We report a case of chronic meningitis due to capsule-deficient Cryptococcus neoformans which could not be diagnosed by routine morphological and immunological cerebrospinal fluid (CSF) examination. Repeated CSF examination and culture did not disclose the fungal body, and the cryptococcal antigen tests were always negative. Culture of ventricular fluid showed non-encapsulated cultured cells which were stained positively by indirect immunofluorescence using diluted sera from another patient diagnosed with cryptococcal meningitis. Inoculation of dispersed colonies into mice resulted in encapsulated C neoformans. It is important to suspect capsule-deficient C neoformans when the conventional diagnostic approaches fail to identify the organism or antigens.

We report a case of chronic meningitis due to capsule-deficient Cryptococcus neoformans which could not be diagnosed by routine morphological and immunological cerebrospinal fluid (CSF) examination. Repeated CSF examination and culture did not disclose the fungal body, and the cryptococcal antigen tests were always negative. Culture of ventricular fluid showed non-encapsulated cultured cells which were stained positively by indirect immunofluorescence using diluted sera from another patient diagnosed with cryptococcal meningitis. Inoculation of dispersed colonies into mice resulted in encapsulated C neoformans. It is important to suspect capsule-deficient C neoformans when the conventional diagnostic approaches fail to identify the organism or antigens.

**CASE REPORT**

A 42 year old man working as a farmer was admitted to a local hospital on July 6, 1992 after having had an episode of loss of consciousness following tonic-clonic convulsion. There was a history of chronic heavy alcohol consumption. A low grade fever and general fatigue preceded the event by several weeks. His cerebrospinal fluid (CSF) cell count from lumbar puncture was increased to 168/mm³ (lymph:polys 1:1) and CSF protein was 116 mg/dl. Aciclovir, ceftriaxone, and sulbenicillin sodium were started intravenously. However, his symptoms did not improve and 1 month later isoniazid, rifampicin, ampicillin, and minocycline hydrochloride were started. On October 26, he had further fits and deterioration. He was transferred on December 16 to a university general hospital because of the difficulty in identifying the cause of meningitis. CSF findings were pleocytosis (74/mm³) and increased protein (151 mg/dl). Herpes simplex virus, varicella zoster virus antibodies, and cryptococcal antigen were negative in CSF. Despite institution of antituberculous regimens again, his clinical course slowly deteriorated and on April 16, 1993, he was transferred to our hospital.

On admission, he was moderately debilitated, his temperature was 37.1°C, and his blood pressure was 162/100 mm Hg. Thereafter, he was conscious most of the time and could express himself, but he was markedly disoriented. He could stand and walk in an unsteady, ataxic manner and Romberg’s test was positive. Deep tendon reflexes were active throughout and the plantar response was bilaterally extensor. The forced grasping and sucking reflexes were seen. Sense of position was diminished in his bilateral lower extremities. The cranial nerves were normal. His muscle power was not fully tested but showed no apparent weakness. Neither stiff neck nor the Kernig’s sign were observed.

A lumbar puncture yielded clear, colourless fluid with an opening pressure of 140 mm CSF, an increased cell count (69/mm³, 78% monocytes), elevated protein (221 mg/dl), and low sugar (5 mg/dl, CSF/blood glucose ratio 8.2%). Gram staining, Indian ink preparations, CSF cytology with Ziehl-Neelsen staining, cultures for fungi, and cryptococcal antigen were all negative, and CSF examinations for Mycobacterium tuberculosis by culture and the polymerase chain reaction method were also negative. Full blood count showed RBC 4.10 million/mm³, platelets 75 000/mm³, WBC 6100/mm³, lymphocytes 2257/mm³, CD4 45.4%, and CD8 23.1%. Laboratory studies revealed moderately impaired liver function (GOT 35 IU/l, GPT 16 IU/l, LDH 443 IU/l, γ-GTP 63 IU/l, cholinesterase 185 IU/l), total serum protein 7.4 g/dl, and albumin 3.9 g/dl. Fasting blood sugar was 97 mg/dl. Hepatitis B surface antigen, hepatitis C antibody, and HIV antibody were negative. CT brain scan with contrast showed a small, circumscribed enhancing area at the right side of the basal cistern, suggesting the presence of a small granuloma (fig 1A). On MRI T2 image, a circumscribed high signal intensity area was observed in the right ventral caudoputamen (fig 1B).

The patient complained of nausea. As an endoscopic examination had disclosed rather vulnerable looking oesophageal varices, endoscopic sclerotherapy was performed on several occasions. A liver biopsy indicated advanced cirrhosis. Following generalised epileptic status which lasted intermittently for several hours, the patient became stuporous. CSF examinations, repeated several times, always disclosed lymphocytic pleocytosis (195/mm³) without atypical cells, increased protein (99 mg/dl), and low glucose (48 mg/dl, CSF/blood ratio 18%). However, the differential diagnoses of cryptococcal meningitis, tuberculous meningitis, and meningeval carcinomatosis were considered. We highly suspected cryptococcal meningitis because there were no findings attributable to pulmonary tuberculosis and the antituberculous agents were not effective, and CSF cytology did not show carcinomatous cells. Therefore, an Ommaya reservoir was placed with the catheter tip inserted into the right frontal horn, both to study the ventricular fluid and to administer amphotericin B into the ventricle. However, cryptococcal antigen in the ventricular fluid via the Ommaya reservoir was reported to be equivocally positive. When the ventricular fluid was cultured on Sabouraud dextrose agar, milk-white colonies were seen after several days. The Indian ink preparation showed small round, bright spots with occasional
budding (fig 1C). Inoculation of dispersed colonies into ICR mice resulted in the classical appearance of \textit{C. neoformans} with thick walls in their ascites and also in their brains after 2 weeks (fig 1D). Moreover, the non-encapsulated cultured cells were stained positively by indirect immunofluorescence using diluted sera from another patient diagnosed with cryptococcal meningitis (E).

**DISCUSSION**

In this case, the typical encapsulated fungal body was not observed in or cultured from the CSF. Moreover, cryptococcal antigen was repeatedly negative by latex agglutination assay of the CSF. A diagnosis of chronic meningitis caused by capsule-deficient \textit{C. neoformans} was established by indirect immunofluorescence staining and formation of the thick capsule after inoculation into the murine peritoneal cavity. In fact, only a few cases of cryptococcal meningitis caused by the capsule-deficient form have been reported.\(^1\)\(^2\)

Capsule-deficient \textit{Cryptococcus} produces sufficient capsule to inhibit phagocytosis when incubated with human lung tissue in vitro.\(^2\) In our case, the capsule-deficient type did form thick capsules in 2 weeks when inoculated into the murine peritoneal cavity. The capsule-deficient \textit{C. neoformans} elicited an intense inflammatory response characterised by early suppuration and phagocytosis, followed by a marked histiocytic and fibroblastic reaction, but with limited spread of infection compared to the capsulated yeast form.\(^4\) Presumably, the brain MRI and CT scan showed a focus of parenchymatous inflammation and granuloma in our case. Meningitis due to capsule-deficient \textit{C. neoformans} has previously been reported in AIDS patients (isolated from CSF) and in Japanese non-AIDS patient (isolated from ventricular fluid).\(^1\)\(^2\) The difference in these cases suggests that the host immune response influences the spread of capsule-deficient \textit{C. neoformans}. Similarly, in our case repeated CSF examinations by lumbar puncture failed to detect yeast forms. The susceptibility of the capsule-deficient form to phagocytosis is probably the major reason why repeated CSF cultures failed to grow \textit{Cryptococcus}, as it was quickly engulfed by macrophages on its way to the lumbar CSF space.

The repeated cryptococcal antigen tests of CSF were always negative. The sensitivity of the antigen test was 76% and its specificity was 98.9% for encapsulated \textit{Cryptococcus}, however capsule deficiency would make the detection of cryptococcal antigen rather difficult, since the latex agglutination test for cryptococcal antigens detects soluble capsular polysaccharide antigen rather difficult, since the latex agglutination test for cryptococcal antigens detects soluble capsular polysaccharide antigens with the antibody fixed to latex particles.\(^5\)

It is important to consider the possibility of this capsule-deficient form when the presence of cryptococcal meningitis is strongly suspected, keeping in mind that the conventional diagnostic approaches fail to identify both the organism and the antigens.

**REFERENCES**

Difficulty in diagnosing chronic meningitis caused by capsule-deficient *Cryptococcus neoformans*

Y Sugiura, M Homma and T Yamamoto

*J Neurol Neurosurg Psychiatry* 2005 76: 1460-1461
doi: 10.1136/jnnp.2004.052662

Updated information and services can be found at:
http://jnnp.bmj.com/content/76/10/1460

These include:

**References**
This article cites 5 articles, 1 of which you can access for free at:
http://jnnp.bmj.com/content/76/10/1460#BIBL

**Email alerting service**
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Topic Collections**
Articles on similar topics can be found in the following collections

- Infection (neurology) (494)
- Immunology (including allergy) (1943)

**Notes**

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/