Frontal cortical synaptophysin in Lewy body diseases: relation to Alzheimer’s disease and dementia

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Abstract

Objectives—Dementia in Alzheimer’s disease correlates closely with loss of neocortical synapses. Similar synaptic loss has been shown in patients whose Alzheimer’s disease is also associated with neocortical and brain stem Lewy bodies. The aim was to determine if dementia in Lewy body disease was associated with diminished concentrations of midfrontal cortex synaptophysin.

Methods—An immunobinding assay was used to measure synaptophysin in post-mortem samples of midfrontal cortex from 89 patients with Alzheimer’s disease (ages 59–100, mean 79), 22 with combined Lewy body disease and Alzheimer’s disease (ages 69–103, mean 79), 15 demented patients with “pure” Lewy body disease (ages 57–80, mean 74), nine with neocortical and brain stem Lewy bodies who had Parkinson’s disease but were not demented (ages 68–85, mean 79), and 20 neurologically normal controls (ages 58–89, mean 75). The diagnosis was confirmed in all cases by detailed neuropathological examination of the contralateral hemisphere. Seven of the patients in the pure Lewy body disease with dementia group had initially presented with parkinsonism and eight with dementia.

Results—Synaptophysin concentrations (arbitrary units (AU)/µg) in patients with Alzheimer’s disease (mean 79 (SD 28)) or combined Lewy body disease and Alzheimer’s disease (mean 83 (SD 33)) were significantly lower than in controls (mean 115 (SD 29)) (p=0.002). Synaptophysin concentrations in demented patients with pure Lewy body disease (mean 106 SD 39) and patients with Lewy body disease who were not demented (mean 101 (SD 18)) did not differ significantly from control values or from each other.

Conclusion—Loss of midfrontal cortex synapses probably contributes to dementia in Lewy body disease when Alzheimer’s disease is also present but not to the dementia of pure Lewy body disease.
Mean (SD) concentrations of synaptophysin

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Mean age (years)</th>
<th>Mean PM delay (h)</th>
<th>Mean synaptophysin (AU/µg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LBD without dementia</td>
<td>9</td>
<td>79 (5)</td>
<td>15 (14)</td>
<td>101 (18)</td>
</tr>
<tr>
<td>LBD with dementia</td>
<td>15</td>
<td>74 (7)</td>
<td>26 (17)</td>
<td>106 (39)</td>
</tr>
<tr>
<td>Mixed LBD + AD</td>
<td>22</td>
<td>79 (8)</td>
<td>11 (10)</td>
<td>83 (33)</td>
</tr>
<tr>
<td>AD</td>
<td>89</td>
<td>79 (9)</td>
<td>NA</td>
<td>79 (28)</td>
</tr>
<tr>
<td>Controls</td>
<td>20</td>
<td>75 (8)</td>
<td>42 (35)</td>
<td>115 (29)</td>
</tr>
</tbody>
</table>

LBD = Lewy body disease; AD = Alzheimer’s disease; NA = not available, but < 24 hours; PM = postmortem.

The non-demented Lewy body disease group consisted of nine patients with clinically diagnosed idiopathic Parkinson’s disease (age range 68–85, mean 74), whose brains showed only typical Parkinson’s disease pathology, including a few neocortical Lewy bodies in every instance, without Alzheimer’s disease changes. The DLB group included 37 patients, both those whose initial and primary clinical diagnosis was dementia, and patients with Parkinson’s disease who developed dementia before death. These 37 DLB specimens were separated into those with and those without concomitant Alzheimer’s disease; a neuropathological diagnosis of Alzheimer’s disease was assigned if a brain had enough senile plaques to meet both National Institute on Aging criteria for Alzheimer’s disease4 and criteria from the Consortium to Establish a Registry for Alzheimer Disease for “definite”, or “probable” Alzheimer’s disease.5 There were 22 such mixed Lewy body disease and Alzheimer’s disease brains. All 22 of these patients (age range 69–103, mean 79) had presented with dementia, not Parkinson’s disease. The remaining 15 DLB cases (age range 57–80, mean 74) did not meet the above described neuropathological criteria for Alzheimer’s disease. Seven of these patients had presented initially with Parkinson’s disease, and eight with dementia. The Alzheimer’s disease comparison group consisted of 89 patients with pure Alzheimer’s disease (age range 59–100, mean 79). Finally, we also analysed synaptophysin concentrations from 20 age matched non-demented controls (age range 58–89, mean 75).

NEUROPATHOLOGICAL EVALUATION
At necropsy, the brains were divided midsagittally, and the right hemibrain was frozen at −70°C for neurochemical analysis, and the left was fixed in formalin for 10 days to three weeks before dissection. Standard neuropathological evaluation was performed on sections of frontal, temporal, and parietal lobes, hippocampus, entorhinal cortex, basal ganglia, substantia innominata, mesencephalon, and rostral pons. Neuronal loss, gliosis, and Lewy bodies were identified in the pigmented nuclei of the brain stem and nucleus basalis of Meynert in haematoxylin and eosin stained sections. All brains with Lewy bodies in the brain stem and nucleus basalis of Meynert included at least some neocortical Lewy bodies, predominately distributed in the limbic and entorhinal cortices and demonstrable in sections stained with haematoxylin and eosin and immunohistochemically for ubiquitin. The extent of Alzheimer’s disease pathology was evaluated using either thioflavin-S preparations viewed under ultraviolet illumination, or with modified Bielschowsky silver impregnation for plaques and tangles.

MIDFRONTAL CORTEX SYNAPTOPHYSIN QUANTIFICATION
Specimens from the frozen right hemibrain midfrontal cortex were analysed with a dot immunobinding assay for quantification of synaptophysin-like immunoreactivity. Post-mortem intervals before freezing of the brain tissue varied considerably, from two to 103 hours, but we have shown this method of synaptophysin measurement to be stable over prolonged postmortem intervals,16 and the controls had both the longest average postmortem delay interval (42 hours) and the highest mean synaptophysin concentrations (115 arbitrary units (AU)/µg). Determination of neocortical synaptophysin was undertaken by homogenising 0.1 g frozen brain tissue in 1 ml cold homogenising buffer with a Dounce glass/glass tissue grinder. A low speed spin (5000 ×g for 10 minutes) was performed and the supernatant separated into particulate and cytosolic fractions by ultracentrifugation (35 000 ×g for 4°C). After resuspendling pellet with buffer, 2µg particulate fraction was blotted on to nitrocellulose membrane using a dot-blot manifold. A serial dilution of aliquoted control sample was included as a normalisation standard. Blots were blocked using phosphate buffered saline (PBS) and 0.1% Tween-20 for two hours. They were then incubated in primary antibody (antisynaptophysin, Boehhringer-Mannheim Biochemical, Indianapolis, IN, USA, mouse monoclonal, 0.1 µg/ml in PBS-BSA) for 12–18 hours, followed by rabbit anti-mouse IgG (0.85 µg/ml in PBS-BSA), and 125I-protein A (0.1 µCi/ml in PBS-BSA) for two hours each. After washing in PBS-Tween, an autoradiogram was produced on a phosphor imaging screen and quantified on a Molecular Dynamics phosphorImager SE using the ImageQuant Software (Molecular Dynamics, Sunnyvale, CA, USA). Results are expressed as arbitrary units (AU)/µg blotted protein after normalising against the standard curve.

Results
The table shows the result of midfrontal cortex synaptophysin assay in each group. Unpaired two tailed t tests were used for the statistical analyses of the differences in midfrontal cortex synaptophysin concentrations. p Values<0.01 were regarded as statistically significant. Midfrontal cortex synaptophysin concentrations in patients with Alzheimer’s disease, or combined Lewy body disease and Alzheimer’s disease were significantly lower than in controls (p=0.001 and p=0.002, respectively). Further statistical analysis by one way analysis of variance (ANOVA) showed that mean synaptophysin values for the four groups differed significantly (F (3,62)=3.62, p<0.02). Post hoc comparisons by the least squares method showed that the combined Lewy body disease and Alzheimer’s disease groups had a
Frontal cortical synaptophysin in Lewy body disease

significantly lower synaptophysin value than controls (p<0.003). Synaptophysin concentrations in demented patients with only Lewy body disease and patients with Lewy body disease who were not demented did not differ significantly from control values or from one another.

Discussion

The results indicate that dementia in Lewy body disease is not associated with loss of midfrontal synaptophysin unless there is concomitant Alzheimer’s disease pathology. Synaptic loss in the midfrontal cortex cannot, therefore, serve as a unitary explanation for dementia across the clinical range of Lewy body diseases. These findings confirm those of a previous investigation into synaptic loss in the frontal cortex in DLB, which had employed far fewer cases, and they extend the finding to include patients who present initially with Parkinson’s disease and subsequently become demented. Samuel et al reported that brains showing mixed Lewy body disease and Alzheimer’s disease (LBV in their nomenclature) had significantly lower midfrontal cortex synaptophysin concentrations than had specimens from demented patients with only Lewy body disease (referred to by the authors as diffuse Lewy body disease, DLBD). Midfrontal cortical synapse loss in LBV was similar to that seen in Alzheimer’s disease, whereas synaptophysin concentrations in DLBD did not differ from controls.

Our results imply that dementia in Lewy body disease is due to neuropathological changes other than synaptic loss in the midfrontal cortex. Modified Braak stages of Alzheimer’s disease neurofibrillary pathology and neocortical neuritic plaque counts correlate with dementia in DLB, and increasing Braak stages are also associated with worsening cognition in combined Parkinson’s disease and Alzheimer’s disease. It is not surprising that progressively increasing burdens of Alzheimer’s disease pathology correlate with dementia, when such lesions are present in many patients with Lewy body disease. Neuropathological overlap of Alzheimer’s disease and Lewy body disease is well established, both neuropathologically and genetically. Lewy bodies have been found in some cases of familial Alzheimer’s disease due to a codon 717 mutation in the amyloid precursor protein gene. Cortical Lewy bodies have also been reported in patients with Down’s syndrome, who invariably develop Alzheimer’s disease if they live long enough. Finally, brains showing Lewy body disease with concomitant Alzheimer’s disease plaques and entorhinal cortex neurofibrillary tangles, whether termed Alzheimer’s disease plus Parkinson’s disease or LBV, have increased frequencies of the apolipoprotein E4 allele, similar to those encountered in Alzheimer’s disease. In patients with pure Lewy body disease, whether or not associated with dementia, the frequency of the apolipoprotein E4 allele is similar to that in the normal population. Concomitant Alzheimer’s disease cannot, however, explain impaired cognition in neuropathologically pure DLB or Parkinson’s disease. Neuropsychological comparisons of DLBD and LBV show significantly less dementia in patients with DLBD, but average mini mental state examination scores of 17 and Blessed scores of 13 among the patients with DLBD clearly establish the presence of disabling mental impairment without Alzheimer’s disease changes.

Neocortical choline acetyltransferase (ChAT) is strikingly diminished in DLB, far more so than in Alzheimer’s disease, which nevertheless shows significant reductions compared with controls. Patients with Parkinson’s disease, ChAT reductions in the temporal neocortex correlate with the degree of mental impairment assessed by a test of memory and information, and with neuronal counts in the nucleus of Meynert. Depletion of ChAT in the midfrontal cortex has also been shown to correlate with severity of dementia in DLB, and similar results have been reported in the pathologically identical condition termed senile dementia of the Lewy body type.

Neocortical Lewy body counts correlate with dementia in DLB, LBV, and DLBD. As neocortical Lewy bodies are generally, although not invariably, more frequent in DLB than Parkinson’s disease, their appearance and proliferation may constitute an additional explanation for the emergence of dementia in patients with Parkinson’s disease without concomitant Alzheimer’s disease pathology.

Damage to dopaminergic frontostriatal neurochemical mechanisms has been implicated as a possible neuropathological substrate for dementia in DLB and Parkinson’s disease with dementia. A recent analysis of concentrations of homovanillic acid in CSF found significantly lower concentrations in LBV than in Alzheimer’s disease. The authors concluded that Lewy bodies and mesolimbic and mesocortical regions may be partly responsible for the reduced dopamine metabolism and turnover reflected in low CSF concentrations of homovanillic acid, and that this may be related to the frequency of psychotic symptoms in this disorder. Brains from patients with Parkinson’s disease also show much lower amounts of dopamine and its metabolites in the hippocampus, and entorhinal, cingulate, and frontal cortex than do controls, and such dopaminergic deficiency may contribute to mental impairment in Parkinson’s disease. Depletion of other subcortical projecting systems in Lewy body diseases could also impair cognition. In particular, decreased concentrations of serotonin and noradrenaline, secondary to neuronal loss in the locus ceruleus and raphe nuclei, may contribute to dementia in DLB and Parkinson’s disease.

Our finding that neocortical synaptic loss does not contribute to dementia in DLB or Parkinson’s disease, unless there is concomitant Alzheimer’s disease pathology, applies only to the midfrontal cortex. Although neocortical Lewy bodies do occur in the frontal lobe in DLB and Parkinson’s disease, they are more numerous in the limbic cortex, and so synaptic loss in the entorhinal cortex, cingulate gyrus, or
insular may yet prove to impair cognition in Lewy body diseases. The data available at present, however, imply that concomitant Alzheimer's disease pathology in the form of neuritic plaques, higher Braak stages, and midfrontal cortex synaptic loss worsens the moderately severe dementia encountered in neuropathologically pure Lewy body diseases. In the absence of Alzheimer's disease changes, dementia in Lewy body diseases seems to be associated with a complex combination of neocortical Lewy body formation, loss of neocortical ChAT and dopamine, and progressive neurodegeneration in multiple cortically projecting brain stem nuclei.

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J Neurol Neurosurg Psychiatry 1998 64: 653-656
doi: 10.1136/jnnp.64.5.653

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