</td>
</tr>
<tr>
<td></td>
<td>serum (a)</td>
</tr>
<tr>
<td></td>
<td>(b)</td>
</tr>
<tr>
<td></td>
<td>(c)</td>
</tr>
<tr>
<td>2</td>
<td>CSF</td>
</tr>
<tr>
<td></td>
<td>serum</td>
</tr>
<tr>
<td>3</td>
<td>CSF</td>
</tr>
<tr>
<td></td>
<td>serum</td>
</tr>
<tr>
<td>4</td>
<td>CSF</td>
</tr>
<tr>
<td></td>
<td>serum</td>
</tr>
<tr>
<td>5</td>
<td>CSF</td>
</tr>
<tr>
<td></td>
<td>serum</td>
</tr>
</tbody>
</table>

*Optical density at nm values, read at 90 min. Background subtracted. CSF and serum (a) drawn at the same occasion; serum (b) six months before, and serum (c) six months afterwards. Arbitrarily, samples with OD values 0-100 or higher are regarded as positive and underlined.
Serum and CSF antibodies against myelin basic protein and their IgG subclasses in multiple sclerosis

We thank Professor Hans Link, Department of Neurology, Karolinska Institutet, Huddinge University Hospital, Stockholm, for valuable critical advice, and Dr Isabel Millán for her help in statistical calculations.

References

5. Chou C-HJ, Tourtellotte WW, Kibler RF. Failure to detect antibodies to myelin basic protein or peptic fragments of myelin basic protein in CSF of patients with MS. Neurology (Cleveland) 1983;33:24–8.
15. Persson MAA, Hammarström L, Smith CIE. Enzyme-


20 Hammarström L, Granström M, Oxelius V, Persson MAA, Smith CIE. IgG subclass distribution of anti-

Garcia-Merino, Persson, Ernerudh, Diaz-Gil, Olsson


Serum and cerebrospinal fluid antibodies against myelin basic protein and their IgG subclass distribution in multiple sclerosis.

A García-Merino, M A Persson, J Ernerudh, J J Díaz-Gil and T Olsson

*J Neurol Neurosurg Psychiatry* 1986 49: 1066-1070
doi: 10.1136/jnnp.49.9.1066

Updated information and services can be found at:
[http://jnnp.bmj.com/content/49/9/1066](http://jnnp.bmj.com/content/49/9/1066)

**Email alerting service**

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
[http://group.bmj.com/group/rights-licensing/permissions](http://group.bmj.com/group/rights-licensing/permissions)

To order reprints go to:
[http://journals.bmj.com/cgi/reprintform](http://journals.bmj.com/cgi/reprintform)

To subscribe to BMJ go to:
[http://group.bmj.com/subscribe/](http://group.bmj.com/subscribe/)