A study on a new antineural antibody in a case of paraneoplastic sensory neuropathy associated with breast carcinoma

Teruaki Iwahashi, Atsushi Inoue, Chang-Sung Koh, Nobuo Yanagisawa

Abstract
Paraneoplastic sensory neuropathy is a remote effect of cancer, usually associated with small cell lung carcinoma and anti-Hu antibody. This report details the case of a 59 year old woman with a breast carcinoma and a paraneoplastic sensory neuropathy characterised by chronic asymmetric sensory neuropathy. Anti-Hu antibody was not detected in her serum; nor were other known antineuronal antibodies such as anti-Ri and Yo. However, we have found an antineural antibody that reacted to a 106 kDa mouse neural antigen which has not yet been reported. Immunohistochemically, this antineural antibody bound to the posterior grey horn. This finding suggests that this antineural antibody may play an important part in the pathogenesis of the sensory neuropathy of this patient.

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Neurological paraneoplastic syndrome is a remote effect of carcinoma and some other types of neoplasia on the central or peripheral nervous system. Paraneoplastic sensory neuropathy is one of the neurological paraneoplastic syndromes, characterised by a subacute clinical course of progressive, usually severe, sensory ataxia that often precedes or closely follows the diagnosis of cancer.1–3 Although the pathogenesis of neurological paraneoplastic syndromes is not known, recent discoveries of several types of antibodies to neural tissue components in patients with paraneoplastic syndromes, such as paraneoplastic sensory neuropathy, paraneoplastic cerebellar degeneration, paraneoplastic encephalomyelitis, opsoclonus-myoclonus syndrome, and Lambert-Eaton myasthenia syndrome have led to speculation that an autoimmune mechanism for neurological paraneoplastic syndromes may exist. Especially in paraneoplastic sensory neuropathy and paraneoplastic encephalomyelitis, the most common cancer is small cell lung carcinoma and it is associated with anti-Hu antibody.4–8 At high titres, it is a sensitive and specific marker for paraneoplastic sensory neuropathy and paraneoplastic encephalomyelitis with small cell lung cancer. We describe a patient with sensory neuropathy and intraducal spreading breast cancer. We detected an IgG antibody in her serum which reacted with mouse neural tissue. This seems to be a new antibody which is different from the known antineuronal antibodies, and is important in the pathogenesis of sensory neuropathy.

Case report
A 59 year old woman became aware of numbness in her left arm in 1991. After one month, she noticed difficulty in moving the left arm. In 1992, numbness and dysesthesia developed in her left leg. In October 1993, she began to have difficulty ascending and descending stairs. In January, 1994, she noticed numbness in her right hand fingers. She was admitted to Shinshu University hospital on 6 October 1994.

On admission, routine physical examination was unremarkable with the exception of a left axillary lymphadenopathy. Neurological examination disclosed pseudoathetosis in her left hand fingers. Muscle strength was almost normal, but movements of the left hand were ataxic. Tendon reflexes were absent in the limbs. Babinski’s sign was elicited bilaterally. Pinprick and touch sensation were decreased in the areas described by the patient as numb. Vibratory sensation was decreased in her limbs and position sensation was lost in her left handfingers. Laboratory findings on admission were as follows. No abnormal values were found in blood and chemistry tests. A test for M protein was negative. The examination of autoantibodies as a screening for autoimmune diseases was normal with all tumour markers within the normal range. Examination of CSF showed 1 lymphocyte/mm³ and a total protein value of 43 mg/dl with 5.1% IgG. Electromyography, motor nerve conduction velocity, and F waves were all normal. She displayed no clinical or
electrophysiological evidence of motor nerve involvement. Sensory nerve conduction velocity was not detected in ulnar and tibial nerves. The chronic onset of illness associated with sensory neuropathy and the axillary lymphadenopathy in this patient were suggestive of sensory neuropathy with malignancy of unknown origin. A biopsy of the left axillary lymph node disclosed metastatic adenocarcinoma. Repeated examinations failed to disclose the primary adenocarcinoma lesion. Immunohistological study using antithyroglobulin antibody, anti amyloid antibody, anti-CA19-9 antibody, anti-CA15-3 (DF3, 115D8), and BCA225 suggested that metastatic adenocarcinoma of the left axillary lymph node originated from a breast cancer. On 16 December 1994, she underwent left radical mastectomy. Histological examination of a surgically obtained specimen showed intraductal spreading type breast cancer. Sural nerve biopsy was performed at the time of mastectomy, and showed pronounced axonal degeneration but no neoplastic cell infiltration. The patient was discharged in good health in January 1995, but sensory neuropathy remained. After surgery, her sensory neuropathy did not progress.

Methods

IMMUNOHISTOCHEMICAL IDENTIFICATION OF ANTI NEURAL ANTIBODIES IN THE PATIENT’S SERUM.

Immunohistochemical studies were performed on frozen acetone fixed tissue sections using immunoperoxidase staining. Under deep anaesthesia, a mouse was perfused and the brain and spinal cord were removed. Frozen sections 4 µm thick were prepared by cryostat from coronal slices of cerebellum and spinal cord from the mouse, and from the patient’s left axillary lymph node. The sections were incubated with the serum from the patient or a normal healthy subject, diluted 1:2000 in 1% blotto (non-fat dry milk), for two hours at room temperature. After washing with phosphate buffered saline (PBS) the sections were incubated with biotinilated goat antihuman IgG antibody (KPL Inc), diluted 1:400 in 1% blotto, for two hours at room temperature. After washing with PBS, they were reacted by an ABC method using histostain-DS kit (Zymed Lab Inc, San Francisco, CA, USA).

WESTERN BLOT ANALYSIS

IgG antibodies against a 106 kDa antigen in mouse cerebellum were detected in the serum of the patient (figure 2). Conversely, serum samples from the control did not react. Anti-Hu antibody was not detected in the serum of the patient or the control. The patient’s serum did not react with her metastatic lymph node. Normal control serum did not react with the 106 kDa protein in either cerebellum or lymph node. We also performed western blot analysis using mouse liver, lung, kidney, and skeletal muscle. No 106 kDa antigen was found.

Results

IMMUNOHISTOCHEMICAL IDENTIFICATION OF ANTI NEURAL ANTIBODY

Immunohistochemistry analysis of serum samples from the patient showed staining of the molecular layer and Purkinje cell layer of the cerebellar cortex as detected by light microscopy. In the spinal cord, the posterior grey horn was stained (figure 1). These findings could not be detected in samples from the normal control subject.

Figure 1 Immunohistochemical staining of frozen sectioned mouse spinal cord. Serum samples were diluted 1:2000 in 1% blotto. The posterior grey horn of the spinal cord was stained.
neural antibodies have been identified. Anti-neurological paraneoplastic syndrome, several breast cancers have also been reported. Of paraneoplastic sensory neuropathy with the Hu antigen in neuronal nuclei. Some cases and it is associated with autoantibodies against small cell lung cancer, the most common cancer with paraneoplastic sensory neuropathy may be caused by immune mediated mechanisms. Autoantibodies have been detected in the serum of many patients, with various paraneoplastic neurological diseases, using immunohistochemistry and immunoblotting. In neurological paraneoplastic syndrome, several neural antibodies have been identified. Anti-Purkinje cell autoantibody, so-called “Yo” antibody, has been a marker of gynaecological and breast carcinoma in the context of subacute cerebellar degeneration. Antineuronal nuclear antibodies, which are called “Hu” and “Ri”, were found in patients with small cell lung cancer and associated sensory neuropathy and encephalomyeloradiculopathies. Paraneoplastic sensory neuropathy is often associated with anti-Hu antibody and small cell lung cancer.

Although already known antineural antibodies associated with neurological paraneoplastic syndrome, such as anti-Hu, Ri, or Yo antibody, were negative in our patient, we found an IgG antibody that reacted against mouse neural tissue in the patient’s serum. Immunohistochemical study disclosed that IgG antibody was reacting with the molecular layer and Purkinje layer of cerebellum and dorsal roots and posterior grey horn of the spinal cord. Western blot analysis showed that an antibody was directed against a 106 kDa mouse neural protein. It is very difficult to prove that the antibody that gives the 106 kDa band in immunoblots of mouse cerebellum is the same that gives immunostaining in cerebellum or spinal cord. However, there is a possibility that the antibody that gives the 106 kDa band in immunoblotted mouse neural tissue may be the same as that which gives immunostaining in cerebellum or spinal cord, although there is also a possibility that additional antibodies against conformational epitopes would have been missed by immunoblotting and be the ones that immunoreact with cerebellum and spinal cord.

Characteristic features of the pathology of paraneoplastic sensory neuropathy are degeneration of the dorsal root ganglion neurons and mononuclear infiltration of the dorsal root ganglia. Secondary Wallerian degeneration of peripheral sensory nerves, posterior nerve roots, and posterior columns of the spinal cord is evident. Although the relation between the antibody we described and sensory neuropathy is not clear, immunohistochemical study disclosed that this antibody seems to be related to paraneoplastic sensory neuropathy and may play an important part in the pathogenesis of this neuropathy.

The role of antineural antibodies in the pathogenesis of paraneoplastic sensory neuropathy is unknown. Some reports assert that paraneoplastic sensory neuropathy results from an autoimmune cross reaction between a tumour cell antigen and brain nucleoprotein. On the other hand, antibodies against neural tissue are also detected in the serum of normal subjects. Sillevis et al reported that immunisation of the recombinant Hu fusion protein did not cause neurological disease in mice even though they had a high titre of anti-Hu antibodies. Taken together, these findings suggest that antibodies which cross react with a tumour cell antigen and neural tissues do not directly result in the destruction and degeneration of the neural tissues. In this study, we have shown an antineural antibody directed against a 106 kDa neural protein that does not react with the metastatic lymph node. Our results do not support the hypothesis that the paraneoplastic sensory neuropathy of the patients is directly caused by an antibody that cross reacts with a tumour cell antigen and a neural antigen. However, this antibody may play an important part in the pathogenesis of neurological paraneoplastic syndrome. The role of antineural antibody should be clarified. In so far as we are aware, this is the first report of this antineural antibody in the serum of a patient with breast cancer and sensory neuropathy. It is possible that the antibody that we detected is a new antineural antibody. Further study is needed to clarify the specificity and sensitivity of this antibody to paraneoplastic sensory neuropathy or breast carcinoma.

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