Biological activity of interferon betas in patients with multiple sclerosis is affected by treatment regimen and neutralising antibodies

A Bertolotto, A Sala, S Malucchi, F Marnetto, M Caldano, A Di Sapio, M Capobianco, F Gilli

Background: MxA gene expression is one of the most appropriate markers of biological activity of exogenous interferon (IFN) beta.

Methods: We quantified MxA mRNA for five consecutive days in 62 patients treated with IFN beta (16, Avonex; 10, Betaferon; 24, Rebif 22; 12, Rebif 44), by quantitative-competitive polymerase chain reaction. Ever three months, IFN beta induced neutralising antibodies (NAbs) were evaluated in sera using a cytopathic effect assay.

Results: Two categories of patients were identified: one group (49/62) had a sharp post-injection increase in MxA expression (defined as “IFN beta biological responder”), whereas the other group (13/62) had no MxA induction after IFN beta administrations (defined as “IFN beta biological non-responder”). In 11/13 biological non-responders, the persistent presence of NAbs correlated with abolished biological activity, independently of treatment regimen. The two remaining IFN beta biological non-responders were NAb−. Among the 49 IFN beta biological responders, biological activity was comparable between the four preparations on day 2 and 3 (+12 and +36 hours post-injection), but it was greater in Betaferon and both Rebif preparations on day 1, 4, and 5. In biological responders treated three times a week, only 82% (59/72) of injections were considered effective, compared with 100% (13/13) of Avonex injections.

Conclusion: Our results suggest that an optimal IFN beta regimen is not yet available: Avonex, given once a week, shows lower cumulative biological activity. On the other hand, both Betaferon and Rebif, given three times a week, show 18% biologically ineffective injections and higher risk of developing NAbs, which abolish biological activity.

PATIENTS AND METHODS

Patients

Blood samples were obtained from a total of 62 patients with MS (22 men and 40 women) who received treatment with recombinant IFN beta. Of these, 16 received IFN beta-1a (Avonex; Biogen, Cambridge, USA) 30 micrograms intramuscularly (IM) once a week, 36 received IFN beta-1a (Rebif; Serono, Basel, Switzerland), either 22 micrograms (n = 24) or 44 micrograms (n = 12) subcutaneously (SC) three times a week, and 10 received IFN beta-1b (Betaferon; Schering, Berlin, Germany) 250 micrograms SC three times a week. The mean duration of therapy was 20 (SD 17) months (range 3–60 months) (table 1).

Patients were not randomised and enrolled retrospectively. Eligibility criteria included a diagnosis of MS according to the McDonald criteria,22 relapsing–remitting (RR) clinical course, Expanded Disability Status Scale (EDSS) score 0–6.5, and informed consent. All patients included were clinically inactive and steroid free in the three months preceding the enrolment. Exclusion criteria included significant other medical illnesses, previous switch in type of IFN beta treatment, and prior immunosuppressive therapy with cytotoxic activity.

Study design

Eligible patients had been under treatment for at least three months prior to the study and had been screened for the

Abbreviations: AUC, area under the concentration time curve; EDSS, Expanded Disability Status Scale; IFN, interferon; MS, multiple sclerosis; MxA, myxovirus resistance protein A; NAbs, neutralising antibodies; OAS, oligoadenylate synthetase; PBMC, peripheral blood mononuclear cell; TRU, tenfold reduction unit
Quantification of MxA mRNA
MxA mRNA was quantified using our previously published protocol. Briefly, peripheral blood mononuclear cells (PBMCs) were separated on a Ficoll-Hypaque (Pharmacia, Uppsala, Sweden) density gradient and total RNA was extracted using RNAwiz reagent, following the manufacturer’s instructions (Ambion, Austin, TX). Complementary DNA (cDNA) was then prepared, using 10 mM of random hexamer primers (Perkin Elmer, Norwalk, CT) and 100 U of Moloney murine leukaemia virus reverse transcriptase (Gibco Laboratories, Grand Island, NY). For the qc-PCR reaction two competitor cDNA fragments (co-MxA and co-glyceraldehyde phosphate dehydrogenase (GAPDH)) were generated and co-amplified with target cDNA. PCR amplification products were then resolved following separation by 2% agarose gel electrophoresis. Bands were visualised by Etbr staining and quantified by densitometric scanning of the gel using a GelDoc 1000 UV fluorescent system (Bio-Rad, Richmond, CA).

The ratios between competitors and target cDNA were evaluated as ratios between bands values, taking as ratio = 1 an amount of starting targets (MxA or GAPDH) equal to the amount of each competitor. The MxA mRNA levels, expressed as fgMxA/pgGAPDH, were normalised using as housekeeping gene, to avoid differences due to expressed as fgMxA/pgGAPDH, were normalised using


densitometer at 620 nm. The neutralisation titre of a serum sample was calculated according to Kawade’s formula and expressed in tenfold reduction unit (TRU). A level of >20 TRU was considered as the threshold for positivity.

RESULTS
Patients
Table 1 shows the baseline demographic and clinical characteristics of the study patients. There were no significant differences between the groups with regard to demographic or clinical characteristics, except for the duration of treatment with Rebif 44, which was shorter compared with the other three treatment regimens (Rebif 22, p = 0.0051; Betaferon, p = 0.0318; Avonex, p = 0.0456). Most of the patients (65%) were women and the mean age was 35.4 years.

Neutralising antibody status
No patient was positive for NAb at baseline. Of the 12 persistent NAb+ patients, nine were evaluated as positive on day 1 and had mean NAb titres >45 TRU; two patients were negative on day 1 and had always had NAb titres <45 TRU in previous NAb tests. One patient was positive on day 1 and presented previous and present NAb titres <45 TRU.

The patients were not randomised, but retrospectively included, since the present study was not conducted to determine the incidence and prevalence of NAb in serum samples from patients with MS who were treated with Betaferon, Avonex, Rebif 22, or Rebif 44. Hence a direct comparison of the percentage of NAb+ and Nabs—patients reported in this study is not possible.
IFN beta biological responders and non-responders

In a previous report, we examined the MxA gene expression in PBMCs from 99 untreated patients with MS and calculated an upper threshold of normal as mean baseline expression+3 SD = 132 fgMxA/pgGAPDH.8

Two categories of patients treated with IFN beta were identified based on MxA mRNA levels after IFN beta administration: IFN beta biological responders had at least one MxA mRNA value higher than the established threshold (>132 fgMxA/pgGAPDH), and IFN beta biological non-responders had MxA mRNA values lower than 132 fgMxA/pgGAPDH during the whole study.

Based on the above threshold, 49/62 (79%) patients were IFN beta biological responders, whereas the remaining 13/62 (21%) patients were IFN beta biological non-responders as MxA expression was unaffected by IFN beta administration.

Among the biological responders, 13 were treated with Avonex (81% of the patients treated with Avonex), 7 were treated with Betaferon (70%), 19 were treated with Rebif 22 (79%), and 10 were treated with Rebif 44 (83%). Moreover, of the 49 IFN beta biological responders, 45 (92%) patients were NAb–, three patients were isolated NAb+, and one patient was persistent NAb+. However, the three isolated NAb+ and the single persistent NAb+ patient presented NAb titres <45 TRU during their follow up and were negative during the study.

Among the 13 IFN beta biological non-responders, 10 were found to be persistent NAb+ and positive during the study, one was persistent NAb+ and negative during the study, and two patients were NAb–.

MxA expression and neutralising antibody status

A comparison of MxA expression and NAb status showed that changes in MxA mRNA levels were greater in NAb– than in persistent NAb+ patients (fig 1). Such analysis was not possible for isolated NAb+, because they were too few in number.

Not unexpectedly, abolished MxA gene expression was more commonly found in patients with high NAb titres (>45 TRU) than in patients with low (<45 TRU) NAb titres: all patients (100%) with NAb titres >45 TRU showed no biological activity, as indicated by MxA mRNA levels <132 fgMxA/pgGAPDH during the whole study (fig 1). Of the three remaining persistent NAb+ patients with NAb titres <45 TRU, two subjects showed no biological activity, but one patient showed significant increases in MxA mRNA.

When the AUCs were compared, the total augmentation of MxA mRNA was fourfold greater in the NAb– group than in persistent NAb+ (p<0.0001).

Comparison of the biological activity of the four interferon beta preparations

Following the administration of IFN beta in biological responders, MxA mRNA concentrations peaked at 12 hours and then declined to baseline levels.15 As expected, MxA mRNA levels in patients given Avonex were lowest on day 1, peaked on day 2 (+12 hours after injection) and then decreased in the following days (fig 2A). No such decrease was seen with Betaferon or Rebif (Rebif 22 and Rebif 44) because, for both preparations, a booster was given three times a week rather than once a week. In particular, the average profiles of MxA expression, in patients treated with both Betaferon and Rebif, showed a second peak of expression on day 4, +12 hours after the second IFN beta injection (fig 2B–D).

MxA expression for both Betaferon and Rebif groups was statistically greater than that for the Avonex group on day 1 (all p<0.038), 4 (all p<0.045), and 5 (all p<0.044) (table 2). On the other hand, differences of MxA mRNA levels among the four preparations of IFN beta were not significant on day 2 and 3 (+12 and +36 hours after the first IFN beta injection) (all p>0.077) (table 2). Interestingly, no differences in the MxA expression were found between Rebif 22 and Rebif 44 during the five days of treatment (all p>0.24) (table 2).

There were no statistical differences among the MxA mRNA levels induced at each time point in persistent NAb+ patients treated with Avonex versus Betaferon, Rebif 22, and Rebif 44 (all p>0.12) (fig 2).

Although this study analysed only five treatment days and did not consider the effect of the third weekly injection of Betaferon and both Rebif preparations, the MxA mRNA AUC values approached statistical significance for both Rebif preparations and Betaferon versus Avonex (0.058>p>0.07).

Biologically effective injections

The profiles of the MxA concentration time curve were similar in all biological responders under treatment with Avonex. On the other hand, some subjects treated three times a week with either Betaferon or Rebif presented an unexpected profile, as MxA expression did not increase after one of the two injections considered. To examine this phenomenon, we evaluated the biological efficacy of every injection separately: injections were considered “biologically effective” when they induced MxA mRNA levels higher than the established threshold (132 fgMxA/pgGAPDH).

Among the 49 biological responders, 13 were treated once a week, and 36 were treated three times a week (7, Betaferon; 19, Rebif 22; 10, Rebif 44). Of these 36 patients, 13 (36%) presented a single “biologically effective” injection, instead of two: 2/7 (29%) were treated with Betaferon, 7/19 (37%) were treated with Rebif 22, and 4/10 (40%) were treated with Rebif 44. Ineffective injections were detected indiscriminately after both the first and second injection. As a whole, in biological responders treated three times a week, only 82% injections (59/72) were “biologically effective”, compared with 100% (13/13) of Avonex injections.

DISCUSSION

Four IFN beta preparations are presently used in the treatment of MS: Avonex, Betaferon, Rebif 22, and Rebif 44. The differences among the four preparations are in their biochemical structure, dose, dosing frequency, route of administration, and vehicle. Despite these differences,
significant therapeutic effects were observed with each preparation. However, the optimal dosing regimen for IFN beta therapy in this indication is still under debate. Moreover, several studies have demonstrated different degrees of immunogenicity, probably due to one or more of the above mentioned differences.

To compare the degree and duration of modulation of the biological response induced by the four IFN beta preparations, in the present study we evaluated in vivo changes in MxA gene expression in patients with MS during five days of treatment. As far as we know, such temporal characterisation of biological activity has not been described previously. In the other investigations in which IFN beta biological activity was studied for several consecutive days, the following topics were analysed: (a) healthy volunteers treated with IFN beta rather than treated patients with MS; (b) study of only one or two preparations instead of four, and (c) data obtained ex vivo instead of in vivo. Moreover, the authors measured protein markers such as β2-microglobulin, OAS, and MxA. Although these proteins are considered as classic markers of the biological activity of IFN beta, they are characterised by slow decay. Therefore, although the protein quantification using methods such as enzyme-linked immunosorbent assay (ELISA) is simpler than the measurement of the specific transcript, we preferred mRNA quantification because mRNA has a shorter half-life than the protein, and its level reflects biological activity of every single injection allowing the detection of small fluctuations in expression.

In the present study, two categories of patients were identified based on MxA mRNA induction: IFN beta biological non-responders and IFN beta biological responders.

As previously demonstrated, the lack of biological activity correlated with both higher NAb titres and persistent presence of NAb. Under these conditions, IFN beta injections always failed to increase MxA mRNA levels independently of dose or dosing frequency of the treatment. On the other hand, the profiles of MxA AUCs in patients presenting persistently low NAb titres (<45 TRU) show a MxA induction, although at significantly lower levels (fig 1). This could imply that low NAb titres can be overcome by increasing the dose of IFN beta, whereas in the presence of higher NAb titres, the biological response to IFN beta is always abolished, even by higher doses and dosing frequency of the treatment.

Analysis of MxA mRNA levels in IFN beta biological responders clearly demonstrated higher biological responses in patients treated three times a week instead of once a week. The cumulative biological activities, as measured by AUC, approached statistical significance, although only two out of the three weekly injections were considered in this study. The third weekly injection influenced the level of MxA mRNA on day 1 (+156 hours for patients treated with Avonex and +60 hours for patients treated with Betaferon and the two Rebifs) as it was significantly higher in patients treated with Betaferon and the Rebifs compared with those treated with Avonex.

In addition, in IFN beta biological responders, the profiles of MxA concentration time curves demonstrate that the IFN signalling pathway and specific cell surface receptors can be stimulated more than once a week. However, the use of a three times a week dosing schedule induces 18% biologically “ineffective” injections, identified by the absence of a clearly detectable biological activity. Indeed, MxA increase after every IFN beta injection was observed only in 64% of responder patients treated three times a week compared with 100% responder patients treated once weekly. The remaining 36% of responder patients treated three times a week, failed to show increased Mxa expression after one of the two injections. This phenomenon could be due to non-compliance, however, this seems unlikely as a small increase in MxA gene expression was observed after a few “ineffective” injections. Moreover, all patients treated with Avonex

Table 2  MxA mRNA expression in patients negative for neutralising antibodies during five days of treatment

<table>
<thead>
<tr>
<th></th>
<th>Avonex (n=14)</th>
<th>Betaferon (n=6)</th>
<th>Rebif 22 (n=17)</th>
<th>Rebif 44 (n=10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 1</td>
<td>69 (43)*</td>
<td>284 (243)</td>
<td>142 (88)</td>
<td>109 (57)</td>
</tr>
<tr>
<td>Day 2</td>
<td>351 (193)</td>
<td>429 (352)</td>
<td>397 (331)</td>
<td>258 (112)</td>
</tr>
<tr>
<td>Day 3</td>
<td>206 (102)</td>
<td>189 (76)</td>
<td>235 (213)</td>
<td>147 (74)</td>
</tr>
<tr>
<td>Day 4</td>
<td>159 (122)*</td>
<td>498 (242)</td>
<td>321 (264)</td>
<td>360 (431)</td>
</tr>
<tr>
<td>Day 5</td>
<td>95 (83)*</td>
<td>261 (272)</td>
<td>191 (160)</td>
<td>201 (110)</td>
</tr>
</tbody>
</table>

Values are mean (SD) fgMxA/pgGAPDH.

*Statistical difference was found between the Mxa mRNA levels induced by Avonex versus Betaferon, Rebif 22, and Rebif 44 on day 1, 4, and 5. No difference was found on day 2 (+12 hours after the first injection) and day 3 (+36 hours).
showed increase of MxA mRNA level after their single injection. On the other hand, the absence, or greatly reduced, MxA induction could have a biological basis. Therefore, it seems to be more likely that PBMCs of patients treated with IFN beta, undergo a process of desensitisation in response to repeated exposure to the cytokine. Accordingly, recently it has been observed that in vitro T cells become desensitised as a result of persistent IFN beta-1a stimulation, regaining full responsiveness to treatment by 168 hours.\(^1\)

Despite clear evidence of higher biological response in patients treated three times a week, it is unclear whether this difference is clinically relevant, as the lower biological activity of Avonex may be counterbalanced by its lower incidence of NAb induction observed in all therapeutic trials.\(^2\)\(^3\)\(^4\)\(^5\)\(^6\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\)\(^12\)\(^13\) In longitudinal comparisons between the different types of IFN beta,\(^2\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\)\(^12\)\(^13\)\(^14\)\(^15\) and in comparison with placebo controlled clinical trials,\(^2\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\)\(^12\)\(^13\)\(^14\)\(^15\) it has been observed that in vitro T cells become desensitised as a result of persistent IFN beta-1a stimulation, regaining full responsiveness to treatment by 168 hours.\(^1\)

In conclusion the results of the present study demonstrate that an optimal IFN beta regimen is not yet available: Avonex, given once a week, shows significantly lower cumulative biological activity, but significantly lower incidence of NAb, compared with both Betaseron and Rebif.\(^1\)\(^2\)\(^3\)\(^4\)\(^5\)\(^6\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\)\(^12\)\(^13\)\(^14\)\(^15\)\(^16\)\(^17\)\(^18\) Hence, to tailor the best treatment in both patients with newly diagnosed MS and those already receiving treatment, the neurologist must carefully consider the results of clinical trials, the pharmacokinetetic data, the risk of loss of therapeutic efficacy due to the development of NAb, and the peculiar clinical and prognostic characteristics of each patient.

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Competing interests: AB, SM, MC, and ADS have been reimbursed by Farmades, Serono, and Dompé Biotech for attending several conferences; AB received fees for lectures by Serono, Dompé Biotech and Biogen; AB received funds for research and for staff members from Serono and Dompé Biotech. Farmades is the Italian distributor of Betaseron; Serono is the manufacturer of Rebif. Biogen of Avonex and Dompé Biotech is the Italian distributor of Avonex. The other authors have nothing to declare.

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HISTORICAL NOTE

Cotugno and cerebrospinal fluid

In 1761 Cotugno gave the first reliable account of ventricular and subarachnoid fluid. Until Cotugno, anatomists had found empty spaces around the brain and cord and thought that in life they were filled by vapour. Willis said the ventricles were empty spaces, or served the “vile duty of a blood-siphon.” He also wrote about typhus and gave a fine syndrome was subsequently applied to unilateral sciatic pain.”

Further, he noted the incoagulability of CSF in health, but like urine in nephritis, which he observed some 50 years before Bright, it clouded on boiling, only in disease. This work was overlooked until Magendie reprinted it in 1827.

In this crucial work, Cotugno, an astute observer and clinician, differentiated sciatic nerve pain from arthritis of the hip, probably for the first time. The eponym Cotugno’s syndrome was subsequently applied to unilateral sciatic neuralgia. He also wrote about typhus and gave a fine description of the pathology of smallpox pustules.

Domenico Felice Antonio Cotugno (1736–1822)

Near the heel of Italy lies the town of Ruvo Pugliese, the birthplace (29 Jan 1736) of Cotugno. Most of his life he spent in Naples. His family were poor and hardship was his constant companion in his formative years. After medical training in Salerno, he worked in the University of Naples and the Ospedale degli Incurabili. Cotugno surmounted serious illness while resident at the hospital. He became an assistant at the Ospedale degli Incurabili. In 1766 he became professor of anatomy, the leading physician in Naples, and director of the Ospedale. By the age of 31 he was widely acclaimed for his excellent publications, including two books.

When he was only 25, in 1761, his dissertation, 

_“As in the pericardium...a thin humour constantly exaltes from the arteries into the ventricles of the brain and is constantly drawn back through the veins...so often the collected moisture turns into water and even distends the ventricles...A great abundance of water has been found in the ventricles of apoplexies, the soporose, convulsives, paralytics, and victims of epidemic fevers; hydrocephalus even more.”_

Cotugno studied 20 adult male bodies. He established the free circulation between the cranial and spinal dura of cerebrospinal fluid (sometimes referred to as liquor Cotunnii). His lucid description indicating its formation and absorption from blood vessels is contained in his work on sciatica.

“Not only does this water contained in the tube of dura mater enmeshing the spinal marrow [cord] from the occiput to the os sacrum, surround the marrow constantly, but it also abounds in the hollow of the skull and fills all the spaces found between the brain and the encompassing dura mater... It seems to be a human law that the space around the spinal marrow that is filled with water increases with man’s age...Hitherto anatomists have not observed this large collection of water in the spine and around the brain because of the ridiculous method usually employed for the dissection of bodies...they cut off the head with the neck...all the fluid collected around the brain and spinal marrow is at once lost... and the anatomist is misled by the appearance of empty spaces...It seems beyond all doubt that the spinal fluid, as well as that which humectifies all other cavities of the body, constantly oozes from the extremities of the smallest arteries and, finally is absorbed through very small inhaling veins, so that there is a continual state of renovation.”

References

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