Relation between trinucleotide GAA repeat length and sensory neuropathy in Friedreich’s ataxia

L Santoro, G De Michele, A Perretti, C Crisci, S Cocozza, F Cavalcanti, M Ragno, A Monticelli, A Filla, G Caruso

Abstract

Objective—To verify if GAA expansion size in Friedreich’s ataxia could account for the severity of sensory neuropathy.

Methods—Retrospective study of 56 patients with Friedreich’s ataxia selected according to homozygosity for GAA expansion and availability of electrophysiological findings. Orthodromic sensory conduction velocity in the median nerve was available in all patients and that of the tibial nerve in 46 of them. Data of sural nerve biopsy and of a morphometric analysis were available in 12 of the selected patients. The sensory action potential amplitude at the wrist (wSAP) and at the medial malleolus (m mal SAP) and the percentage of myelinated fibres with diameter larger than 7, 9, and 11 µm in the sural nerve were correlated with disease duration and GAA expansion size on the shorter (GAA1) and larger (GAA2) expanded allele in each pair. Pearson’s correlation test and stepwise multiple regression were used for statistical analysis.

Results—A significant inverse correlation between GAA1 size and wSAP, m mal SAP, and percentage of myelinated fibres was found. Stepwise multiple regression showed that GAA1 size significantly affects electrophysiological and morphometric data, whereas duration of disease has no effect.

Conclusion—The data suggest that the severity of the sensory neuropathy is probably genetically determined and that it is not progressive.

Keywords: Friedreich’s ataxia, sensory neuropathy, GAA expansion

Friedreich’s ataxia is the most frequent early onset autosomal recessive inherited ataxia. The disease is clinically characterised by progressive ataxia with onset before 20 years of age, absence of lower limb tendon reflexes, dysarthria, Babinski’s sign, limb weakness, decreased vibration sense, and skeletal deformities. Cardiomyopathy and diabetes may also be found.1 Electrophysiological and pathological studies have suggested that axon degeneration and secondary demyelination constantly occur in peripheral sensory nerves.2,4

The gene causing the disease (X25) has been recently identified.7 X25 encodes a 210 amino acid protein with unknown function called “frataxin”. Most patients (90%) were homozygous for an unstable GAA trinucleotide expansion in the first intron of X25. Normal chromosomes contained 7–22 GAA units, whereas Friedreich’s ataxia chromosomes carried 200 to above 900 repeats. The remaining patients were heterozygous for the expansion and a few point mutations have been described in some of them. Advances in molecular genetics broadened the disease phenotype. In fact, the form with onset later than 20 years (late onset Friedreich’s ataxia),7 and that with retained tendon reflexes are in linkage with the FRDA locus. Moreover, we recently described patients with minimal GAA expansion on one allele (ranging from 120 to 156 triplets) and without clinical and electrophysiological signs of sensory neuropathy.9

A clear relation between expansion size and phenotype variability has also been found. The expansion size inversely correlates with age at onset and directly with the presence of diabetes mellitus and cardiomyopathy. The best correlation was found with the size of the shorter allele in each pair.4,12

To verify that GAA expansion size could account for the severity of sensory neuropathy, we studied the relation between molecular data and peripheral nerve electrophysiological and pathological findings. We also investigated if disease duration might affect the severity of peripheral neuropathy.

Patients and methods

Fifty six patients were selected among 160 patients with Friedreich’s ataxia examined since 1973 at the Department of Neurology of the Federico II University of Naples. The criteria of selection were homozygosity for GAA expansion and availability of electrophysiological findings. Forty three patients had typical Friedreich’s ataxia, 11 had onset after 20 years of age, six had preserved tendon reflexes, four of them both late onset and pre-
Table 1  Clinical, electrophysiological, and molecular findings

<table>
<thead>
<tr>
<th>Patients</th>
<th>Age</th>
<th>Disease duration</th>
<th>Wrist SAP amplitude</th>
<th>m mal SAP amplitude</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>µV</td>
<td>%</td>
</tr>
<tr>
<td>Electrophysiological study</td>
<td>56</td>
<td>23.8 (13.7)</td>
<td>8.3 (9.2)</td>
<td>1.32 (1.62)</td>
</tr>
<tr>
<td></td>
<td>12</td>
<td>28.7 (12.6)</td>
<td>8.7 (6.9)</td>
<td>1.46 (1.64)</td>
</tr>
</tbody>
</table>

Values are mean (SD); SAP=sensory action potential; m mal=medial malleolus; control values: n=68; age range=15–44 years; wrist SAP mean=14.3 µV (range 6–16); m mal SAP mean=2.16 µV (range 0.2–3).

Table 2  Pearson’s regression coefficients

<table>
<thead>
<tr>
<th>Wrist SAP *</th>
<th>m. mal. SAP *</th>
<th>Percent of fibres</th>
</tr>
</thead>
<tbody>
<tr>
<td>µV</td>
<td>%</td>
<td>µV</td>
</tr>
</tbody>
</table>

Pearson’s regression coefficient:
GAA1 size -0.639*** -0.660*** -0.353* -0.570*** -0.872*** -0.802** -0.856***
GAA2 size -0.027 -0.023 -0.095 -0.066 -0.342 0.129 0.057
Disease duration 0.214 0.257 0.098 0.279 0.779** 0.672* 0.690*

*p<0.05; **p<0.01; ***p<0.001.
SAP=sensory action potential; m mal=medial malleolus; control values: n=68; age range=15–44 years; wrist SAP mean=14.3 µV (range 6–16); m mal SAP mean=2.16 µV (range 0.2–3).

Table 3  Stepwise multiple regression: partial regression coefficients (SE)

<table>
<thead>
<tr>
<th>Wrist SAP</th>
<th>m mal SAP</th>
</tr>
</thead>
<tbody>
<tr>
<td>µV</td>
<td>%</td>
</tr>
</tbody>
</table>

Stepwise multiple regression: partial regression coefficients (SE)
GAA1 size -0.0051 (0.0080)*** -0.0503 (0.0071)*** -0.0006 (0.0003)* -0.0695 (0.0152)***
GAA2 size 0.0029 (0.0012)* 0.0301 (0.0113)* 0.0001 (0.0005) 0.0378 (0.0248)
Disease duration 0.0249 (0.0180) 0.3139 (0.1654) 0.0013 (0.0064) 0.5417 (0.3542)

*p<0.05; **p<0.01; ***p<0.001.
SAP=sensory action potential; m mal=medial malleolus; control values: n=68; age range=15–44 years; wrist SAP mean=14.3 µV (range 6–16); m mal SAP mean=2.16 µV (range 0.2–3).
Table 3 (continued)

<table>
<thead>
<tr>
<th>Per cent of fibres</th>
<th>&gt;7 µ</th>
<th>&gt;9 µ</th>
<th>&gt;11 µ</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>−0.0338 (0.0108)***</td>
<td>−0.0241 (0.0083)**</td>
<td>−0.0141 (0.0038)***</td>
<td></td>
</tr>
<tr>
<td>0.0106 (0.0100)</td>
<td>0.0208 (0.0077)</td>
<td>0.0086 (0.0036)</td>
<td></td>
</tr>
<tr>
<td>0.7176 (0.3993)</td>
<td>0.6007 (0.3063)</td>
<td>0.2488 (0.1421)</td>
<td></td>
</tr>
</tbody>
</table>

Trinucleotide GAA repeat length and sensory neuropathy in Friedreich's ataxia

The continuous line and the full symbols refer to the correlation between the number of GAA1 repeats and sensory action potential at the wrist (wSAP) in 56 patients (r =−0.639; p< 0.001). The dashed line and the open symbols refer to the correlation between the number of GAA1 repeats and percentage of sural nerve fibres>7 µm in 12 patients (r =−0.872; p< 0.001).

Discussion

Friedreich’s ataxia is the first autosomal recessive disease due to an expanded trinucleotide repeat.1 The discovery of the Friedreich’s ataxia molecular defect has implications for the diagnostic criteria and for the explanation of the phenotypic variability of the disease. Filla et al2 showed that patients with late onset ataxia and preserved reflexes are homozygous for the GAA expansion, confirming the genetic homogeneity between typical Friedreich’s ataxia and the atypical forms. The recent finding of patients homozygous for the expansion and with no signs of peripheral neuropathy9 prompted us to investigate the relation between the sensory neuropathy severity, as expressed by SAP amplitude, and the expansion size.

Loss of large peripheral nerve fibres is considered to be constant in Friedreich’s ataxia,2–4 but there is no agreement about the pathogenesis of axon loss. The severity of myelinated fibre reduction seemed to increase with the patients’ age,15 but it was not related to disease duration or severity.1 Said et al16 suggested a slow dying back degeneration of abnormally developed sensory neurons. We performed sural nerve biopsies and serial sensory nerve conduction studies in 15 patients with Friedreich’s ataxia.15 In three of them a contralateral nerve biopsy was repeated after 6–7 years. Typical electrophysiological findings did not change with time, suggesting a very early but stable involvement of peripheral nerve, due to a defective development of the largest sensory neurons. Goto and Hirano16 suggested a selective involvement of these neurons by using immunohistochemical staining for substance P and synaptophysin. Frataxin expression is high in dorsal root ganglia (M Pandolfo, personal communication).

The present study shows a clear relation between expansion size and severity of sensory neuropathy. In fact, we showed an inverse correlation between the number of GAA1 repeats and neurophysiological (wSAP and m mal SAP amplitude) and morphological findings (percentage of the largest fibres in the sural nerve). GAA1 size is the main factor determining severity of sensory neuropathy as also shown by multiple regression analysis. It accounts for 41% of the variation of wSAP amplitude and a difference in GAA1 size of 100 triplets determines a mean change of about 0.5 µV in wSAP. The results of the analysis for sural nerve biopsy data were even more evident. GAA1 size accounts for 76% of variation of fibres >7 µm. A difference in GAA1 size of 100 triplets determines a mean change of about 3.4% in fibre percentage.

The size of GAA2 is correlated neither with neurophysiological nor with morphological findings. However, multiple regression analysis showed a GAA2 effect on wSAP amplitude with higher amplitude values for higher GAA2 sizes. This effect, marginally significant (p<0.05), and biologically unlikely, may be due to chance.

In the group of patients with available peripheral nerve morphological findings, the number of remaining large fibres was inversely correlated with the size of GAA1 expansion and directly with duration of disease. An explanation of the last finding is that patients with lower GAA size had milder disease progression and underwent sural nerve biopsy later. There was a high inverse correlation between disease duration at biopsy and the number of repeats on GAA1 allele (r =0.697; p<0.05), and multiple regression analysis showed that duration of disease is not a significant determinant of the loss of large fibres. A trend towards a direct correlation between dis-
Severe sensory neuropathy has been considered for a long time a hallmark of electrophysiology of patients with Friedreich's ataxia; however, the molecular diagnosis allowed us and others to detect a few patients with Friedreich's ataxia without neuropathy. The present results show that very small expansions can be associated with very mild or absent sensory neuropathy, as measured by SAP amplitude. On the other hand, patients with an expansion above 600 GAA repeats have a full clinical expression of the involvement of peripheral nerves. This threshold is close to the 700 GAA repeats indicated by Dürr et al as the level above which the residual expression is too low to influence the clinical presentation. Our data suggest that the severity of the sensory neuropathy is probably genetically determined and that it is not progressive. Tissue mosaicism caused by mitotic instability, as has been shown in several triplet diseases including Friedreich's ataxia, might be a possible further mechanism that could explain the variability of severity of sensory neuropathy in patients with the disease. Studies on X25 transcription and translation in the peripheral nervous system will elucidate this issue.

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