Skeletal muscle bioenergetics in the chronic fatigue syndrome

Phillip R J Barnes, Doris J Taylor, Graham J Kemp, George K Radda

Abstract
Skeletal muscle bioenergetics and control of intracellular pH have been investigated in 46 patients with chronic fatigue syndrome by phosphorus magnetic resonance spectroscopy. The results have been compared with those from healthy controls and from a group of patients with mitochondrial cytopathies affecting skeletal muscle. No consistent abnormalities of glycolysis, mitochondrial metabolism or pH regulation were identified in the group when taken as a whole, although in 12 of the 46 patients the relationship between pH and phosphocreatine utilisation during exercise fell outside the normal range. Of these, 6 patients showed increased acidification relative to phosphocreatine depletion while 6 showed reduced acidification. These findings do not support the hypothesis that any specific metabolic abnormality underlies fatigue in this syndrome although abnormalities may be present in a minority of patients.

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Methods and subjects
Phosphorus magnetic resonance spectroscopy
Spectra were collected from the forearm muscle flexor digitorum superficialis using techniques as previously described. Briefly, the subject lay supine with the arm abducted to 90° and placed so that the flexor digitorum superficialis lay in the centre of a 1.89 Tesla 30 cm bore superconducting magnet which was interfaced with a Fourier transform spectrometer (Oxford Research Systems, Oxford, UK). Radiofrequency pulses of 20 μs length were delivered at 2 s intervals using a 2.5 cm diameter surface coil, thus optimising 31P signal collection. The 90° pulse with this coil was 16 μs. A spectrum consisting of 128 accumulated free induction decays (FIDs) was collected from resting muscle to provide a baseline for exercise and recovery. During exercise (see below) spectra were collected over 64s (32 FIDs) and during recovery they were collected over 32 or 64s for a period of 11 minutes.

Determination of intracellular metabolite concentrations and pH
Metabolite ratios were calculated from the peak areas of phosphocreatine (PCr), inorganic phosphate (P), and β-ATP (cor-
Table 1 Metabolite ratios in resting skeletal muscle

<table>
<thead>
<tr>
<th>Group</th>
<th>Controls</th>
<th>Mito Myopathy</th>
<th>CFS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>P*</td>
<td>P*</td>
<td></td>
</tr>
<tr>
<td></td>
<td>(A vs B)</td>
<td>(B vs C)</td>
<td></td>
</tr>
<tr>
<td>n</td>
<td>19</td>
<td>21</td>
<td>46</td>
</tr>
<tr>
<td>pH</td>
<td>7.03 (0.03)</td>
<td>6.77</td>
<td>7.03 (0.05)</td>
</tr>
<tr>
<td>PCr/ATP</td>
<td>3.08 (0.09)</td>
<td>0.0001</td>
<td>2.52 (0.40)</td>
</tr>
<tr>
<td>P/ATP</td>
<td>0.34 (0.07)</td>
<td>0.05</td>
<td>0.40 (0.11)</td>
</tr>
<tr>
<td>Pi/P</td>
<td>9.40 (1.90)</td>
<td>0.002</td>
<td>6.90 (3.20)</td>
</tr>
<tr>
<td>PCr(PCR + Pi)</td>
<td>0.90 (0.02)</td>
<td>0.001</td>
<td>0.86 (0.05)</td>
</tr>
<tr>
<td>ADP (μM)</td>
<td>6 (3)</td>
<td>0.0001</td>
<td>22 (15)</td>
</tr>
</tbody>
</table>

All values given as means with standard deviations in parentheses.

*p values refer to differences between the groups stated at the head of the column in which they appear. There was no significant difference between patients with CFS and controls for any parameter (p > 0.05 for each).

Figure 1 (A) Changes in the ratio PCr/(PCR + Pi) during exercise in patients with CFS (dotted line and closed symbols) and controls (solid line and open symbols). Means (1 SD); (B) Changes in pH, during the same exercise protocol (symbols as for fig 1A).

Exercise protocol
All controls and patients performed an exercise protocol which consisted of squeezing a rubber sphgmonomanometer bulb 22 times per minute. During the first 5 minutes of exercise the maximum pressure that could be generated by emptying the bulb was preset at 100 mm Hg and this was increased to 300 mm Hg for the final 2.5 minutes. Subjects who could not achieve these pressures simply squeezed as hard as they could.

Statistical analysis
All data are expressed as their mean with standard deviation (SD) in parentheses unless otherwise stated, and a range is given for some control parameters. Significance was calculated using the Mann-Whitney U test.

Subjects
The data presented are from 46 patients (24 men and 22 women aged 18–69 years; mean 36) referred by hospital specialists with a diagnosis of a chronic fatigue syndrome, and include the cases previously reported by us.8 In all cases there was a clear clinical history of a preceding viral-type illness followed by incapacitating fatigue and in no case was there evidence of any other illness to account for the fatigue or other symptoms. Symptoms had been present for a mean of 35 months (range 6–132) at the time of study and all cases satisfied the Green College criteria for diagnosis of a chronic fatigue syndrome.1 In 21 cases there was additional laboratory evidence of recent viral infection at the time of referral (thus satisfying the strict criteria for diagnosis of a post-infectious fatigue syndrome); in the remaining 25 patients tests had either not been performed or they were negative. Data are also shown from 21 patients with biopsy-proven mitochondrial cytopathies (ages 12–70 years; mean 33). Control subjects were sedentary volunteers (ages 20–79; mean 41) and do not include any trained athletes. All studies were approved by the Central Oxford Research Ethics Committee and informed verbal and written consent was obtained in all cases.

Results
Resting muscle
Results for resting skeletal muscle are shown in table 1. There were no significant differences between patients with chronic fatigue syndrome and controls. Patients with mitochondrial myopathies had significantly lower PCr/ATP and PCr/Pi ratios and higher resting [ADP] compared with either controls or patients with CFS. Subgroup analysis of those patients with CFS in whom positive evidence was found of viral infection showed no difference in resting muscle compared...
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Table 2 Recovery of metabolites after exercise

<table>
<thead>
<tr>
<th>Group</th>
<th>Controls</th>
<th>Mitochondrial myopathy</th>
<th>CFS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>P*</td>
<td>P*</td>
<td></td>
</tr>
<tr>
<td>Init rate PCr recovery (mM/min)</td>
<td>A (A vs B)</td>
<td>B (B vs C)</td>
<td>C</td>
</tr>
<tr>
<td>t0 PCr (s)</td>
<td>20 (13)</td>
<td>13 (8)</td>
<td>0-007 20 (9)</td>
</tr>
<tr>
<td>t0P (s)</td>
<td>50 (17)</td>
<td>92 (62)</td>
<td>0-02  53 (30)</td>
</tr>
<tr>
<td>t0ADP (s)</td>
<td>28 (7)</td>
<td>65 (44)</td>
<td>0-003 31 (11)</td>
</tr>
<tr>
<td></td>
<td>14 (7)</td>
<td>53 (68)</td>
<td>0-0001 14 (5)</td>
</tr>
</tbody>
</table>

All values given as means with standard deviations in parentheses.

Exercise and recovery

The changes in PCr (expressed as PCr/(PCr + P]) and pH during exercise are summarised in fig 1. There are no significant differences between the CFS patient group and controls, although the scatter in the pH data from patients was greater. Data from recovery are shown in table 2. Again there are no differences between the control and CFS groups (including the CFS subgroup with positive evidence of viral infection), whereas patients with mitochondrial myopathy have prolonged recovery half times for [PCr]). Recovery data for pH were only obtained in a minority of patients and controls due to loss of the P, signal during recovery and therefore are not presented here.

The relationship between changes in [PCr] and pH during exercise may be expressed as the calculated free [ADP], since this is derived from the creatine kinase equilibrium expression in which [PCr] and pH are the only other major variables. When pH is lower for a given [PCr] than that in controls, the calculated [ADP] is lower than in controls. Figure 2 shows the [ADP] at the end of exercise ([ADP]), for the three groups of subjects, and the majority of CFS patients fall within the control range. It is possible, however, to identify 6 patients who have lower than normal [ADP], due to greater than normal acidification during exercise. Analysing these separately, there were no significant differences between this subgroup and the control group at rest but the half time for recovery of [PCr] was significantly prolonged (63s [SD 33] vs 50s [17]; p = 0-002). There were also 6 patients with CFS who show an [ADP], above the control range (signifying less acidification of muscle than is usual for the degree of PCr depletion). When compared with controls this subgroup had a lower ratio for PCr/ATP in resting muscle (2.79 [SD 0-21] vs 3.08 (0-19); p = 0-002) and higher [ADP] at rest (11 µM [SD 3] vs 6 µM [3]; p = 0-007). Recovery half times were not significantly different from controls, however. If this subgroup is also compared with those patients with mitochondrial myopathy whose [ADP], is above the normal range (11/21 patients) the latter have a still lower resting PCr/ATP (2-42 [SD 0-48]; p = 0-03) and higher resting [ADP] (25 µM [SD 15]; p = 0-02) and the recovery half time for [ADP] is significantly longer in the mitochondrial myopathy group (65s [SD 79] vs 12s [2] in CFS; p = 0-006). Three of these 6 CFS patients with high [ADP], had had muscle biopsies and all were normal without evidence of ragged red fibres.

Discussion

The pathophysiological basis of the chronic fatigue syndrome (including post-viral fatigue syndrome, “myalgic encephalomyelitis” and neurasthenia) remains a mystery. Many of the symptoms relate to muscle with prominent fatigue and pain at rest and on minimal exertion. Exertional muscle pain is also a feature of a number of metabolic myopathies including glycogenoses of muscle, fatty acid oxidation defects and adenylate deaminase deficiency although it is uncommon in mitochondrial myopathies. A previous publication from our group reported results from a single patient with post-varicella fatigue syndrome who showed a repeatable abnormal metabolic response to aerobic exercise with greater acidification than that seen in controls and slow recovery of [PCr] following exercise. This was interpreted as being due to abnormal control of glycolysis leading to increased lactic acid production, although others have

Figure 2 Distribution histogram of the free [ADP] at the end of exercise ([ADP]) in controls, patients with CFS and patients with mitochondrial myopathies (MM). The dotted lines show the upper and lower limits of the control range.
used the data as evidence of mitochondrial dysfunction.2 We have shown here that in a group of 46 patients with CFS following a standard exercise protocol there are no consistent bioenergetic abnormalities. In particular in the group as a whole there is no evidence of a mitochondrial defect nor of a consistent abnormality of pH regulation. This is in keeping with other in vitro studies of mitochondrial and glycolytic function8,9 and with preliminary results using 31P MRS to study the tibialis anterior.10 Nevertheless, the variation in the results from CFS patients was greater than that seen in controls and, since there are difficulties in the diagnosis of CFS and the possibility exists that it represents the clinical manifestations of a number of as yet unidentified pathologies, it is pertinent to consider further the metabolic responses of those subgroups of patients described above who fall outside the normal range.

Because the degree of metabolic change in muscle during exercise will depend to a large degree on the level of fitness and “training” it is most appropriate to compare metabolic parameters rather than to look at each separately. There is known to be a well defined relationship between changes in pH and the utilisation of PCR.12 In particular, significant acidification of muscle does not occur until PCR falls to approximately 60% of its resting level. In those patients with CFS who acidify more than controls and whose [ADP], therefore is low (fig 2), this “threshold” for acidification is unchanged (data not shown). This pattern of pH response is also quite different from that in patients with mitochondrial myopathies11 (see results). In the latter the degree of intracellular acidification is less than in controls despite the lactic acidemia which is often present16 and possibly represents a compensatory increase in proton extrusion from the cell. The group of CFS patients with marked acidification do have slow [PCR] recovery but this is to be expected since their [ADP], is low and rates of mitochondrial ATP synthesis (and thus PCR resynthesis)17 depend upon [ADP] in vitro.14 Further evidence against a significant mitochondrial defect in this subgroup is the normal recovery half time for [ADP] which is itself a sensitive indicator of mitochondrial function.18 The response to exercise in this subgroup resembles that seen in a group of healthy volunteers exercising at a much higher than normal workload (D Taylor, G Kemp, unpublished data) in which the pH fell rapidly and [ADP], was low. In the high workload control group the recovery half time for [PCR] was low (as expected for the low [ADP],) but that for [ADP] itself was normal. This shows that changes in relative work done affect the relative aerobic and anaerobic contributions to ATP synthesis and suggest that different degrees of fitness may explain the results. In particular, studies of trained athletes show that they have improved oxidative metabolism and reduced acidification during exercise both in specifically trained and other muscle groups.18,19 “Detraining” due to reduced activity is likely to produce the reverse changes and would explain the present results in this small subgroup.

A similar consideration of that subgroup of CFS patients who have high [ADP], (reflecting less intracellular acidification as compared to PCR utilisation than controls) shows that they have slightly lower resting PCR/ATP and higher [ADP] than controls, although they remain significantly different from the mitochondrial myopathy group. Recovery data from this group of CFS patients are entirely normal, however, so that there is no evidence of significant mitochondrial dysfunction. The resting abnormalities may reflect the non-specific mild reductions in mitochondrial enzyme activities reported by others2 but they show no evidence of any specific metabolic defect.

In conclusion we have failed to demonstrate any specific metabolic abnormalities in the skeletal muscle of patients with chronic fatigue syndrome. In particular there is no evidence of a functional bioenergetic correlate of the morphological abnormalities reported by Behan et al in 80% of their patients. We have found a relationship between intracellular acidification and PCR utilisation that lies outside the control range in 12 out of 46 patients, but equal numbers showed abnormally great and abnormally little acidification. We cannot rule out minor metabolic abnormalities in some of these patients but it is clear that there is no specific bioenergetic abnormality in CFS that can explain the prominent symptoms of fatigue. Furthermore, although some minor electrophysiological abnormalities have been reported on single fibre EMG20 and in protein synthetic rates,21 careful assessment of strength and fatigability have shown no evidence for a peripheral neuromuscular component to the symptoms found in the disease and a central cause for fatigue seems more likely.22,23

These experiments were carried out over a number of years and many members of the MRC Biochemical and Clinical Magnetic Resonance Unit have contributed to them and their contributions are most gratefully acknowledged. In particular Drs D Arnold, P Bore, B Raimagepalan and P Styles were closely involved in the planning of studies and in early experiments. The assistance of Mrs R Cooper in collating the data is also acknowledged. We thank those clinicians who have referred their patients to us. PRJB and QJK acknowledge personal support from the Muscle Dystrophy Group of Great Britain and Northern Ireland. The Unit is supported by the Medical Research Council and the British Heart Foundation.

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