Effects of psychotropic drugs on canine cerebral metabolism and circulation related to EEG—diazepam, clomipramine, and chlorpromazine

ATSUO SARI1, YASUO FUKUDA, TAKEFUMI SAKABE, TSUYOSHI MAEKAWA, AND TOSHIZO ISHIKAWA

From the Department of Anesthesiology, Yamaguchi University School of Medicine, Ube, Yamaguchi, Japan

SYNOPSIS The effects of diazepam, clomipramine, and chlorpromazine upon cerebral metabolism and blood flow were examined separately in 18 dogs. After the administration of diazepam or clomipramine, cerebral cortical oxygen consumption (CMR$_{O_2}$) decreased significantly by a maximum of 17% and 13% of control within 10 minutes and 15 minutes, and returned to control at 120 minutes and 90 minutes, respectively. Chlorpromazine, however, decreased by a maximum of 10% of control, a level which continued throughout the period of observation. It was observed that reduction in CMR$_{glucose}$ was followed by the reduction in CMR$_{O_2}$ at an interval during the early stages of CMR$_{O_2}$ depression. Diazepam produced a significant decrease in CBF accompanied by a reduction in CMR$_{O_2}$, but neither clomipramine nor chlorpromazine had any effect on CBF in spite of reduction in CMR$_{O_2}$. Reduction in CMR$_{O_2}$ both with diazepam and clomipramine was accompanied by slow wave activities of EEG, but with chlorpromazine reduction in CMR$_{O_2}$ was accompanied with less pronounced slow wave activities. It was concluded that the three drugs examined were cerebral metabolic depressants.

It has been generally assumed that the effects of psychotropic drugs upon the central nervous system are related to their therapeutic action in patients with psychiatric disorder. This assumption follows from extensive pharmacological, biochemical, and electrophysiological studies obtained with these drugs. In the present study we examined the effect of three psychotropic drugs—diazepam, clomipramine, and chlorpromazine—on the canine cerebral metabolism and circulation.

Although the effects of chlorpromazine on cerebral oxygen consumption have been extensively investigated in animals (Frowein et al., 1955) or men (Fazekas et al., 1955; Morris et al., 1955; Aizawa et al., 1956; Moyer et al., 1956; Ehrmantraut et al., 1957; Sutherland et al., 1960), where the other two drugs are concerned, despite their profound effects on the central nervous system, little information has been reported regarding their effect on cerebral metabolism (in vivo) and circulation. It was, therefore, the purpose of the present study to attempt to discover whether or not these drugs produce any changes in cerebral metabolism and circulation related to EEG.

A part of the study on diazepam has been reported elsewhere (Maekawa et al., 1974).

METHODS

Eighteen fasting unpremedicated dogs weighing 8 to 26 kg were anaesthetized with halothane (1.0-2.0%) inspired in oxygen. The trachea was intubated with a cuffed endotracheal tube with the aid of succinylcholine and thereafter 10 mg/kg/h was given to maintain muscle paralysis. Ventilation was controlled with an animal respirator, AR-300 (Acoma, Tokyo, Japan). Cannulae were placed in the femoral artery.
Effects of psychotropic drugs on canine cerebral metabolism and circulation related to EEG

for blood sampling and pressure determination, in one femoral vein for the reinfusion of blood, and in another femoral vein for drug administration. Thereafter, the dog was placed in a prone position with the head supported on a block.

The surgical preparation used in the present study for direct measurement of cerebral blood flow (CBF) was originally described in detail by Michenfelder et al. (1968). With this technique, blood flow from the sagittal sinus is diverted through a cannula to an external reservoir at the level of the sinus, measured by timed collection, and returned by pump into the femoral vein. The percentage of the drained region was determined by staining the veins with Sudan III dissolved in vinyl acetate monomer injected from the cannulated portion of the sagittal sinus at the completion of the study. These percentages and the individual brain weights of the dogs studied were used to convert unit of flow per minute to per 100 g of brain weight per minute. After isolation by obliteration of the diploic veins that communicate with the sagittal sinus, the collection and the measurement of the venous drainage of a known portion of the brain provides a ready source for sampling mixed venous blood, which is exclusively representative of the brain tissue.

The oxygen content of arterial and sagittal sinus blood was calculated from measurements of oxyhaemoglobin (IL 182 CO-oximeter, Instrumental Laboratories, Boston, Mass., U.S.A.) and oxygen tension (IL 113 electrodes). Additional measurements included arterial pressure (strain gauge), pH and PaCO₂ (electrode 37°C), and brain temperature (parietal epidural thermistor). Cerebral metabolic rate for oxygen (CMR₀₂) was calculated as the product of CBF and arterial sagittal sinus blood oxygen content difference (C(a−v)O₂). Blood glucose level was determined by an enzymatic method. CMR₉₀₂ was calculated as the product of CBF and arterial sagittal sinus blood glucose difference (C(a−v)glucose). Cerebral vascular resistance (CVR) was calculated as the ratio of the mean arterial pressure (MAP) to the CBF. The oxygen-glucose index (OGI) was calculated as described by Cohen et al. (1967). The EEG was recorded from the parietal lobes using bipolar silver–silver chloride disc electrodes and was analysed every 10 seconds with a frequency analyser, MAF-5 (Nihonkoden, Tokyo, Japan) throughout the study. Every activity was expressed as a percentage of integrated voltage of δ (2–4 Hz), θ (4–8 Hz), α (8–13 Hz), β₁ (13–20 Hz) and β₂ (20–30 Hz) waves.

After completion of the surgical preparation, the inspired halothane was maintained at 0.2% for 1 h so that any possible effects of residual halothane on CMR₀₂, CBF or EEG could be kept constant

FIG. 1 Effects of diazepam, clomipramine, and chlorpromazine on CMR₀₂; and each wave activity of EEG. Three figures represent sequential changes in CMR₀₂; accompanied by changes in the EEG after administration of diazepam, clomipramine, and chlorpromazine. Key: O: δ wave, ●: θ wave, ×: α wave, △: β₁ wave, ○: β₂ wave.
throughout the study. Ventilation was adjusted to maintain normocapnia (PaCO₂ 39 ± 2 mmHg). Sodium bicarbonate was given as needed to maintain a normal buffer base. The epidural temperature was maintained at 37 ± 0.1°C by external means, and haemoglobin levels were maintained above 13 g/dl. Blood loss due to sampling was replaced by fresh heparinized blood.

Control measurements were obtained over a 30 minute period and mean values were calculated from five to eight consecutive determinations of CBF and C(a − v)O₂ and two to three determinations of C(a − v)glucose. The dogs were divided into three groups. Five dogs received 0.25 mg/kg diazepam, five dogs received 2 mg/kg clomipramine, and eight dogs received 0.5 mg/kg chlorpromazine. All drugs were administered intravenously. After control determinations, measurements were done subsequently at 2, 5, 10, 20, 30, 45, 60, 90, 120 and 150 (except for clomipramine) minutes after injection of each drug. No evidence of extracerebral contamination of blood or other cerebral vascular anomalies was found at necropsy.

Statistical significance was tested by Student’s t test for paired data (P < 0.05, considered statistically significant).

RESULTS

The sequential effects of diazepam, clomipramine, and chlorpromazine upon cerebral haemodynamics, metabolism, and EEG are shown in Tables 1, 2, and 3, and Fig. 1.

CEREBRAL METABOLISM Two minutes after the administration of 0.25 mg/kg diazepam, CMRO₂ was reduced significantly and remained decreased over a 90 minute period. Thereafter, it returned to control value. Its minimum level was 84% of control at 10 minutes. OGI at two minutes decreased significantly to 77%.

| TABLE 1 |
| EFFECTS OF DIAZEPAM (0.25 MG/KG) ON CEREBRAL METABOLISM AND CIRCULATION |
| Time (min) | CMRO₂ (ml/100g/min) | CMRglucose (mg/100g/min) | OGI (%) | CBF (ml/100g/min) | MAP (mmHg) | CVR (mmHg/ml/100g/min) | PaCO₂ (mmHg) | PscO₂ (mmHg) |
| Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE |
| Control | 6.3 | 0.4 | 9.1 | 0.6 | 96 | 4 | 60 | 2 | 110 | 9 | 1.85 | 0.19 | 40.8 | 1.5 | 34 | 2 |
| 2 | 5.3 | 0.4* | 9.4 | 1.2 | 77 | 5* | 51 | 10* | 98 | 12 | 1.94 | 0.25 | 41.8 | 1.3 | 33 | 1 |
| 10 | 5.2 | 0.4* | 7.4 | 0.6* | 100 | 5 | 53 | 2* | 101 | 10 | 1.94 | 0.25 | NE | NE | 33 | 1 |
| 30 | 5.7 | 0.4* | 7.9 | 1.0 | 99 | 6 | 54 | 3 | 103 | 9 | 1.98 | 0.26 | NE | NE | 32 | 3 |
| 90 | 5.9 | 0.4* | 9.6 | 1.2 | 87 | 9 | 55 | 3 | 114 | 8 | 2.14 | 0.24 | 39.3 | 1.8 | 29 | 3 |
| 150 | 6.1 | 0.4 | 8.8 | 0.7 | 93 | 5 | 57 | 4 | 110 | 7 | 1.99 | 0.23 | 39.1 | 1.9 | 30 | 3 |

* Significantly different from control (P < 0.05). NE = not examined.

| TABLE 2 |
| EFFECTS OF CLOMIPRAMINE (2 MG/KG) ON CEREBRAL METABOLISM AND CIRCULATION |
| Time (min) | CMRO₂ (ml/100g/min) | CMRglucose (mg/100g/min) | OGI (%) | CBF (ml/100g/min) | MAP (mmHg) | CVR (mmHg/ml/100g/min) | PaCO₂ (mmHg) | PscO₂ (mmHg) |
| Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE | Mean | SE |
| Control | 6.4 | 0.2 | 9.1 | 0.3 | 95 | 4 | 58 | 2 | 123 | 9 | 2.17 | 0.21 | 37.6 | 0.9 | 34 | 2 |
| 5 | 6.3 | 0.4 | 9.3 | 0.5 | 90 | 3 | 59 | 4 | 112 | 13 | 1.98 | 0.32 | 38.1 | 0.4 | 34 | 2 |
| 10 | 5.8 | 0.3* | 9.4 | 0.6 | 84 | 6* | 55 | 4 | 96 | 11* | 1.78 | 0.25* | 37.6 | 0.4 | 33 | 3 |
| 15 | 5.6 | 0.4* | 9.6 | 0.6 | 80 | 4* | 53 | 4 | 79 | 12* | 1.53 | 0.22* | 38.7 | 0.5 | 33 | 4 |
| 30 | 5.9 | 0.3* | 8.5 | 0.6 | 95 | 8 | 54 | 3 | 83 | 13* | 1.56 | 0.22* | 38.4 | 0.4 | 32 | 3 |
| 60 | 6.1 | 0.4 | 9.3 | 0.6 | 88 | 6 | 57 | 3 | 106 | 10 | 1.88 | 0.17 | 38.2 | 0.4 | 32 | 2 |
| 120 | 6.5 | 0.2 | 9.1 | 0.5 | 97 | 3 | 57 | 3 | 115 | 8 | 2.04 | 0.19 | 39.2 | 1.1 | 31 | 2 |

* Significantly different from control (P < 0.05).
A significant reduction in CMRO₂ was observed between 10 and 30 minutes after administration of 2 mg/kg clomipramine. And thereafter CMRO₂ gradually returned to control value during 45 to 120 minutes. The minimum level in CMRO₂ in the clomipramine group was 87% of control at 15 minutes after the injection. There were no significant changes in CMRglucose. The OGI showed a transient significant decrease between 10 to 15 minutes, and thereafter returned to control. With 0.5 mg/kg chlorpromazine, CMRO₂ decreased significantly to 92% of control 10 minutes after injection and continued to decrease throughout a 150 minute period of observation. At five minutes, chlorpromazine briefly, but significantly, caused an increase of CMRglucose while not significantly decreasing CMRO₂. As a result, a significant reduction in OGI occurred.

CEREBRAL HAEMODYNAMICS A statistically significant reduction in CBF was produced at two and 10 minutes in the diazepam group, but was not accompanied by significant changes in MAP or CVR. In each group that was given clomipramine or chlorpromazine, MAP started to decrease significantly at 10 minutes with a concomitant reduction in CVR and gradually returned to control value within 60 minutes in the clomipramine group, but continued to decrease throughout the period observed in the chlorpromazine group. There were no significant changes in CBF in the two groups, which indicates that CBF was maintained by a significant reduction in CVR accompanied by a considerable fall in MAP. No significant changes in PaCO₂ or sagittal sinus PO₂ (PssO₂) were observed during the period of observation in all groups after the administration of each drug.

**EEG AND CMRO₂** Representative EEG changes and corresponding values of CMRO₂ of the three groups are illustrated in Figs 2, 3, and 4. With diazepam, high voltage (100–150 μV) slow

---

**TABLE 3**

EFFECTS OF CHLORPROMAZINE (0.5 MG/KG) ON CEREBRAL METABOLISM AND CIRCULATION

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>CMRO₂ (ml/100g/min) Mean</th>
<th>SE</th>
<th>CMRglucose (mg/100g/min) Mean</th>
<th>SE</th>
<th>OGI (%) Mean</th>
<th>SE</th>
<th>CBF (ml/100g/min) Mean</th>
<th>SE</th>
<th>MAP (mmHg) Mean</th>
<th>SE</th>
<th>CVR (mmHg/ml 100g/min) Mean</th>
<th>SE</th>
<th>PaCO₂ (mmHg) Mean</th>
<th>SE</th>
<th>PssO₂ (mmHg) Mean</th>
<th>SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>6.3</td>
<td>0.1</td>
<td>8.8</td>
<td>0.3</td>
<td>97</td>
<td>2</td>
<td>60</td>
<td>3</td>
<td>94</td>
<td>6</td>
<td>1.57</td>
<td>0.07</td>
<td>39.2</td>
<td>0.7</td>
<td>35</td>
<td>2</td>
</tr>
<tr>
<td>5</td>
<td>6.3</td>
<td>0.1</td>
<td>11.5</td>
<td>1.0*</td>
<td>73</td>
<td>5*</td>
<td>58</td>
<td>3</td>
<td>90</td>
<td>6</td>
<td>1.56</td>
<td>0.10</td>
<td>39.5</td>
<td>0.7</td>
<td>34</td>
<td>2</td>
</tr>
<tr>
<td>10</td>
<td>5.8</td>
<td>0.2*</td>
<td>9.8</td>
<td>0.7</td>
<td>82</td>
<td>6</td>
<td>59</td>
<td>3</td>
<td>84</td>
<td>6*</td>
<td>1.46</td>
<td>0.08</td>
<td>39.6</td>
<td>0.6</td>
<td>34</td>
<td>3</td>
</tr>
<tr>
<td>30</td>
<td>5.8</td>
<td>0.2*</td>
<td>8.8</td>
<td>0.3</td>
<td>89</td>
<td>3</td>
<td>57</td>
<td>2</td>
<td>77</td>
<td>5*</td>
<td>1.33</td>
<td>0.09*</td>
<td>40.0</td>
<td>0.8</td>
<td>32</td>
<td>3</td>
</tr>
<tr>
<td>90</td>
<td>5.8</td>
<td>0.2*</td>
<td>8.4</td>
<td>0.4</td>
<td>93</td>
<td>4</td>
<td>53</td>
<td>2</td>
<td>74</td>
<td>4*</td>
<td>1.42</td>
<td>0.12</td>
<td>39.9</td>
<td>1.3</td>
<td>28</td>
<td>2</td>
</tr>
<tr>
<td>150</td>
<td>5.6</td>
<td>0.2*</td>
<td>8.0</td>
<td>0.3</td>
<td>93</td>
<td>5</td>
<td>49</td>
<td>3</td>
<td>70</td>
<td>6*</td>
<td>1.45</td>
<td>0.15</td>
<td>38.6</td>
<td>0.7</td>
<td>26</td>
<td>2</td>
</tr>
</tbody>
</table>

* Significantly different from control (P < 0.05).

---

**FIG. 2** A typical EEG tracing before and after injection of diazepam with corresponding values of CMRO₂.
wave activity (δ) predominated within two minutes after the injection with a concomitant decrease in fast waves. These EEG changes were statistically significant and thereafter gradually returned to control levels over a 20 to 30 minute period. Such predominant increases in slow wave activity corresponded with the decreases in CMRO₂ (Fig. 2). With clomipramine, slow wave (δ) activities showed a tendency to increase, and fast wave activities decreased within five minutes after injection and reached their lowest value within 15 to 30 minutes, corresponding to a reduction in CMRO₂ (Fig. 3). These EEG changes with clomipramine gradually returned to control levels within 90 to 120 minutes after injection. Chlorpromazine showed different EEG changes from the two former drugs; significant increases in δ and α wave activities were observed from 30 minutes and 10 minutes after injection, respectively, accompanied by a significant reduction in β₂ waves (Fig. 4).

**DISCUSSION**

In dosage similar to that used in clinical practice, diazepam, clomipramine, and chlorpromazine cause a significant reduction in CMRO₂ in dogs.

Reduction in CMRO₂ with diazepam accompanied by a concomitant reduction in CBF occurred within two minutes after the injection. The appearance of the effect of diazepam on CMRO₂ was faster than either clomipramine or chlorpromazine, which possibly indicates that diazepam can pass the blood–brain barrier faster than the other two. This is in agreement with findings reported by Kleijn (1969) that uptake of [³¹C]diazepam in the mouse brain reaches its maximum about one minute after intravenous injection. *In vitro*, Davis *et al.* (1971) demonstrated a reduction in oxygen consumption in rat brain mitochondria.

There has been no available report concerning
the effect of clomipramine or other types of antidepressant drugs on cerebral oxygen consumption in vivo. However, in vitro, Ernsting et al. (1960) investigated the effect of several psychotropic drugs on the oxygen consumption in rat brain and concluded that imipramine reduced oxygen consumption significantly. Di Mascio et al. (1964) found the tricyclic antidepressants, imipramine or desipramine, produced hypnotic-like action in normal men. The present study indicated that their hypnotic-like action may be related to decrease in CMRO₂.

In the dog, Frowein et al. (1955) could not observe any significant changes in CMRO₂ and CBF after the injection of 2–10 mg/kg chlorpromazine. CMRO₂ in man (Fazekas et al., 1955; Morris et al., 1955; Aizawa et al., 1956; Moyer et al., 1956; Ehrmantraut et al., 1957; Sutherland et al., 1960) also showed no changes under chlorpromazine. However, in the present study, the reduction in CMRO₂ observed within 10 minutes after the injection of 0.5 mg/kg chlorpromazine continued throughout the period of observation. This reduction in CMRO₂ lasted much longer than with the other two drugs. Cassano et al. (1965) demonstrated the distribution of chlorpromazine in cat brain using an autoradiographic technique and showed that retention of this agent in the cortex was reduced four hours later. Although many of the reports cited above are contrary to our findings, a possible explanation for this discrepancy could be the difference in the methods used for measuring CBF. The above-mentioned studies determined CBF by the nitrous oxide method of Kety and Schmidt (1948), while, in our study, CBF was determined by the direct measurement method described originally by Michenfelder et al. (1968). Theye and Michenfelder (1968) pointed out greater values for CMRO₂ in their method than those generally observed in whole brain studies. The direct method is valid for a portion of the cerebral cortex, not the whole brain. Therefore, our findings indicate that chlorpromazine may reduce CMRO₂ in the cortex and is in agreement with the results of electrophysiological studies by Gangloff and Monsier (1957), who demonstrated that chlorpromazine does affect the cerebral cortex.

The decrease in OGI observed with diazepam and chlorpromazine is due to disproportional change in CMRglucose to a reduction in CMROG. This was also suggestive in the chlorpromazine group at 10 minutes. Our findings showed that the changes in CMRglucose were slower than those of CMROG at early states of their alteration. This delayed change in CMRglucose was probably due to differences in equilibration of oxygen and glucose metabolism. Chlorpromazine produced a significant increase in CMRglucose at five minutes. Aizawa et al. (1956) found an increase in CMRglucose 30 minutes after injection of chlorpromazine in man that was probably due to a rise in glucose level in the blood, but the blood glucose level was not mentioned. In the present study, a transitory but significant increase in CMRglucose was not accompanied by an elevated blood glucose level. This finding is contrary to previous reports (Norman and Hiestand, 1955; Ryall, 1956; Bonaccorsi et al., 1964; Bachelard and Lindsay, 1966), which suggest hyperglycaemia with this drug. However, Norman and Hiestand (1955) found a profound species variation in hyperglycaemic response to chlorpromazine. At the present, it is difficult to evaluate the significance of the increase in CMRglucose with chlorpromazine.

Changes in CMROG were paralleled by slow wave activities in the EEG (Fig. 1). During a period of reduction in CMROG by both diazepam and clomipramine, slow wave activities were predominant and returned to control levels when CMROG did. With chlorpromazine, however, an increase in slow wave activity was not so marked, whereas a decrease in fast wave activity was significant. Gleichmann et al. (1962) and Ingvar et al. (1962) demonstrated a correlation between cortical oxygen consumption and EEG pattern. The correlation which they found consisted of findings of predominant fast waves with high oxygen consumption and high voltage slow waves with low oxygen consumption. However, we found that chlorpromazine showed a different pattern of EEG changes from the other two drugs, even though there was considerably less reduction in CMROG than the other two; less pronounced slow wave activities. The reduction in CBF in the diazepam group was about the same magnitude as the reduction in CMROG, agreeing with the finding that PssO₂ remained unchanged. After administration of either
clomipramine or chlorpromazine, CBF did not change significantly but roughly paralleled a reduction in CMRO\textsubscript{2}, resulting again in unchanged PssO\textsubscript{2}. Therefore, these changes in CBF seem to be mostly metabolic dependent.

From these results, it was concluded that diazepam and chlorpromazine are cerebral cortical metabolic depressants. This evidence suggests that the reduction in cerebral cortical oxygen consumption may play an important role in their therapeutic action in psychiatric patients.

The authors are grateful to Dr Hiroshi Takeshita, Department of Anesthesiology, Yamaguchi University, Japan, for his valuable advice and criticism throughout this study. We are also grateful to the publishers of Anesthesiology for permission to make use of some of the material used in Table 1.

REFERENCES


