

## LESSON OF THE MONTH

## Sporadic lower limb hypertrophy and exercise induced myalgia in a woman with dystrophin gene deletion

Denis Malapert, Dominique Recan, France Leturcq, Jean-Denis Degos, Romain K Gherardi

### Abstract

**A 25 year old woman, without family history of muscular dystrophy, had had an isolated lower limb hypertrophy since infancy and later experienced exercise-induced myalgia. Genomic DNA analysis showed a deletion of exons 45 to 52 of the dystrophin gene. Uncommon phenotypes of dystrophinopathies and consequences in genetic counselling in women are emphasised.**

(*J Neurol Neurosurg Psychiatry* 1995;59:552-554)

**Keywords:** lower limb hypertrophy; myalgia; muscle dystrophy; dystrophin gene deletion

Recent advances in molecular genetics have focused interest on previously unrecognised phenotypic expressions of dystrophinopathies.<sup>1</sup> These conditions are suspected in males with a family history suggestive of an X linked disorder. Because of the high rate (30%) of spontaneous dystrophin gene mutations, however, immunocytochemistry for detection of dystrophin on muscle fibres and DNA analysis on muscle or blood samples are also important in the evaluation of isolated or sporadic muscle dystrophies. Analysis of DNA is also useful to assess symptomatic female carriers of Duchenne and Becker muscular dystrophies (DMD, BMD), which are recognised with increasing frequency.

We report the case of a young woman, without family history, who had dystrophin gene deletion manifest by bilateral hypertrophy of her calves and thighs, and myalgia on exertion.

### Case report

A 25 year old woman was referred by her occupational medical control for pronounced bilateral lower limb hypertrophy. She had no children. Her parents and her six brothers and sisters were asymptomatic. Hypertrophy of calves and thighs was first noticed by her parents in her infancy. During childhood, she developed a progressive toe walking gait. Achilles tenotomy was performed at the age of 16 with a good result on gait. Cramps and painful sensations induced by exercise developed as she grew older. In adulthood, she

experienced discomfort in her lower limbs during prolonged walks, running, or climbing stairs. In recent years, some exercise induced myalgia developed in the upper limbs, but, as a whole, worsening of muscular symptoms was minimal.

Examination showed global and symmetric pseudohypertrophy of her lower limbs, predominating in her calves and thighs (fig 1). Upper limbs were normal. The muscles in the lower limbs were rubbery and percussion elicited no pain. Testing of muscle strength and tendon reflexes was normal. Other physical examination was normal.

Serum creatine kinase concentration was 680 (normal range 40-225) U/l. Electro-

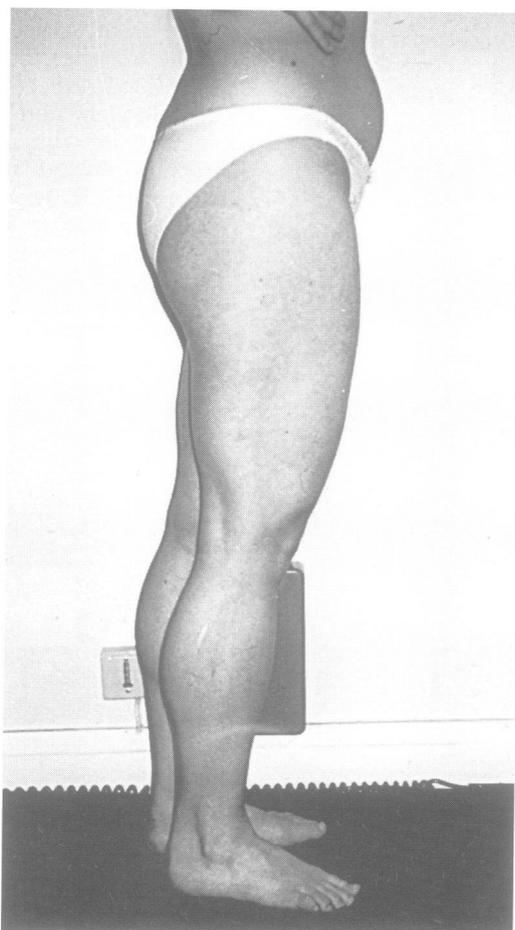


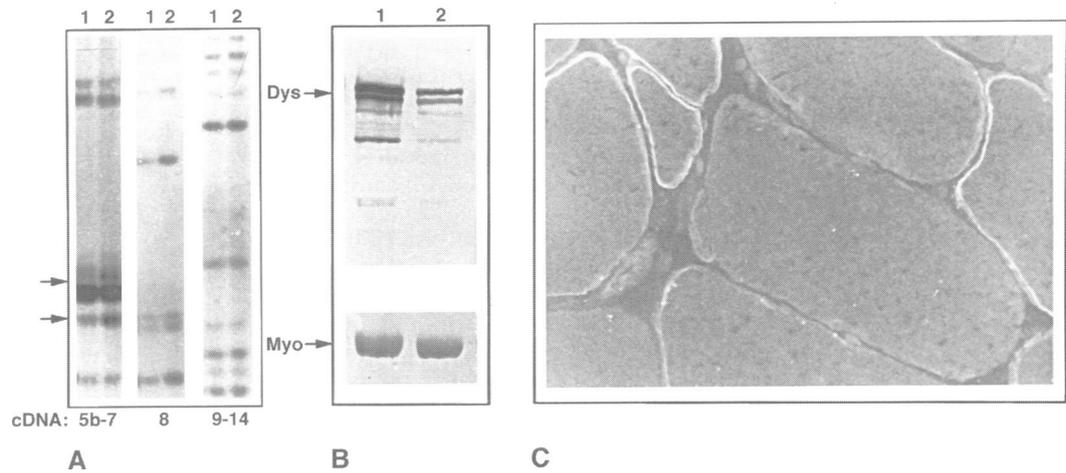
Figure 1 Bilateral hypertrophy of calves and thighs.

**Groupe Nerf-Muscle (ER 269),  
Département de Neurosciences,  
Centre Hospitalo-Universitaire Henri Mondor, 94010-Créteil, France**  
D Malapert  
J-D Degos  
R K Gherardi

**Service de Biochimie Génétique, Hôpital Cochin, 75014-Paris, France**  
D Recan  
F Leturcq

Correspondence to:  
Professor R K Gherardi,  
Groupe Nerf-Muscle (ER 269), Département de Pathologie, Hôpital Henri Mondor, 94010-Créteil, France.

Received 3 April 1995 and in revised form 30 May 1995  
Accepted 23 June 1995



**Figure 2** DNA and dystrophin analysis. (A) Southern blot performed with three dystrophin cDNA probes after *Hind III* digestion. cDNA probe 5b-7 (corresponding to exons 30 to 46): 2 bands (black arrows) containing exons 45 and 46 show half intensity of labelling in the patient (lane 1) compared with a female control DNA (lane 2). The other bands, corresponding to non-deleted exons, are similarly expressed in the patient and the control. cDNA probe 8 (corresponding to the deleted exons 47 to 52): all bands show half intensity of labelling in the patient (lane 1) compared with the control (lane 2). cDNA probe 9-14 (corresponding to the non-deleted exons 53 to 64): all bands show a similar intensity of labelling in the patient (lane 1) and the control (lane 2). (B) Western blot analysis with antibody dys-1. The doublet of dystrophin is normal sized for the patient (lane 2) compared with the control (lane 1) but in reduced amount (20% estimated by densitometry). Dys = dystrophin; Myo = myosin. (C) Immunofluorescence study with dystrophin antibody dys-2 showing a dystrophic myopathological pattern with a mixture of positive and completely negative fibres

myography, ECG, and echocardiography were normal; CT of the lower limbs was in keeping with a fatty degeneration of muscles. The karyotype was normal (46, XX).

A biopsy of quadriceps femoris muscle showed abnormalities consistent with a muscular dystrophy, including considerable variability in fibre size with numerous hypertrophic fibres, splitting, central nuclei, necrotic fibres, small groups of regenerating fibres, endomysial fibrosis, and mild fatty degeneration. An immunofluorescence study of dystrophin was performed with monoclonal antibodies against the rod domain (dys 1), the C-terminal (dys 2), and the N-terminal (dys 3) of dystrophin (Novocastra, Newcastle-upon-Tyne, UK). It showed a mixture of positive fibres with continuous sarcolemmal staining, and completely negative fibres that accounted for 30% of muscle fibres (fig 2). Negative fibres were found with all three antidystrophin monoclonal antibodies. The quantity of dystrophin was estimated at 20% of the normal with western blot analysis using the monoclonal antibody dys 1.

Genomic DNA was analysed by hybridisation with  $6^{(32P)}$ -dCTP labelled cDNA probes encompassing the dystrophin gene, after digestion by restriction enzymes *Hind III* and Southern blotting. The gene dosage showed a deletion ("simple dose" intensity of labelling) of exons 45 to 52 (fig 2).

### Discussion

This woman was evaluated for sporadic bilateral lower limb hypertrophy without weakness, and exercise induced myalgias. She had mildly increased creatine kinase concentrations, and muscle biopsy showed dystrophic changes with absent dystrophin expression in 30% of muscle fibres. Karyotype was normal. Analysis of DNA showed a deletion of exons 45 to 52

of the dystrophin gene on one X chromosome.

Dystrophinopathies are due to mutations in the dystrophin gene on the short arm of the X chromosome at p21,<sup>2</sup> and, therefore, are virtually confined to males. In about 70% of cases of BMD/DMD, mutations of the dystrophin gene result in detectable deletions and duplications.<sup>3-5</sup> Deletion of exons 45 to 52 of the dystrophin gene found in our patient disrupts the transductional reading frame, and gives a shortened and non-functional dystrophin product. This deletion is found on the DNA of male patients with DMD in whom no dystrophin is detected in muscle tissue.<sup>6</sup>

A range of 45-70% of definite female carriers of DMD have increases in serum creatine kinase concentrations,<sup>7,8</sup> and about 10% have clinical symptoms.<sup>9</sup> The proportion of manifesting BMD female carriers is probably much lower.<sup>10,11</sup> So called "manifesting" or "symptomatic" DMD carriers were first recognised in the early 1960s.<sup>12-14</sup> It was the presence of unilateral calf hypertrophy and associated cramps on exercise that prompted Dubowitz in 1963 to report the first muscle biopsy of a known carrier of DMD, and to show overt pathological changes in the muscle.<sup>12</sup> Manifesting female carriers with previously X linked history for DMD in males usually present with proximal limb weakness (80%), and much less often with muscle symptoms without weakness, including calf hypertrophy, myalgia, or cardiac failure.<sup>15</sup> Unlike asymptomatic carriers of DMD, manifesting DMD carriers do tend to show a mosaic pattern on dystrophin immunocytochemistry in their muscle.<sup>16</sup> A lack of correlation between dystrophin expression and clinical weakness, however, has been found in manifesting DMD carriers.<sup>16</sup>

Some reports have described DMD in women with chromosomal abnormalities such as Turner (X0) or Turner mosaic (X/XX or X/XX/XXX) syndromes, a structurally abnor-

mal X chromosome, or an X autosomal translocation.<sup>3</sup> Manifesting female carriers of DMD usually have a normal karyotype, however, and they express the disease as the result of skewed lyonisation of the X chromosome<sup>11</sup> (inactivation of the normal paternal X chromosome in a large proportion of embryonic cells<sup>17</sup>).

In females without a family history of dystrophy, immunohistochemistry for dystrophin and DNA analysis are key tests to assess dystrophinopathy. The first well documented sporadic female cases of dystrophinopathy were three 10 year old girls who had moderate calf hypertrophy, moderate to severe weakness, raised creatine kinase in serum, and dystrophin deficiency assessed by immunohistochemical study of muscle and immunoblot analysis.<sup>18</sup> In a large multicentre study of 25 sporadic cases of manifesting female carriers of DMD,<sup>15</sup> 40% had proximal limb weakness before the age of 10, 24% had cramps or myalgia, 24% had grossly increased serum creatine kinase, 8% were easily tired, and 4% (one patient) had progressive proximal limb weakness beginning at the age of 45. Female patients initially believed to have sporadic limb girdle dystrophy have occasionally been found to be carriers of DMD,<sup>19,20</sup> as have 27–38% of male patients with sporadic limb girdle dystrophy.<sup>21,22</sup> By contrast, our patient had no weakness at the age of 25. Myalgia on exertion is now well recognised in male patients with dystrophinopathy.<sup>1</sup> Hypertrophy of the calves is a typical sign in up to 77% of manifesting female carriers of DMD.<sup>15</sup> Unusual phenotypic expressions of dystrophinopathies in males include quadriceps myopathy,<sup>23</sup> X linked myoglobinuria,<sup>24</sup> cramps and myalgia,<sup>25</sup> and dilated cardiomyopathy.<sup>26</sup> The clinical range of the disease could even include some entities of obscure pathogenesis such as monomelic hypertrophy with progressive myopathy<sup>27</sup>; these rare patients should be screened for dystrophin because asymmetry of symptoms can be seen in more than 25% of manifesting carriers.<sup>12,28</sup>

Deficiencies in dystrophin associated glycoproteins (DAGs), such as the 50 kDa DAG encoded by chromosome 17, adhalin, can cause autosomal recessive muscular dystrophies mimicking DMD.<sup>29</sup> To what extent adhalin deficiencies may account for clinical symptoms resembling those of DMD carriers in heterozygotes remains to be determined.

We conclude that isolated hypertrophy of the thighs and calves may be the only manifestation of sporadic dystrophinopathy, and this has practical consequences in genetic counselling.

1 Comi GP, Prella A, Bresolin N, *et al.* Clinical variability in Becker muscular dystrophy. Genetic, biochemical and immunohistochemical correlates. *Brain* 1994;117:1–14.

- 2 Koenig M, Hoffman EP, Bertelson CJ, Monaco AP, Feener C, Kunkel LM. Complete cloning of the Duchenne muscular dystrophy (DMD) cDNA and preliminary genomic organization of the DMD gene in normal and affected individuals. *Cell* 1987;50:509–17.
- 3 Engel AG, Yamamoto M, Fischbeck KH. Dystrophinopathies. In: Engel AG, Franzini-Armstrong C, eds. *Myology, basic and clinical* Vol II. 2nd ed. New York: McGraw-Hill, 1994:1133–87.
- 4 Ioannou P, Christopoulos G, Panayides K, Kleanthous M, Middleton L. Detection of Duchenne and Becker muscular dystrophy carriers by quantitative multiplex polymerase chain reaction analysis. *Neurology* 1992;42:1783–90.
- 5 Yau SC, Roberts RG, Bobrow M, Mathew CG. Direct diagnosis of carriers of point mutations in Duchenne muscular dystrophy. *Lancet* 1993;341:273–5.
- 6 Monaco A, Bertelson C, Liechti-Gallati S, Kunkel L. An explanation for the phenotypic differences between patients bearing partial deletions of the DMD locus. *Genomics* 1988;2:90–5.
- 7 Griggs RC, Mendell JR, Brooke MH. Clinical investigation in Duchenne dystrophy: V. Use of creatine kinase and pyruvate kinase in carrier detection. *Muscle Nerve* 1985;8:60–4.
- 8 Moser H. Duchenne muscular dystrophy: pathogenetic aspects and genetic prevention. *Hum Genet* 1984;66:17.
- 9 Moser H, Emery AEH. The manifesting carrier in Duchenne muscular dystrophy. *Clin Genet* 1974;5:271–84.
- 10 Haginova K, Yamamoto K, Iinuma K, *et al.* Dystrophin immunohistochemistry in a symptomatic carrier of Becker muscular dystrophy. *J Neurol* 1991;238:375–8.
- 11 Tihy F, Vogt N, Recan D, *et al.* Skewed inactivation of an X chromosome deleted at the dystrophin gene in an asymptomatic mother and her affected daughter. *Hum Genet* 1994;93:563–7.
- 12 Dubowitz V. Myopathic changes in a muscular dystrophy carrier. *J Neurol Neurosurg Psychiatry* 1963;26:322.
- 13 Emery AEH. Clinical manifestations in two carriers of Duchenne muscular dystrophy. *Lancet* 1963;i:1126.
- 14 Pearson CM, Fowler WM, Wright SW. X-chromosome mosaicism in females with muscular dystrophy. *Proc Natl Acad Sci (USA)* 1963;50:24.
- 15 Hoffmann EP, Arahata K, Minetti C, Bonilla E, Rowland LP. Dystrophinopathy in isolated cases of myopathy in females. *Neurology* 1992;42:967–75.
- 16 Sewry CA, Sansome A, Clerk A, *et al.* Manifesting carriers of Xp21 muscular dystrophy; lack of correlation between dystrophin expression and clinical weakness. *Neuromusc Disord* 1993;3:141–8.
- 17 Lyon MF. Gene action in the X-chromosome of the mouse. (*Mus musculus* L). *Nature* 1961;190:372–4.
- 18 Minetti C, Chang HW, Medori R, *et al.* Dystrophin deficiency in young girls with sporadic myopathy and normal karyotype. *Neurology* 1991;41:1288–92.
- 19 Arikawa E, Hoffmann EP, Kaido M, Nonaka I, Sugita H, Arahata K. The frequency of patients with dystrophin abnormalities in a limb-girdle patient population. *Neurology* 1991;41:1491–6.
- 20 Ferrer X, Larivière M, Coquet M, Ellie E, Laguery A, Julien J. Syndrome des ceintures. Etude de 46 cas. *Rev Neurol* 1993;149:788–93.
- 21 Hoffmann EP, Kunkel LM, Angelini C, Clarke A, Johnson M, Harris JB. Improved diagnosis of Becker muscular dystrophy by dystrophin testing. *Neurology* 1989;39:1011–7.
- 22 Norman A, Coakley J, Thomas N, Harper P. Distinction of Becker from limb-girdle muscular dystrophy by means of cDNA probes. *Lancet* 1989;i:466–8.
- 23 Sunohara N, Arahata K, Hoffmann EP, *et al.* Quadriceps myopathy: forme fruste of Becker muscular dystrophy. *Ann Neurol* 1990;28:634–9.
- 24 Doriguzzi C, Palmucci L, Mongini T, Chiado-Piat L, Restagno G, Ferrone M. Exercise intolerance and recurrent myoglobinuria as the only expression of Xp21 Becker type muscular dystrophy. *J Neurol* 1993;240:269–71.
- 25 Gospe SM, Lazaro RP, Lava NS, Grootcholten PM, Scott MO, Fischbeck KH. Familial X-linked myalgias and cramps: a nonprogressive myopathy associated with a deletion in the dystrophin gene. *Neurology* 1989;39:1277–80.
- 26 Palmucci L, Doriguzzi C, Mongini T, *et al.* Dilating cardiomyopathy as the expression of Xp21 Becker type muscular dystrophy. *J Neurol Sci* 1992;111:218–21.
- 27 Shukle A, Hall CD, Bradley WG, Pendlebury WW. Congenital monomelic hypertrophy with progressive myopathy. *Arch Neurol* 1991;48:107–10.
- 28 Merlini L. Calf myopathy with a twist. *Neuromusc Disord* 1994;4:13–5.
- 29 Roberds SL, Leturcq F, Allamand V, *et al.* Missense mutations in the adhalin gene linked to autosomal recessive muscular dystrophy. *Cell* 1994;78:625–33.