Endoneurial capillary abnormalities in mild human diabetic neuropathy


Abstract
Microvascular factors have been implicated in the pathogenesis of human diabetic neuropathy. The extent of microangiopathy was assessed in 15 diabetic patients with clinically mild neuropathy and compared with eight age matched control subjects. Endoneurial capillary density was reduced (p < 0.04) and correlated significantly with reduced myelinated fibre density (p < 0.01). Both basement membrane area (p < 0.0001) and endothelial cell profile number per capillary (p < 0.002) were significantly increased in diabetic patients and correlated significantly with both neurophysiological and neuropathological measures of neuropathic severity. There was no evidence of endothelial cell hypertrophy as assessed by either cross sectional endothelial cell area or a reduction in luminal size. Furthermore, the percentage of closed vessels did not differ between diabetic patients and control subjects and failed to relate to measures of neuropathic severity. It was concluded that microvascular abnormalities are present in patients with clinically mild human diabetic neuropathy, and that these data provide further support for the role of endoneurial capillary disease in the development of this condition.

The cause of human diabetic neuropathy is considered to be multifactorial, with both metabolic and ischaemic factors playing an important part. Whereas much recent research has focused on the polyol pathway and related metabolic abnormalities in relation to the aetiologypathogenesis of human diabetic neuropathy, several studies have characterised the presence of microangiopathy in patients with established human diabetic neuropathy. Thus a considerable amount of evidence has accumulated on the role of microvascular disease and resultant hypoxia in the development of neuropathy. Fibre loss in human diabetic neuropathy has been shown to be multifocal, providing further support for a role of ischaemia in its development. A recent study has also shown that multifocal fibre loss and hence ischaemia may well play an important part in the development of neuropathy in older non-insulin dependent diabetic patients. However, more recent studies on the pattern of fibre loss and extent of microangiography have failed to support these earlier findings.

A microvascular basis to human diabetic neuropathy cannot, however, be discounted and this subject requires further systematic study. We examined microvascular abnormalities in a group of patients with clinically mild neuropathy and attempted to relate these abnormalities to measures of neuropathic severity.

Materials and methods

Subjects
Fifteen patients (12 men and three women) with mild diabetic neuropathy were studied; all fulfilled the following criteria: (a) either no symptoms or a history of mild neuropathic symptomatology; (b) no history of foot ulceration, peripheral vascular disease, renal disease, or any other conditions known to cause neuropathy; (c) evidence of reduced but not absent sensation in the feet to both small and large fibre modalities, and with normal foot pulses; (d) vibration perception threshold by a biothesiometer over the medial malleolus above the 95th centile for the age matched normal value but < 40 volts; (e) peroneal nerve motor conduction velocity below the 5th centile for the age matched normal value, but > 30 m/s; (f) normal serum creatinine concentration (< 130 μmol/l).

All electrophysiological nerve parameters were assessed by the same neurophysiologist (WS). The study was approved by the Central Manchester Hospitals Ethical Committee and all patients gave written informed consent after the nature of the procedures had been explained.

Tissue biopsy
Fascicular sural nerve biopsy was performed under local anaesthesia by a consultant neurosurgeon (RHL). For ultrastructural examination the nerve specimen was fixed primarily in glutaraldehyde in cacodylate buffer and secondarily in osmium tetroxide. After dehydration in a graded series of ethanol the tissue was embedded in epon resin with propylene oxide as an intermediary. Sural nerve biopsies of six age matched organ donors and two patients with traumatic amputation were used as controls. All such patients had normal cardiovascular and renal function and no known cause of peripheral neuropathy before biopsy by the same surgeon.

Histological and morphometric procedures
The myelinated fibre density was derived by directly counting fibres from montages (×1000) of all fascicles from the biopsies.
Mean fascicular area and endoneurial capillary density were assessed directly from semithin sections, stained with thionin and acridine orange, by using a camera lucida (Nachet, Evry, France) and sonic digitiser. All microvessels without a complete layer of cells (pericyte or smooth muscle cells) surrounding the endothelial cells were considered to be capillaries and were photographed at a final magnification of ×10 000–15 000. The luminal, endothelial cell, and basement membrane areas were derived from programme digitisation (Commodore PET Microcomputer System) of each electron micrograph. The endothelial cell profile and nuclear number and pericyte nuclear number per capillary were counted directly from the electron micrographs by using previously described techniques. Closure of endoneurial vessels was assessed by surveying all vessels by electron microscopy. When endothelial cell surfaces were opposed in the midline of any vessel it was considered to be closed (fig 1C).

### Table 1 Results of peripheral nerve electrophysiology and quantitative sensory tests in diabetic patients

<table>
<thead>
<tr>
<th>Test</th>
<th>Diabetic Patients (n = 15)</th>
<th>% Abnormal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (SE) peroneal motor nerve conduction velocity (m/s)</td>
<td>37.3 (3.3)</td>
<td>100</td>
</tr>
<tr>
<td>Mean (SE) sural nerve conduction velocity (m/s)</td>
<td>38.6 (5.4)</td>
<td>66</td>
</tr>
<tr>
<td>Mean (SE) sural sensory action potential amplitude (μV)</td>
<td>5.4 (4.5)</td>
<td>40</td>
</tr>
<tr>
<td>Mean (SE) vibration perception threshold (volts)</td>
<td>26.4 (7.0)</td>
<td>100</td>
</tr>
<tr>
<td>Mean (SE) thermal discrimination (°C)</td>
<td>3.6 (7.2)</td>
<td>63</td>
</tr>
</tbody>
</table>

**Results**

**CLINICAL AND NEUROPHYSIOLOGICAL DETAILS**

The mean age of control subjects was 47.5 (median 47.5, SD 26.5, range 18–85) years. The mean age of the diabetic patients was 46.9 (median 48.0, SD 10.9, range 27–59) years and the mean duration of diabetes was 16.3 years. Seven patients had insulin dependent diabetes and eight had non-insulin dependent diabetes. All but three subjects had preserved ankle reflexes. Details of electrophysiological and quantitative sensory testing are provided in table 1. Sural nerve conduction velocity was normal in one third of patients. Sural action potentials were recordable in all patients and were of normal amplitude in 60%. According to published criteria the patients in this study would be classified as having either stage 2 (asymptomatic) or stage 2 (symptomatic) neuropathy.

**MICROANGIOPATHY**

**Qualitative observations**—Electron micrographs of representative vessels from control subjects and diabetic patients are presented in figure 1. The basement membrane was considerably thickened in the form of concentric, reduplicated layers. Endothelial cells showed hyperplasia as did pericyte profiles and there was no evidence of either endothelial or pericyte cell degeneration. Both semithin and ultrathin sections failed to show vascular occlusion with fibrin or platelets, and no inflammatory infiltrates were observed in the endoneurium, perineurium, or epineurium. The perineurium itself was observed to be thickened in diabetic patients.

**Quantitative observations**—Capillary density (p < 0.04) and myelinated fibre density (p < 0.01) were significantly reduced in diabetic patients, without a significant increase in fascicular area. Basement membrane area (p < 0.0001), endothelial cell profile (p < 0.002), and pericyte nuclear number (p < 0.02) per
Table 2  Endoneurial capillary morphology in diabetic patients and age matched control subjects presented as mean (SE)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control subjects (n = 8)</th>
<th>Diabetic patients (n = 15)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>47 (9.3)</td>
<td>46 (3.0)</td>
<td>NS</td>
</tr>
<tr>
<td>Age (range)</td>
<td>18-85</td>
<td>27-59</td>
<td></td>
</tr>
<tr>
<td>Mean fascicular area (mm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>10 (0.1)</td>
<td>20 (0.5)</td>
<td></td>
</tr>
<tr>
<td>Myelinated fibre density (No/mm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>656 (407)</td>
<td>404 (540)</td>
<td>0.01</td>
</tr>
<tr>
<td>Capillary density (No/mm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>67.9 (4.8)</td>
<td>55.3 (4.4)</td>
<td>0.04</td>
</tr>
<tr>
<td>Basement membrane area (µm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>47.34 (5.37)</td>
<td>156.5 (15.9)</td>
<td>0.0001</td>
</tr>
<tr>
<td>Endothelial cell area (µm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>32.54 (6.18)</td>
<td>25.59 (3.32)</td>
<td>NS</td>
</tr>
<tr>
<td>Luminal area (µm&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>32.61 (3.71)</td>
<td>44.87 (5.45)</td>
<td>NS</td>
</tr>
<tr>
<td>Endothelial cell number</td>
<td>410 (0.30)</td>
<td>582 (0.21)</td>
<td>0.002</td>
</tr>
<tr>
<td>Endothelial cell nuclear number</td>
<td>120 (0.10)</td>
<td>1.68 (0.12)</td>
<td>NS</td>
</tr>
<tr>
<td>Pericyte nuclear number</td>
<td>0.59 (0.05)</td>
<td>0.83 (0.08)</td>
<td>0.02</td>
</tr>
<tr>
<td>Endothelial-pericyte cell nuclear ratio</td>
<td>2.30 (0.20)</td>
<td>2.26 (0.26)</td>
<td>NS</td>
</tr>
</tbody>
</table>

Diabetic patients were significantly increased in diabetic patients (table 2). The endothelial-pericyte cell nuclear ratio were similar in control subjects and diabetic patients (table 2). The endothelial cell area was slightly lower in diabetic patients than in control subjects. Furthermore, the luminal area was slightly greater in diabetic patients than in control subjects, and the percentage of closed capillaries was in fact slightly less in diabetic patients than in control subjects (fig 2). These differences were not statistically significant.

**Discussion**

Microangiopathy is prominent in the endoneurial vessels of patients with established human diabetic neuropathy. We have now studied diabetic patients with clinically mild neuropathy and have shown significant endoneurial microangiopathy, which relates to measures of neuropathic severity. The mild nature of neuropathy is supported by the electrophysiological results where sural nerve conduction velocity was normal in one third of patients. A reduction in capillary density was observed without a significant increase in fascicular area, confirming our previous findings in patients with severe neuropathy. Such a reduction would be expected to increase intercapillary distance and reduce nerve blood flow and hence endoneurial oxygen levels. A recent study showed an increase in capillary density in patients with newly presenting and chronic painful neuropathy and a decrease in those patients with recurrent foot ulceration. Such a disparity in capillary density among different syndromes of diabetic neuropathy would perhaps explain the failure of a recent study to find any difference in capillary density between diabetic patients and control subjects as it incorporated patients with a variety of different syndromes of diabetic neuropathy, including those with foot ulceration.

Endothelial cells exhibit both hyperplasia and hperthropy in established diabetic neuropathy, and they are known to be markers of a range of metabolic, haemostatic, and

**Figure 2** Percentage of closed endoneurial capillaries (top) and endothelial cell and luminal area (bottom) in diabetic patients and control subjects expressed as mean (SE).

**Figure 3** Correlation plots between PMCV (peroneal motor conduction velocity) and MFD (myelinated fibre density) with basement membrane area (top) and endothelial cell profile number (bottom).
pathological abnormalities in diabetic patients.\textsuperscript{26} We found endothelial cell hyperplasia, which again confirms previous findings in patients with established neuropathy.\textsuperscript{4,11-16} Other studies have also found endothelial cell hyperplasia in patients with established neuropathy.\textsuperscript{4,11-15} However, the present study, which included patients with clinically mild neuropathy, failed to show this, in keeping with previous studies that have included some patients with mild neuropathy\textsuperscript{6} and without neuropathy.\textsuperscript{4,15} Thus endothelial cell hyperplasia may well be a late feature of diabetic microangiopathy.

Pericyte cell nuclear hyperplasia was observed, confirming previous findings in established neuropathy.\textsuperscript{13} Basement membrane thickening was also found in the present study.\textsuperscript{4,10-15} It has been described in a range of acquired\textsuperscript{27} and inherited neuropathies\textsuperscript{18} and is presumably due to primary precapillary denervation and hence capillary hypertension.\textsuperscript{28} In diabetes both primary precapillary denervation\textsuperscript{58} and the reduced degradation of basement membrane due to glycosylation\textsuperscript{29} have central roles. It is therefore not surprising that a recent study of hereditary motor and sensory neuropathy found basement membrane thickening and increased luminal size almost comparable to those in diabetic patients with neuropathy.\textsuperscript{15} Therefore these abnormalities result in limited vascular reactivity and hence abnormal microvascular function.\textsuperscript{30-32}

We did not find luminal occlusion on direct electron microscopic observation of vessel closure and from morphometric assessment of luminal size, contrary to the observations of a previous study.\textsuperscript{4} Support for our findings is found in other ultrastructural studies that have shown an increase in luminal size.\textsuperscript{4,12-15} We also failed to find any relation between extent of vessel closure and measures of neuropathic severity, confirming a previous finding.\textsuperscript{13}

A range of microvascular abnormalities was correlated with measures of neuropathic severity such as loss of myelinated fibres and neurophysiological abnormalities. This has been shown previously in patients with established neuropathy.\textsuperscript{13,14} However, all measures of microangiography failed to relate to age and abnormalities of either vibration or thermal perception. This finding may simply reflect the large coefficient of variation inherent in these tests.\textsuperscript{33,34}

In conclusion, we clearly showed a microangiopathy in patients with clinically mild neuropathy, which relates significantly to measures of neuropathic severity. These findings provide further support for the role of small vessel disease in the development of human diabetic neuropathy.

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\footnotesize{
\begin{table}
\centering
\begin{tabular}{|l|c|c|c|c|c|c|}
\hline
Age & PNCV & SNCV & SNAP & MFD & VPT & TDT \\
\hline
\textbf{Capillary density (no/mm\textsuperscript{2})} & 0.11 & 0.25 & 0.44 & 0.29 & 0.65 & 0.30 & 0.12 \\
Basement membrane area (\um\textsuperscript{2}) & 0.21 & 0.55 & 0.55 & 0.29 & 0.52 & 0.19 & 0.38 \\
Endothelial cell profile number & (p<0.05) & 0.46 & 0.25 & 0.39 & 0.44 & 0.36 & 0.46 \\
Endothelial cell area (\um\textsuperscript{2}) & 0.19 & 0.48 & 0.52 & 0.10 & 0.39 & 0.15 & 0.16 \\
Luminal area (\um\textsuperscript{2}) & 0.19 & 0.70 & 0.49 & 0.44 & 0.46 & 0.36 & 0.46 \\
Closed capillaries (%) & 0.17 & 0.15 & 0.08 & 0.29 & 0.08 & 0.09 & 0.07 \\
\hline
\end{tabular}
\caption{Correlation of capillary morphometry with age, nerve electrophysiology, myelinated fibre density and sensory testing presented as Spearman’s rank correlation coefficient and degree of significance}
\end{table}
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\begin{itemize}
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