Antiganglioside antibodies in Guillain-Barré syndrome after a recent cytomegalovirus infection

Azadeh Khalili-Shirazi, Norman Gregson, Ian Gray, Jeremy Rees, John Winer, Richard Hughes

Abstract
Objective—To study the association between anti-ganglioside antibody responses and Guillain-Barré syndrome (GBS) after a recent cytomegalovirus (CMV) infection.

Methods—Enzyme linked immunosorbent assay (ELISA) was undertaken on serum samples from 14 patients with GBS with recent cytomegalovirus (CMV) infection (CMV+GBS) and 12 without (CMV-GBS), 17 patients with other neurological diseases (OND), 11 patients with a recent CMV infection but without neurological involvement, 11 patients with recent Epstein-Barr virus (EBV) infection but without neurological involvement, and 20 normal control (NC) subjects.

Results—IgM antibodies were found at 1:100 serum dilution to gangliosides GM2 (six of 14 patients), GM1 (four of 14), GD1a (three of 14) and GD1b (two of 14) in the serum samples of the CMV+GBS patients, but not in those of any of the CMV-GBS patients. IgM antibodies were also found to gangliosides GM1, GD1a, and GD1b in one of 11 OND patients, to ganglioside GM1 in one of 11 non-neurological CMV patients, and to ganglioside GD1b in one of 20 NC subjects. Some patients with EBV infection had IgM antibodies to gangliosides GM1 (five of 11), GM2 (three of 11), and GD1a (two of 11). However, the antibodies to ganglioside GM2 had a low titre, none being positive at 1:200 dilution, whereas five of the CMV+GBS serum samples remained positive at this dilution.

Conclusion—Antibodies to ganglioside GM2 are often associated with GBS after CMV infection, but their relevance is not known. It is unlikely that CMV infection and anti-ganglioside GM2 antibodies are solely responsible and an additional factor is required to elicit GBS.
Antiganglioside antibodies in Guillain-Barré syndrome after cytomegalovirus

Clinical details and the ELISA results of the serum samples of the GBS patients, with (CMV+GBS) and without (CMV-GBS) a recent CMV infection

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Values ( ) in the last two rows are mean (SD).

+ = positive (a value greater than the NC mean + 3 SD); = weak positive, ± = borderline, (±) = positive or negative by TLC; URTI = upper respiratory tract infection; ab = antibody; vac = vaccine.

Patient 2, but none of the others, had serological evidence of a recent Campylobacter jejuni infection.

An ELISA negative serum (patient 4) gave a positive band in TLC. This might be due to a difference in the epitope specificity of the antibody, but as this result was not confirmed by both methods, it was not regarded as positive.

Patient 8, but none of the others, had positive IgG antibodies to GM1 and GD1b.

2=anaesthesia or analgesia of fingers or feet, 3=anaesthesia or analgesia to elbows or knees or worse.

ANTIBODY ASSAYS

ELISA

Antibodies to CMV were determined by enzyme linked immunosorbent assay (ELISA) (Enzygnost Kit, Behring Diagnostics, Hounslow, Middlesex, UK) with CMV infected fibroblasts and peroxidase labelled antihuman IgM antibody as the detection system. We did not consider any samples as containing CMV antibodies if the optical density value was less than 0.20.

Antibodies against gangliosides were detected by ELISA with the following protocol. The wells of ELISA plates (Immulon 3, Dynatech, UK) were coated with 100 µl gangliosides GM1 (Sigma, Poole, UK), GM2, GD1a, or GD1b (Alexis, Nottingham, UK) at 1 µg/ml and cholesterol (5 µg/ml) in methanol, by evaporation. After washing the plates with high ionic phosphate buffered saline (PBS), non-specific binding was blocked with 1% bovine serum albumin (BSA) fraction V (Sigma, Poole, UK) in PBS (100 µl) for 2 hours at room temperature. One hundred microlitres of the test and control serum samples, appropriately diluted in 1% BSA PBS, were then incubated overnight at 4°C. The plates were washed with PBS, and then antihuman IgG or IgM conjugated to alkaline phosphatase (Sigma, Poole, UK) (100 µl) was added. The plates were further incubated for 2 hours at 4°C and developed by incubating for 1 hour at 37°C, with p-nitrophenol phosphate substrate tablets (Sigma) (100 µl). The absorbance was read at 405 nm. Serum samples were tested in triplicate and considered positive when the mean absorbance was more than 3 SD greater than the mean of the NC serum.

Thin layer chromatography with immuno-overlay

All of the ganglioside ELISA positive serum samples were tested by thin layer chromatography (TLC) with immuno-overlay. A mixture of 40 µl of each of the gangliosides GM1, GM2, GD1a, and GD1b, each at 1 mg/ml, was resolved on 85 mm of aluminium backed TLC plates (1.88 µg of gangliosides/mm) (Merck, Poole, Dorset, UK) in chloroform/methanol/0.05 M CaCl2 (30:20:4, v:v:v) at 4°C. Antibody binding was detected using standard techniques. Positive and negative control serum samples were included in each test.

STATISTICAL ANALYSIS

Multiple groups were compared with the Kruskal-Wallis analysis of variance (ANOVA) and groups of interest were compared with the normal control group with a Mann-Whitney test. Two tailed tests of significance are quoted. Differences in proportions were tested with Fisher's exact test.

Results

ANTI-CMV ANTIBODIES

Anti-CMV IgM antibodies were present in 14 patients with GBS and absent in 12. There was no difference in the sex ratio, age, clinical features, delay between the collection of serum samples and the onset of GBS for the two groups (table). The mean optical density obtained for IgM anti-CMV antibody ELISA for the CMV+GBS group was 0.52 (SD 0.23) and for the CMV-GBS it was 0.03 (SD 0.05). The corresponding values for the patients with CMV infection but no neurological disease were 0.41 (SD 0.11) and for the EBV patients they were 0.07 (SD 0.10). The most frequent preceding illness in both GBS groups was an upper respiratory tract infection, which was reported by eight of 14 of the patients with GBS with CMV antibodies and four of 12 of those without.

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ANTI-GANGLIOSIDE ANTIBODIES

IgM antibody binding to one or more of the gangliosides was detected by ELISA in eight of 14, and IgG antibodies in one of 14 CMV+GBS patients, but not in any of the CMV-GBS patients (table). IgM antibodies were detected against gangliosides GM1 in four of 14, GM2 in six of 14, GD1a in three of 14, and GD1b in two of 14 patients in the CMV+GBS group. The optical density of ganglioside GM2 in the CMV+GBS group was significantly higher than those in the CMV-GBS (Mann Whitney test, p=0.0009), OND (p<0.0001), CMV (p=0.0034), and NC (p=0.0002) groups, but not significantly higher than those in the EBV group (p=0.1323).

Some of the antibodies were titrated, but all were tested at 1:100 and 1:200 dilutions, the results for which are shown as some low titre (positive at 1:100, but negative 1:200) antibodies.

ANTIGANGLIOSIDE ANTIBODIES

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Some of the antibodies were titrated, but all were tested at 1:100 and 1:200 dilutions, the results for which are shown as some low affinity antibodies become negative at 1:200 dilution. The highest anti-ganglioside GM2 IgM antibody binding occurred in patients from the CMV+GBS and the EBV groups. However, the EBV serum samples were all negative to ganglioside GM2 at 1:200 dilution, whereas five of 14 of the CMV+GBS samples were still positive at this dilution. None of the samples from the other groups contained significant concentrations of antibody to this ganglioside (figure). By contrast, IgG antibodies against gangliosides GM1 and GD1b were detected only in one of the CMV+GBS patients (results not shown). The ELISA results for individual patients showed that when antibodies were present, they were often against more than one ganglioside (table).

Compared with the NC subjects, the IgM (1:100 dilution) ELISA absorbance readings of CMV+GBS patients were significantly increased against gangliosides GM1 (p=0.001), GM2 (p=0.0001), GD1a (p=0.002) and GD1b (p=0.001). The only other group of patients with significantly raised concentrations was the EBV group, in which IgM anti-ganglioside binding was raised to gangliosides GM1 (p=0.001) and GD1a (p=0.007).

TLC IMMUNO-OVERLAY

Samples were considered positive, if they were confirmed both by ELISA and TLC. Serum
samples strongly positive by ELISA were all confirmed by TLC immuno-overlay, but some of the borderline ELISA positive samples did not bind on the TLC plates (table). In the serum from CMV+GBS patient 8, ELISA positive IgG antibody binding to gangliosides GM1 and GD1b was also confirmed by TLC immuno-overlay. The TLC immuno-overlay results for the EBV group confirmed some of the ELISA positive results for gangliosides GM1 (five of six), GM2 (three of three), and GD1a (two of three). It did not detect the two borderline ELISA positive serum samples.

Discussion

This study extends previous findings on the relation between CMV infection and anti-ganglioside antibodies in GBS. When our findings are combined with those of previous reports, a total of 22 of 44 CMV+GBS, nine of 187 CMV-GBS, two of 135 OND, seven of 71 CMV, three of 11 EBV patients, and none of the 140 in the NC group were positive to ganglioside GM2. It becomes clear that antibodies to ganglioside GM2 are present more commonly in GBS associated with recent CMV infection, than in patients with GBS not associated with CMV (p=0.0001; Fisher’s exact test). Low affinity antiganglioside GM2 IgM antibodies have been reported in serum samples from normal people and may represent some of the natural antibody repertoire, but they have also been reported in patients with peripheral and central nervous system disease.

Cytomegalovirus can produce both persistent and latent infection in humans, infecting various cell types including neural and endothelial cells. There is no histological evidence of CMV infection within the nerve in GBS, and a polymerase chain reaction study confirmed by TLC immuno-overlay, but some of the ELISA positive results for gangliosides GM1 (five of six), GM2 (three of three), and GD1a (two of three). It did not detect the two borderline ELISA positive serum samples.

anti-ganglioside GM2 antibodies is not known. Ganglioside GM2 is localised at the neuromuscular junction in the rat. The concentration of this ganglioside is increased in the plasma membrane of melanoma cell lines and neuroendocrine tumour cells and anti-ganglioside GM2 antibodies are cytotoxic to these cells. High titre antiganglioside GM2 antibodies are often reactive with other minor gangliosides such as IV4GalNAcGD1a and IV4GalNAcGM1b. Antibodies weakly reactive with ganglioside GM2, but strongly reactive with IV4GalNAcGD1a have been reported in a patient developing GBS after Campylobacter jejuni enteritis. A cross reactive shared sequence with minor gangliosides and ganglioside GM2 itself may be the relevant antigenic target in GBS. Further research into the fine specificity of antiganglioside antibodies generated after CMV and other infections is necessary to determine how these antibodies are involved in the pathogenesis of GBS.

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References

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