Ambiguous value of *Haemophilus influenzae* isolation in Guillain-Barré and Fisher syndromes

**M Koga, S Koike, K Hirata, N Yuki**

**SHORT REPORT**

Guillain-Barré syndrome (GBS) is a heterogeneous disorder in which various infections may occur before and after neurological onset. Such preceding infections as *Campylobacter jejuni*, cytomegalovirus, Epstein-Barr virus, and *Mycoplasma pneumoniae* are generally accepted as causes of GBS. The antecedent infectious agent is usually determined by a serological test or isolation of the agent after GBS onset, sometimes making it difficult to distinguish the antecedent infection from complications of GBS itself. This is a problem in the isolation of respiratory infectious agents, because nosocomial respiratory tract infections resulting from aspiration pneumonia and intratracheal intubation are common in GBS.

*Haemophilus influenzae*, a major pathogen of community acquired respiratory infection, is considered a causative agent of GBS and Fisher syndrome, but the frequency of this infection in GBS is controversial. It is considered a causative agent of GBS and Fisher syndrome, but the frequency of this infection in GBS is controversial.

**Background:** *Haemophilus influenzae* is considered a causative agent of Guillain-Barré syndrome (GBS) and Fisher syndrome, but the frequency of this infection in GBS is controversial.

**Objective:** To determine whether isolation of *H. influenzae* indicates it is a causative agent in GBS and Fisher syndrome.

**Results:** Four (15%) of 27 patients with GBS and Fisher syndrome in whom *H. influenzae* was isolated were also seropositive for *Campylobacter jejuni*. Antiganglioside IgG antibodies in these four patients did not cross react with their *H. influenzae* lipo-oligosaccharides, whereas antiganglioside antibodies in the four patients with positive serology for *H. influenzae* did.

**Conclusions:** The findings suggest that *H. influenzae* isolation is not always indicative of the causative agent in these syndromes and that tests for other infections should be made, even in cases of positive culture.

**RESULTS**

Antiganglioside IgG antibodies were detected in 10 of 11 GBS patients (91%) and in all 16 Fisher syndrome patients. Frequent antibodies were anti-GM1 (73%) and anti-GM1b (55%) in the GBS patients, and anti-GQ1b and anti-GT1a (both 100%) in the Fisher syndrome patients.

Four patients (15%; three GBS; one Fisher syndrome) had positive serology for *C. jejuni* and seven (26%; five Fisher syndrome; one GBS; one Fisher syndrome/GBS) had positive serology for *H. influenzae*, but none for both. All the patients were negative for cytomegalovirus and *M. pneumoniae*. Of the four *C. jejuni* seropositive patients, one had a previous history of soft faeces, but the others had no preceding gastrointestinal symptoms (table 1). Follow up serum was available for one *C. jejuni* seropositive patient (patient 3, table 1) who had a significant anti-*C. jejuni* IgA titre decrease during the recovery phase of illness. IgG antibodies to GM1, GM1b, GT1a, and GQ1b were detected in all four patients. In contrast, all seven patients with positive serology for *H. influenzae* had a previous history of respiratory infection symptoms. Anti-GQ1b and anti-GT1a IgG antibodies were detected in six of these and anti-GM1 IgG antibody in the seventh.

Cross reactivity of antiganglioside IgG antibodies to the LOSs of their *H. influenzae* isolates was examined in eight patients, four with positive serology for *C. jejuni* and four for *H. influenzae*. Cross reactivity was absent in the four who had positive *C. jejuni* serology, whereas it clearly was present in at

**METHODS**

We examined recent infections (*H. influenzae*, *C. jejuni*, cytomegalovirus, and *M. pneumoniae*) serologically, as described elsewhere, in 27 patients with GBS (n = 11), Fisher syndrome (n = 14), and overlapping Fisher syndrome/GBS (n = 2), from whom *H. influenzae* had been isolated in sputum or swab cultures. All the strains were isolated shortly after neurological onset, and artificial ventilation was needed for five patients after culture sampling. Serotypes of the isolates were all non-typable, the biotypes predominantly II and III, like the isolates of uncomplicated respiratory infections. There was a history of antecedent respiratory infection symptoms in 21 patients (78%).

Serum IgG antibodies to GM1, GM1b, GM2, GD1a, GaINAc-GD1a, GD1b, GT1a, GT1b, and GQ1b were assayed by enzyme linked immunosorbent assay (ELISA), as reported elsewhere. An absorption study was done to examine whether these antiganglioside antibodies cross reacted with the LOS on the isolates. Crude LOS was prepared as described previously. A 100 µl portion of diluted serum, which gave an optical density of between 1.0 and 2.0 for antiganglioside antibodies, was incubated in LOS precoated wells (LOS lysate 1 µl per well) of microtitre plates at 4°C for 48 hours. The supernatant was used as the primary antibody in the standard ELISA. Absorption rates are expressed as percentages of the optical densities obtained with and without absorption treatment.

**Abbreviations:** GBS, Guillain-Barré syndrome; LOS, lipo-oligosaccharide
least three of the four *H influenzae* seropositive patients examined (fig 1). The target ganglioside of the antibodies was not related to cross reactivity (data not shown).

**DISCUSSION**

We found that 15% of our patients from whom *H influenzae* was isolated had positive serology for recent *C jejuni* infection. The specificity of the *C jejuni* serological assay system used is 97%; moreover, a significant titre decrease during the recovery phase was confirmed in one patient, indicating that there were no false positive results. In GBS and Fisher syndrome after *H influenzae* and *C jejuni* infections it is hypothesised that the ganglioside mimics present on the bacterial LOSs produce antiganglioside antibody. The antibodies in our *H influenzae* seropositive patients did cross react with their isolate LOSs, whereas those in the *C jejuni* seropositive patients did not. This suggests that production of antiganglioside antibodies in the latter patients was not triggered by *H influenzae*, although *H influenzae* infection had certainly occurred, either before or after the neurological onset. It also is noteworthy that only one of four *C jejuni* seropositive patients had a history of preceding gastrointestinal symptoms, but this does not necessarily indicate false positive results in our *C jejuni* serology assay because *C jejuni* has been isolated from some GBS/Fisher syndrome patients who experienced only upper respiratory infectious symptoms or fever. Our findings suggest that *H influenzae* isolation does not always indicate that this is the causative agent in GBS and Fisher syndrome. Even when *H influenzae* is isolated, we should also test for other infections—in particular *C jejuni*—irrespective of the antecedent symptoms. This also may hold true for cases in which other agents are isolated and serological evidence obtained. We believe that more careful judgment of the causal agent in GBS and Fisher syndrome will lead to a decrease in the numerous case reports of sham causal infection in these syndromes.

Positive *H influenzae* serology was found in 26% of the tested cases in whom *H influenzae* had been isolated. This low frequency may partly reflect the assay’s low sensitivity, or it may indicate that *H influenzae* infection was merely a complication of GBS or Fisher syndrome in some of the patients, because most of the serum samples assayed were obtained soon after neurological onset, at a time when neither antibody response nor *H influenzae* infection may have occurred. However, anti-GQ1b and anti-GT1a antibodies were detected in most of the *H influenzae* seropositive patients, all of whom had Fisher syndrome. This supports our previous results showing that *H influenzae* infection is associated more closely with Fisher syndrome than with GBS and that production of antiganglioside antibody is mediated by a GQ1b/GT1a epitope on the bacterial LOS. A serological *H influenzae* test and examination of the cross reactivity of antiganglioside antibodies with the isolate LOS would help in determining the causal role of the bacterium when *H influenzae* is isolated in such cases.

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**Table 1** Patients from whom *H influenzae* was isolated and who had positive serology for recent *C jejuni* infection

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age (years)/sex</th>
<th>Diagnosis</th>
<th>Antecedent symptoms</th>
<th>Artificial ventilation</th>
<th>IgG antibody against</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>5/F</td>
<td>GBS</td>
<td>RTI and headache</td>
<td>Yes</td>
<td>GM1 and GT1a</td>
</tr>
<tr>
<td>2</td>
<td>33/F</td>
<td>GBS</td>
<td>Soft faeces</td>
<td>No</td>
<td>GM1b</td>
</tr>
<tr>
<td>3</td>
<td>67/F</td>
<td>GBS</td>
<td>Fever and general fatigue</td>
<td>No</td>
<td>GM1</td>
</tr>
<tr>
<td>4</td>
<td>66/M</td>
<td>FS</td>
<td>RTI</td>
<td>No</td>
<td>GM1b, GT1a, and GQ1b</td>
</tr>
</tbody>
</table>

FS, Fisher syndrome; GBS, Guillain-Barré syndrome; RTI, respiratory tract infection.

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**Figure 1** Cross reactivity of antiganglioside IgG antibodies in *H influenzae*-isolate patients to their isolate lipo-oligosaccharide (LOS). Absorption rates are given as percentages of optical densities obtained with and without absorption. Circles represent the mean absorption rates of five wells, error bars the standard errors. Antiganglioside IgG antibodies did not cross react with *H influenzae* LOS in patients with serological evidence of recent *C jejuni* infection, whereas the antibodies did react with the isolate LOS in those with serological evidence of recent *H influenzae* infection. Ab, antibody.
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NEUROLOGICAL PICTURE

Ultra fast resolution of acute post-traumatic subdural haematoma

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Post-traumatic acute subdural haematoma (ASDH) is a life threatening condition. The traditional approach is urgent surgery in haematomas with a mass effect. We report a case with a large post-traumatic ASDH whose haematoma disappeared markedly within 2 hours.

A 19 year old man sustained a head trauma at 22:30 and presented with Glasgow Coma Scale (GCS) score of 9 to the state hospital. CT scan showed a right hemispheric ASDH and a marked midline shift (panel A). He was transferred to our hospital for urgent surgery. In our hospital his GCS score was 12. A repeat CT scan at 00:30 showed substantial decrease of ASDH and oedema (panel B). Six hours later CT revealed total resolution of ASDH and the appearance of subarachnoidal haemorrhage (panel C). On the third day he was completely alert with a GCS of 15. MRI on the fifth day showed mild subarachnoidal haemorrhage (panel D). He was discharged on the tenth day with excellent neurological condition.

Although infrequently reported, rapid spontaneous resolution of an ASDH may be underestimated. Two possible hypotheses for spontaneous resolution have been proposed: (1) the haematoma is diluted by cerebrospinal fluid and is washed out; (2) the haematoma is compressed by the pressure produced by acute cerebral swelling and redistributed but also washing out of haematoma by retrograde flow into the subarachnoidal space may be the causes of ultra fast spontaneous resolution of an ASDH. In cases showing rapid improvement of neurological status a subsequent control CT is indicated before surgery.

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