Enhanced B7 costimulatory molecule expression in inflammatory human sural nerve biopsies

R Kiefer, F Dangond, M Mueller, K V Toyka, D A Hafler, H-P Hartung

Abstract

Objectives—To define the role of the costimulatory molecules B7–1 and B7–2 in inflammatory disorders of the peripheral nervous system. B7 molecules are essential for effective antigen presentation and may determine the differentiation of T cells into a Th-1 or Th-2 phenotype, thus modulating immune response and disease course.

Methods—Forty nine sural nerve biopsies from patients with neuroborreliosis, Guillain-Barré syndrome (GBS), chronic inflammatory demyelinating polyneuropathy (CIDP), CIDP variants and hereditary neuropathies, and those with no detectable abnormality were investigated. The expression of B7–1 and B7–2 mRNA and protein was investigated by polymerase chain reaction (PCR) and immunocytochemistry.

Results—B7–1 mRNA was strongly upregulated in both cases of neuroborreliosis, in two cases of GBS and one case of variant CIDP. Moderate to low levels were detected in the remaining GBS and CIDP biopsies and were rarely found in a non-inflammatory control group consisting of hereditary neuropathy and normal nerves. At the immunocytochemical level, strong expression of B7–1 protein was found in both neuroborreliosis cases, and moderate or low expression in six of eight GBS cases and seven of 17 CIDP cases investigated, whereas only one of five non-inflammatory control nerves showed staining, which was very weak. In neuroborreliosis, B7–1 protein was found very pronounced in epineurial infiltrates, whereas in GBS and CIDP, labelling was predominantly endoneurial and localised to putative macrophages. B7–2 mRNA and protein were expressed only at low levels in neuroborreliosis and selected autoimmune neuropathy cases, and were essentially absent from non-inflammatory controls.

Conclusions—B7 molecules are expressed in the peripheral nervous system and regulated during disease, and their presence in macrophages underlines the putative function of endoneurial macrophages as local antigen presenting cells in the immunopathology of peripheral nerve. B7–1 rather than B7–2 is preferentially upregulated, possibly promoting the induction of a Th-1-type T cell response within the nerve.

Keywords: B7; antigen presentation; peripheral neuropathy; macrophage

Guillain-Barré syndrome (GBS) and chronic inflammatory demyelinating polyneuropathy (CIDP) are inflammatory disorders of presumed autoimmune aetiology of the peripheral nervous system (PNS). Activated autoreactive T cells are probably of key importance in the initiation of disease although humoral immune responses play a part in pathogenesis.3 Activation of T cells not only requires presentation of processed immunogenic peptide fragments on the surface of local antigen presenting cells in the context of major histocompatibility complex (MHC) antigens to interact with the T cell receptor, but also additional antigen-non-specific costimulatory signals. One of these signals is provided through the interaction of the costimulatory molecules B7–1 and B7–2 on antigen presenting cells with their ligands CD28 and CTLA-4.4 Antigen presentation in the presence of B7 results in T cell activation whereas absence of signals from B7 molecules may induce anergy or immunosuppression. Furthermore, antigen presentation in the presence of B7–1 may drive T cells towards the Th-1 phenotype with expression of interleukin (IL)-2, interferon (IFN)–γ and tumour necrosis factor (TNF)–α whereas presentation in association with B7–2 may induce the Th-2 phenotype with predominant expression of IL-4;6 although recent in vitro experiments using antigen presenting cells deficient in B7–1 or B7–2 give less clear results.8 Finally, whereas interaction with CD28 on T cells results in activation, engagement of CTLA-4 may deliver an inhibitory signal to T cells and induce peripheral T cell tolerance.7

The expression of B7 molecules is not restricted to monocytes and macrophages but occurs on other immune cells including dendritic cells, T cells, and B cells.4 In the CNS, microglial cells are the predominant cell type expressing B7 molecules.10–11 In multiple sclerosis lesions, immune cells including putative microglial cells express B7–1 and B7–2,10–12 whereas in stroke B7–2 is exclusively expressed. In vitro, human microglia constitutively express B7–2 and display B7–1 with prolonged culture in a time dependent manner.13–15 Both molecules may also be upregulated on cytokine stimulation in vitro. By virtue of their ability to express B7, microglia may act as local antigen presenting cells in the brain and may play a critical part in lesion formation in multiple sclerosis. In the PNS however, the function of B7 molecules has not been defined. In sural nerve biopsies from patients with inflammatory and non-inflammatory neuropathies, MHC class II antigens have been found both on...
macrophages and Schwann cells, whereas in experimental animals, macrophages seem to be the major site of MHC II expression. Thus macrophages rather than Schwann cells are the likely antigen presenting cell in the CNS and, in addition to executing cytotoxicity towards myelin sheaths, they may be critical components in initiating an autoimmune response such as in GBS and CIDP. To date it is unknown whether and where B7 molecules are expressed in peripheral nerve and how the local microenvironment of axons and Schwann cells may influence B7–1 and B7–2 preponderance. We have therefore investigated the presence of B7–1 and B7–2 mRNA and protein in sural nerve biopsies from patients with inflammatory and non-inflammatory neuropathies.

**Methods**

**PATIENTS**

Sural nerve biopsies from 49 patients were examined. They were performed for diagnostic purposes with informed consent, and scientific investigations on these biopsies were performed under a research protocol approved by the University of Würzburg ethics committee. Most of these nerves were investigated in previous studies, and details of the patients fulfilled only part of the diagnostic criteria of the absence of motor symptoms and signs or disability Chalfont, England) in a total volume of 30 μl. Amplification was conducted for 33 cycles (B7–1 and B7–2) of the resulting first strand cDNA was amplified by PCR using 0.5 μg each of forward and reverse primer, 80 nM dNTP, 1,2 U Taq polymerase (Perkin Elmer, Foster City, CA, USA) and 0.08 μl (32P)dCTP (3000 Ci/mmol; Amersham, Little Chalfont, England) in a total volume of 30 μl. Amplification was conducted for 33 cycles (B7–1 and B7–2) or 30 cycles (β-actin), each of 60 seconds at 94°C for denaturation, 60 seconds at 60°C for annealing, and 90 seconds at 72°C for extension. For B7 amplification, the following primers were used: for B7–1: 5'-AGA AGA GGA CCA CCA TCT CTG-3' and 5'-CTT GTC AGT TTC CAG AAT ACA GAA GAT GGT C-3' for B7–2. PCR using 0.5 μg each of forward and reverse primer, 80 nM dNTP, 1,2 U Taq polymerase (Perkin Elmer, Foster City, CA, USA) and 0.08 μl (32P)dCTP (3000 Ci/mmol; Amersham, Little Chalfont, England) in a total volume of 30 μl. Amplification was conducted for 33 cycles (B7–1 and B7–2) of the resulting first strand cDNA was amplified by PCR using 0.5 μg each of forward and reverse primer, 80 nM dNTP, 1,2 U Taq polymerase (Perkin Elmer, Foster City, CA, USA) and 0.08 μl (32P)dCTP (3000 Ci/mmol; Amersham, Little Chalfont, England) in a total volume of 30 μl. Amplification was conducted for 33 cycles (B7–1 and B7–2) or 30 cycles (β-actin), each of 60 seconds at 94°C for denaturation, 60 seconds at 60°C for annealing, and 90 seconds at 72°C for extension. For B7 amplification, the following primers were used: for B7–1: 5'-AGA AGA GGA CCA CCA TCT CTG-3' and 5'-CTT GTC AGT TTC CAG AAT ACA GAA GAT GGT C-3' for B7–2. After amplification, the samples were subjected to 5% polyacrylamide gel electrophoresis, the gels were dried, and PCR products were visualised by autoradiography on Fuji RX x-ray film for 2–24 hours. Quantification of amplification products was achieved using a phosphorimager Betascope 603 Blot Analyzer (Betagen, Waltham, MA, USA). B7 mRNA content was expressed in arbitrary units as the mean of two independent experiments and semiquantitatively assessed after normalisation for β-actin mRNA content. In some experiments, specificity of non-radioactive PCR products was confirmed by Southern blotting and hybridisation with radioactively labelled internal oligonucleotides (for B7–1: 5'-AGA AAA TGG TGC TGA CTA TGA TGT CTG-3'; for B7–2: 5'-GTA TCA ATG TAT CAT CCA TCA CAA AAA GCC C-3').
IMMUNOCYTOCHEMISTRY

Ten micron thick cryostat sections of sural nerve specimens and human tonsil as positive control tissues were air dried and fixed for 1 minute each in 3.7% phosphate buffered formalin and 50%, 100%, and 50% acetone at room temperature. After blockade of non-specific binding with 10% bovine serum albumin, primary antibodies were applied overnight at 4°C in a humid chamber. Primary mouse anti human B7–1 and B7–2 antibodies, a gift from Dr Vijay Kuchroo, Harvard Medical School, were used at 5 µg/ml for purified antibody and 1:25 for hybridoma supernatant. Bound antibody was detected using secondary biotinylated antimouse antibody and avidin biotinylated peroxidase complex, both at 1:100 for 1 hour at room temperature (Dako, Hamburg, Germany), and 3,3’-diaminobenzidine as chromogen. Sections treated without primary antibody or equivalent concentrations of an irrelevant antibody served as negative controls. For semiquantitative assessment, the number and intensity of the immunocytochemical signals in the biopsy specimens was graded as strong (grade 3), moderate (grade 2), weak (grade 1), or absent (grade 0) without knowledge of the PCR results.

RESULTS

DETECTION OF B7–1 AND B7–2 MRNA BY PCR

Polymerase chain reaction amplification of cDNA derived from sural nerve biopsies demonstrated expression of B7–1 mRNA in a subgroup of biopsy specimens (fig 1). However, the degree of B7–1 mRNA expression showed marked variation between individual biopsies. A semiquantitative estimate, relating the amount of B7 PCR product to that of actin, disclosed strong B7–1 amplification signals in both cases of established neuroborreliosis, two of the 11 GBS cases, and in one of the eight cases with CIDP variant. Moderate to low expression was found in the remaining cases with GBS, CIDP (20 cases), and CIDP variants, and the non-inflammatory control group of normal and hereditary neuropathies (seven cases), and was virtually absent in some (fig 2 A).

By contrast, B7–2 mRNA was only rarely expressed. Low levels of B7–2 amplification products were found in both cases with neuroborreliosis, two of 11 cases with GBS, three of 20 cases with CIDP and four of the eight CIDP variant cases. No expression was found in the remaining cases including all seven patients with hereditary neuropathies or no abnormality (fig 2 A).

IMMUNOCYTOCHEMICAL DETECTION OF B7–1 AND B7–2

Twenty three monoclonal antibodies against human B7–1 and B7–2 were tested for their usefulness in immunocytochemistry. Seven out of 14 antibodies directed against B7–1 and two out of nine antibodies directed against B7–2 consistently labelled round cells in human tonsils serving as positive control tissue,23 with comparable staining patterns.

Figure 1  Representative PCR experiment to detect B7–1 mRNA in nerve biopsies showing upregulation of B7–1 mRNA in selected cases with inflammatory neuropathies. Variability of β-actin signal intensity is due to different amounts of tissue in the biopsy samples.

Figure 2  (A) Semiquantitative estimation of B7–1 and B7–2 mRNA content in sural nerve biopsies of two cases with neuroborreliosis (BORR), 11 cases of Guillain-Barré syndrome (GBS), 18 cases of chronic inflammatory demyelinating polyneuropathy (CIDP), eight CIDP variants (CIDP Var), and six non-inflammatory controls (CO). Results are mean values of two independent experiments normalised for β-actin mRNA content. (B) Semiquantitative estimation of B7–1 and B7–2 immunocytochemical staining intensities in sural nerve biopsies of two cases with neuroborreliosis, eight cases of GBS, 17 cases of CIDP, and six non-inflammatory controls. Grading was grade 3=strong, grade 2=moderate, grade 1=weak, and grade 0=absent.
B7–1 was strongly expressed in both cases with neuroborreliosis. Intense immunocytochemical reaction product was found in ramified cells in the epineurium often located within perivascular round cell infiltrates (fig 2 B and fig 3 C). Endoneurial expression was also marked but somewhat weaker and present in scattered cells throughout the endoneurium (fig 3 D). By morphology and distribution, these cells strongly resembled endoneurial and epineurial macrophages rather than T cells or other cellular components of the peripheral nerve. Immunocytochemical stains on neighbouring sections for T cells and macrophages confirmed similar staining patterns between B7–1 and macrophages, although fewer cells were stained with B7–1 antibody than with macrophage markers (not shown).

In six of eight GBS cases and seven of 17 CIDP cases investigated, B7–1 signals were detected but were weaker than in neuroborreliosis (fig 2 B). Expression in GBS was generally stronger than in CIDP. There was limited expression on scattered epineurial perivascular cells (fig 3 F), with occasional perivascular clusters of B7–1 positive cells. In the endoneurium, there consistently were numerous scattered cells with small ramifications or an elongated shape resembling endoneurial macrophages that were weakly or moderately labelled with B7–1 antibody (fig 3 G, H). Occasionally, rounded cell profiles in the vicinity of myelin sheaths were noted that might represent Schwann cells. Comparing the staining pattern with the neuroborreliosis cases, endoneurial staining was relatively more pronounced than epineurial staining in GBS and CIDP, whereas the opposite was true for the neuroborreliosis cases.

In the hereditary neuropathy cases studied, either no specific staining or only very occasional epineurial cells staining for B7–1 were found, with minimal or absent endoneurial staining (fig 2 B, fig 3 I).
B7–2 immunoreactivity was only very rarely found. In the neuroborreliosis cases, B7–2 was identified in round cell infiltrates (fig 3 E) and occasional epineurial ramified cells, with no definite staining within the endoneurium. In the other inflammatory neuropathies, no consistent endoneurial staining was demonstrable but unequivocal albeit weak staining for B7–2 was noticeable in two CIDP cases. Some labelled epineurial cells were also occasionally found which could also be seen in one of the hereditary neuropathy cases.

COMPARISON OF PCR AND IMMUNOHISTOCHEMICAL RESULTS
Twenty seven biopsies were available both for PCR and immunohistochemistry. In 21 of these biopsies PCR and immunohistochemical data were supportive of each other. In three samples, there were B7 PCR amplificates above background but no immunohistochemical signals, and in another three biopsies we found immunohistochemical signals with low or absent B7 mRNA.

Discussion
The initiation of a local immune response in the peripheral nerve as a prerequisite for autoimmune inflammation requires effective presentation of antigen to surmounting T cells. Local resident macrophages are main candidates for antigen presentation as they readily express MHC class II molecules both in a resting state and after activation. We now report that the costimulatory molecules B7–1 and to a lesser extent B7–2 may also be found in peripheral nerve and are regulated during autoimmune disease. These data demonstrate that local cells within the PNS may effectively present antigen to T cells enabling them to actively participate in the initiation and maintenance of an autoimmune response. They also underline the apparent immunocompetence of the PNS.

B7–1 mRNA and protein were not present in all biopsies but were found at high levels mainly in an infectious neuropathy caused by *Borrelia burgdorferi* and in a subset of cases with GBS, CIDP, and CIDP variants, with only low levels in others and complete absence in some. In interpreting these data, it should be noted that due to the low amounts of RNA available no proper quantitative PCR with internal competitors could be performed, and that immunocytochemical quantification reflects no more than estimates as the stains were generally too weak to allow for reliable numerical quantification of labelled cells. However, immunocytochemical and PCR data are largely consistent and supportive of each other, although mRNA was occasionally found in biopsies that were negative at the immunocytochemical level, and vice versa. This might be due to differences in sensitivity or sampling errors, or due to B7 mRNA present in contaminating blood cells within the biopsy samples. It should further be noted that immunocytochemistry, although weak, was well reproduced within individual nerves providing reliable results, whereas PCR quantification was less controlled due to lack of an internal competitor. Despite these technical considerations it seems clear that B7–1 mRNA and particularly B7–1 protein are upregulated in several inflammatory neuropathies compared with subjects with normal nerves and patients with hereditary neuropathy. B7–1 expression, particularly in the epineurium, seemed to be more pronounced in inflammatory infiltrates from patients with more acute neuropathies of short duration such as neuroborreliosis and GBS. This is consistent with our findings in multiple sclerosis where B7–1 was upregulated particularly in acute plaques from early disease cases. However, it should be noted that the absence of B7 in other inflammatory biopsy specimens does not necessarily reflect true absence because of the well known sampling issues involved in obtaining peripheral nerve biopsy specimens. Thus B7 expression may go undetected even if present elsewhere or at another time in the same patient. In non-inflammatory controls, only minimal immunocytochemical signal for B7–1 protein was present in one single biopsy whereas it was absent in the others. Low levels of B7–1 PCR amplification product in these biopsies may again either reflect the higher sensitivity of the method or the amplification of mRNA derived from contaminating blood cells inevitably present in the specimens. In addition, inflammatory infiltrates may occasionally be present in hereditary neuropathies. Similarly, B7–2 was noted in biopsies from patients with inflammatory neuropathies but was virtually absent from non-inflammatory controls. Unfortunately, only two normal nerves were available for comparison as such nerves usually are not being biopsied.

The preferential overexpression of B7–1 in GBS and CIDP, in addition to the neuroborreliosis cases, emphasises the immunoinflammatory aetiology of these conditions. Interestingly, expression was particularly strong epineurially in neuroborreliosis and more conspicuous endoneurally in GBS and CIDP. These differences may reflect the actual site of the inflammatory response, which is an epineurial perivasculitis in neuroborreliosis and an immune response against myelin and other endoneural components in the case of GBS and CIDP. Although published data on MHC II expression were not reproduced in the present study, it seems that B7 expression does not correlate closely with MHC II expression as MHC II positive macrophages are also abundantly present in degenerative neuropathies. MHC II expression may thus be part of a more general activation programme of macrophages, much as it is used by activated microglial cells of the CNS which readily express MHC II molecules once activated. By contrast, B7 expression may be more restricted and specific for inflammatory conditions where it may be required as a costimulatory signal in antigen presentation. Thus as a preliminary speculation, B7 expression might even be useful as a diagnostic marker to differentiate inflammatory neuropathies from degenerative conditions, as has been described for other inflammatory macrophage markers.
Macrophages seemed to be the main source of B7 molecules in the present study. This assumption is based on the typical morphology and distribution of B7 positive cells both epineurally and endoneurally. It is additionally supported by colocalisation of B7–1 and the macrophage antigen CD 68 in some double immunofluorescence experiments not shown in the present paper as B7–1 immunofluorescence was too weak to be unequivocally reproduced and photodocumented. In addition, some occasional B7–1 positive putative Schwann cells were seen, and B7–2 positive round cells might either represent macrophages, T cells, or B cells. Thus, B7 expression was found on those cells already known to express B7 molecules in vitro and present antigen, and those expressing MHC II in peripheral nerve—namely, macrophages.

Our data thus further support the accepted notion that macrophages rather than Schwann cells are the main immunocompetent cell of the PNS. In most biopsies, B7–1 mRNA and protein were much stronger and more widely expressed than B7–2. These findings differ somewhat from in vivo studies in multiple sclerosis where B7–2 staining was also widely found. In an from in vivo studies in multiple sclerosis where B7–1 mRNA and protein were much stronger and more widely expressed than B7–2. These findings differ somewhat from in vivo studies in multiple sclerosis where B7–2 staining was also widely found.

31 Satoh J, Lee YB, Kim SU. T-cell costimulatory molecules B7–1 (CD80) and B7–2 (CD86) are expressed in human microglia but not in astrocytes in culture. Brain Res 1995;704:92–6.

This work was supported by grants from NIH (NS24247 to DAIH, Deutsche Forschungsgemeinschaft (Ki 532/3–1), IWF Munich Medical Faculty (IMF program), and Gemeinschaftliche Herte-Stiftung, and University Research Funds of the University of Würzburg. We thank Dr Vijay Kuchroo, Harvard Medical School, Boston, for kindly supplying us with anti-B7–1 and anti-B7–2 monoclonal antibodies, and Heidrun Pischel and Anne Stöber for skillful technical assistance. Early parts of this study were performed while RR and HPH were at the Department of Neurology, University of Würzburg, Germany.