Tourette’s syndrome: a cross sectional study to examine the PANDAS hypothesis

A J Church, R C Dale, A J Lees, G Giovannoni, M M Robertson

Background: The classical neurological disorder after group A β haemolytic streptococcal infection is Sydenham’s chorea. Recently a tic disorder occurring after group A streptococcal infection has been described and termed PANDAS (paediatric autoimmune neuropsychiatric disorders associated with streptococcal infection). It is proposed that antibodies induced after group A streptococcal infection react with basal ganglia neurones in Sydenham’s chorea and PANDAS. Anti-basal ganglia antibodies (ABGA) are present in most cases of acute Sydenham’s chorea, but rarely in controls.

Objective: To investigate the hypothesis that Tourette’s syndrome may be associated with group A streptococcal infection and ABGA.

Methods: 100 patients with Tourette’s syndrome (DSM-IV-TR) were enrolled in a cross sectional study. Children with neurological disease (n = 50) and recent uncomlicated streptococcal infection (n = 40), adults with neurological disease (n = 50), and healthy adults (n = 50) were studied as controls. Recent group A streptococcal infection was defined using antistreptolysin O titre (ASOT). ABGA were detected using western immunoblotting and indirect immunofluorescence.

Results: ASOT was raised in 64% of children with Tourette’s syndrome compared with 15% of paediatric neurological disease controls (p < 0.0001), and in 68% of adults with Tourette’s syndrome compared with 12% of adult neurological controls and 8% of adult healthy controls (p < 0.05). Western immunoblotting showed positive binding in 20% of children and 27% of adults with Tourette’s syndrome, compared with 2–4% of control groups (p < 0.05). The most common basal ganglia binding was to a 60 kDa antigen, similar to the proposed antigen in Sydenham’s chorea. Indirect immunofluorescence revealed autoantibody binding to basal ganglia neurones. Serological evidence of recent group A streptococcal infection, assessed by a raised ASOT, was detected in 91% (21/23) of Tourette’s Syndrome patients with positive ABGA compared with 57% (44/77) with negative ABGA (p < 0.01).

Conclusions: The results support a role of group A streptococcal infection and basal ganglia autoimmunity in a subgroup of patients with Tourette’s syndrome and suggest a pathogenic similarity between Sydenham’s chorea and some patients with Tourette’s syndrome.

Abbreviations: ABGA, anti-basal ganglia antibodies; ADHD, attention deficit hyperactivity disorder; ASOT, antistreptolysin O titre; DSM, Diagnostic and Statistical Manual of Mental Disorders; ICD, International Classification of Diseases; PANDAS, paediatric autoimmune neuropsychiatric disorders associated with streptococcal infection.
controversial. The proposed disease mechanism in Sydenham's chorea and PANDAS is cross reactive antibodies induced by group A streptococcal infection that bind specifically to basal ganglia antigens. The presence of anti-basal ganglia antibodies (ABGA) in Sydenham's chorea and PANDAS supports this hypothesis.

We aimed to examine the association of streptococcal infection and basal ganglia autoimmunity in Tourette's syndrome by screening for recent streptococcal infection and ABGA in a large cohort of children and adults with this syndrome.

METHODS

Patients

Permission for the study was obtained from the local ethics committee of the National Hospital for Neurology and Neurosurgery. Index patients were interviewed by one of us (MMR) using standardised instruments including the National Hospital interview schedule, the diagnostic interview schedule, and the Yale global tic severity rating scale. To be diagnosed as having Tourette's syndrome, patients had to satisfy DSM-IV-TR (APA, 2000) and ICD-10 (WHO, 1992) criteria. Thus they had to have multiple motor and one or more vocal tics, with symptoms lasting longer than a year. Comorbid diagnosis of obsessive-compulsive disorder and ADHD conformed to DSM-IV and ICD-10 criteria.

Controls

To determine the significance of both streptococcal serology and anti-basal ganglia antibodies, we enrolled controls for comparison (table 1), as follows:

- **Children with neurological disease** (n = 50). This group contained patients with dystonia (n = 30) of inflammatory (n = 12), vascular (n = 6), metabolic (n = 6), and genetic (n = 2) aetiology. Additional dystonic aetiologies included variant Creutzfeldt-Jakob disease, basal ganglia tumour, athetoid cerebral palsy, and juvenile Parkinson's disease (n = 1 each). Also within the neurology control group were patients with invasive viral encephalitis (n = 20).

- **Children with recent uncomplicated streptococcal infection** (n = 40).

- **Adults with neurological disease** (n = 50), including multiple sclerosis (n = 12), dementia (n = 7), paraneoplastic syndromes (n = 3), autoimmune neuropsychiatry (n = 3), acute cerebellitis (n = 3), and other mixed neurological disease.

- **Healthy adults from laboratory staff and paediatric hospital workers** (n = 50).

The Tourette's syndrome patients and controls were recruited during the same time period and blood samples were stored at −80°C with identification data coded.

**Streptococcal serology**

Evidence of recent streptococcal infection was examined using antistreptolysin O titres (ASOT). All patient and control samples were analysed using standardised Dade Behring II nephelometry; an ASOT above 200 IU/ml is considered to indicate recent infection (WHO guidelines).

### Table 1 Demographics of the sample

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Mean age (years)</th>
<th>Sex (M/F)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Child Tourette syndrome</td>
<td>56</td>
<td>12.8 (8 to 17)</td>
<td>40/16</td>
</tr>
<tr>
<td>Adult Tourette syndrome</td>
<td>44</td>
<td>37.8 (18 to 61)</td>
<td>32/12</td>
</tr>
<tr>
<td>Child streptococcal infection</td>
<td>40</td>
<td>9.8 (2 to 15)</td>
<td>25/15</td>
</tr>
<tr>
<td>Child neurological disease</td>
<td>50</td>
<td>7.6 (0.5 to 18)</td>
<td>24/26</td>
</tr>
<tr>
<td>Adult healthy controls</td>
<td>50</td>
<td>35.6 (19 to 57)</td>
<td>25/25</td>
</tr>
<tr>
<td>Adult neurological disease controls</td>
<td>50</td>
<td>41.1 (19 to 70)</td>
<td>30/20</td>
</tr>
</tbody>
</table>

F, female; M, male.

Basal ganglia homogenate

Caudate, putamen, and globus pallidus from a patient with no evidence of neurological disease was kindly provided by the Queen Square brain bank for neurological disorders, Institute of Neurology, London. The block of tissue was homogenised with a small volume of saline containing protease inhibitors (Sigma Chemicals, Poole, Dorset, UK) and centrifuged for 30 minutes at 7500 × g. Equal volumes of supernatant and Di-isopropyl ether were mixed and centrifuged at 600 × g for 10 minutes to remove lipid from the supernatant. The protein fraction was then collected and stored at −80°C until required.

**Basal ganglia antibody western immunoblotting**

As previously described, the basal ganglia homogenate was mixed with lithium dodecyl sulphate sample buffer (Invitrogen, USA), containing 0.05 M dithiothreitol and heated at 65°C for 15 minutes. A total of 30 µg of protein was loaded onto a 4–12% bis-tris gel (Invitrogen, Paisley, Scotland, UK) and electrophoresed. The proteins were blotted onto nitrocellulose (Sartorius, Epsom, Surrey, UK) and blocked with 2% milk proteins for two hours. Samples were diluted 1/300, applied to the blot, and incubated overnight at 4°C. The nitrocellulose was washed with 10 changes of 0.9% saline containing 0.2% milk proteins and 0.025% Tween. The blot was incubated for two hours with rabbit anti-human IgG conjugated with horseradish peroxide diluted 1/1000 (Dako, Cambridge, UK). After washing, the substrate 4-chloro-1-naphthol (Sigma) was added and the blot was allowed to develop for 15 minutes. Western immunoblotting was done in all patients and controls.

**Basal ganglia antibody indirect immunofluorescence**

Indirect immunofluorescence was undertaken in 10 western immunoblot positive Tourette's syndrome patients and 10 controls from each group. Using previously described methods, 10 µm sections of normal human basal ganglia were cut from a snap frozen tissue block of caudate and putamen. Control and patient sera were diluted 1:10 in phosphate buffered saline (PBS), overlaid onto prepared slides, and incubated for 30 minutes. A control slide was also prepared and incubated with PBS alone for 30 minutes to assess background fluorescence. All slides were washed in PBS and incubated with anti-human IgG conjugated with fluorescein isothiocyanate (Dako) and analysed using a fluorescence microscope.

**Statistical analysis**

All statistical analyses were done using SAS software (SAS Institute Inc, Cary, North Carolina, USA). Streptococcal serology was compared using the non-parametric two sample exact Wilcoxon rank-sum test. Positive ABGA Western immunoblotting was compared in each subgroup using χ² tests. Ninety-five per cent confidence intervals (CI) are given.

**RESULTS**

**Demographics**

Demographic data on the patients are summarised in table 1. The serology results were analysed in paediatric and adult
groups separately, because of the higher incidence of streptococcal infection in childhood. The Tourette’s syndrome cohort had a mean (SD) diagnostic confidence index of 64 (20)%, range 14% to 100%. The prevalences of obsessive-compulsive disorder and ADHD in the total cohort were 31% and 43%, respectively. The paediatric Tourette’s syndrome patients had a diagnostic confidence index of 63 (21)%, range 14% to 100%, a 17% incidence of obsessive-compulsive disorder, and a 49% incidence of ADHD. The adult Tourette’s syndrome patients had a diagnostic confidence index of 63 (20)%, range 16% to 100%, a 51% incidence of obsessive-compulsive disorder, and a 31% incidence of ADHD in childhood.

**Streptococcal serology**

**Paediatric**

As predicted, ASOT was raised in 80% of the children with recent streptococcal infection (mean 349 IU/ml (95% CI, 270 to 427)). ASOT was also raised in 64% of the children with Tourette’s syndrome (299 IU/ml (262 to 335)) and in 18% of the children with neurological disease (151 IU/ml (101 to 201)). ASOT was raised in the paediatric Tourette’s syndrome cohort compared with the neurological controls (p < 0.0001), but not with the streptococcal controls (p = 0.4).

**Adult**

ASOT was raised in 68% of the adults with Tourette’s syndrome (mean 2981 IU/ml (95% CI, 243 to 353)), in 12% of the adults with neurological disease (140 IU/ml (92 to 188)), and in 8% of the healthy adults (122 IU/ml (101 to 143)). ASOT was statistically raised in the adult Tourette’s syndrome patients compared with the neurological controls (p < 0.05) and the healthy controls (p < 0.05).

**ABGA western immunoblotting**

**Paediatric**

Twenty per cent of the paediatric Tourette’s syndrome group (12/56) had positive western immunoblotting, compared with 4% of the neurological controls (1/50) and 2% of the streptococcal controls (1/40). The difference between the Tourette’s syndrome group and both control groups was significant (p < 0.05 and p < 0.05, respectively). Common bands of reactivity were seen in the Tourette’s syndrome group, with IgG binding to large basal ganglia neurones (fig 1). The common antibody binding was also to 60 kDa (n = 7), 40 kDa (n = 5), 45 kDa (n = 2), 80 kDa (n = 2), 67 kDa (n = 1), and 98 kDa antigens (n = 1). The positive neurological control bound to a 40 kDa antigen. The positive healthy controls bound to a 55 kDa antigen. In all paediatric and adult Tourette’s syndrome patients the most common basal ganglia autoantigen detected was to a 60 kDa protein (n = 13) and then to a 40 kDa protein (n = 9).

**ABGA indirect immunofluorescence**

To define the localisation of antibody binding, we carried out indirect immunofluorescence on 10 Tourette’s syndrome patients (five paediatric and five adult) with positive western immunoblotting. All patients had the same binding pattern, with IgG binding to large basal ganglia neurones (fig 2). The
immunofluorescence staining pattern seen using these Tourette’s syndrome patients was identical to that previously described in Sydenham’s chorea. None of the controls tested had reactivity against any cellular component of the basal ganglia.

Raised streptococcal serology and positive ABGA Ninety one per cent of Tourette’s syndrome patients with positive ABGA western immunoblotting (21/23) had a positive ASOT, compared with 57% of Tourette’s syndrome patients with negative ABGA western immunoblotting (44/77) (p < 0.01).

Clinical comparison between ABGA positive and ABGA negative groups The clinical characteristics (obsessive-compulsive disorder/ADHD and family history of neuropsychiatric disease) did not differ between patients who were ABGA positive or ABGA negative (table 2). The numbers of patients with a significantly raised ASOT were greater in the ABGA positive group (91.3%) than in the ABGA negative group (58.4%). We were able to retest the blood of four of the Tourette’s syndrome patients who had negative ABGA and positive ASOT 3.5 months after first testing. This showed that ASOT levels had decreased to normal in three cases but ABGA had become positive in all four (table 3).

**Table 2 Clinical comparisons between ABGA positive and ABGA negative Tourette’s syndrome patients**

<table>
<thead>
<tr>
<th>Finding</th>
<th>ABGA positive (n=24)</th>
<th>ABGA negative (n=77)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at first tic (years) (range)</td>
<td>6.56 (2 to 13)</td>
<td>6.81 (2 to 16)</td>
<td>NS</td>
</tr>
<tr>
<td>Diagnostic confidence index (mean [SD])</td>
<td>62 [20%] [36% to 96%]</td>
<td>64 [21%] [14% to 100%]</td>
<td>NS</td>
</tr>
<tr>
<td>Obsessive-compulsive disorder (mean [SD])</td>
<td>27%</td>
<td>33%</td>
<td>NS</td>
</tr>
<tr>
<td>Attention deficit hyperactivity disorder (mean [SD])</td>
<td>22%</td>
<td>50%</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>History of neuropsychiatric disease in 1st degree family member (mean [SD])</td>
<td>69.6%</td>
<td>753%</td>
<td>NS</td>
</tr>
<tr>
<td>ASOT &gt;200 IU/ml</td>
<td>91.3%</td>
<td>584%</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

*By western immunoblotting.
†Family history of neuropsychiatric disease includes tic disorders, Tourette’s syndrome, and obsessive-compulsive disorder.
ABGA, anti-basal ganglia antibodies; ASOT, antistreptolysin O titre.

**Table 3 Follow up studies**

<table>
<thead>
<tr>
<th>Case</th>
<th>Test</th>
<th>1st Test result</th>
<th>2nd Test result</th>
<th>Time after 1st test (months)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Case 1</td>
<td>ASOT</td>
<td>500 IU/ml Negative</td>
<td>133 IU/ml 60 kDa</td>
<td>3</td>
</tr>
<tr>
<td>Case 2</td>
<td>ASOT</td>
<td>260 IU/ml Negative</td>
<td>185 IU/ml 45 and 60 kDa</td>
<td>3.5</td>
</tr>
<tr>
<td>Case 3</td>
<td>ASOT</td>
<td>710 IU/ml Negative</td>
<td>220 IU/ml 40 kDa</td>
<td>4</td>
</tr>
<tr>
<td>Case 4</td>
<td>ASOT</td>
<td>315 IU/ml Negative</td>
<td>&lt;50 IU/ml 40 and 95 kDa</td>
<td>3</td>
</tr>
<tr>
<td>Case 5</td>
<td>ASOT</td>
<td>500 IU/ml Negative</td>
<td>133 IU/ml 60 kDa</td>
<td>3.5</td>
</tr>
<tr>
<td>Case 6</td>
<td>ASOT</td>
<td>260 IU/ml Negative</td>
<td>185 IU/ml 45 and 60 kDa</td>
<td>4</td>
</tr>
<tr>
<td>Case 7</td>
<td>ASOT</td>
<td>710 IU/ml Negative</td>
<td>220 IU/ml 40 kDa</td>
<td>4</td>
</tr>
<tr>
<td>Case 8</td>
<td>ASOT</td>
<td>315 IU/ml Negative</td>
<td>&lt;50 IU/ml 40 and 95 kDa</td>
<td>3</td>
</tr>
</tbody>
</table>

ABGA, anti-basal ganglia antibodies; ASOT, antistreptolysin O titre.

DISCUSSION

The Tourette’s syndrome patients were recruited from a dedicated clinic with all patients fulfilling strict diagnostic criteria. The mean age of onset (6.75 years) and the male predominance were consistent with previously published data. Similarly, 100% of the patients in this study presented in childhood (< 18 years), which is characteristic of Tourette’s syndrome. As longitudinal studies have shown that 50% of patients will be free of tics by 18 years of age, and tic severity gradually diminishes in adulthood, it is likely that this cohort of adult patients with Tourette’s syndrome represents a severe form of the disease.

The pathogenesis of Tourette’s syndrome remains obscure but it is considered to be an inherited neurodevelopmental disorder resulting in disinhibition of the cortico-striatal-thalamic-cortical circuitry. Although various neurotransmitters have been implicated in the pathogenesis of the syndrome, the most favoured hypothesis is that there are subtle abnormalities of the dopaminergic system. Tourette’s syndrome appears to be a familial disorder, with early reports supporting an autosomal dominant inheritance. As a genetic basis for Tourette’s syndrome has yet to be uncovered, alternative pathological models are being considered, and Tourette’s syndrome may turn out to be a heterogeneous disorder.

The recognition that PANDAS may be pathologically related to Sydenham’s chorea, along with the clinical similarity of PANDAS and Tourette’s syndrome, has led to the notion that Tourette’s syndrome may occur as a result of basal ganglia dysfunction secondary to post-streptococcal autoimmunity. The observation that patients with tic disorders or Tourette’s syndrome are more likely to have positive streptococcal serology than control subjects supports this hypothesis, though not all studies have replicated such an association. Preliminary analysis of antibody reactivity to streptococcal M proteins has shown raised titres to M12 and M19—but not to M1, M4, and M6—in 25 adult patients with Tourette’s syndrome.
syndrome compared with 25 control subjects. This may suggest that only certain strains of group A streptococcal infection could be linked to Tourette’s syndrome. This is compatible with observations that other post-streptococcal immune mediated disorders, such as acute rheumatic fever and glomerulonephritis, are also strain specific. The proposal that Sydenham’s chorea and PANDAS occur as the result of an immune mediated insult is supported by the presence of ABGA which bind to basal ganglia neurons. In a study of patients with Sydenham’s chorea, we have recently shown that ABGA were present in all 20 patients (100%) with acute disease and in 11/16 patients (69%) with persistent disease, but in none of a group of 11 healthy controls. Increased autoantibody binding to putamen in Tourette’s syndrome and identification of basal ganglia antigens of molecular weights 60, 67, and 83 kDa, have been reported. A further study also suggested that 60 kDa was the most prevalent basal ganglia autoantigen in Tourette’s syndrome, although a complex multivariate analysis is required to establish this association. Preliminary results from another group also suggested that a 60 kDa and an 83 kDa antigen represented the dominant response in patients with tics, Tourette’s syndrome, or obsessive-compulsive disorder, and the same investigators suggested that these antibodies may cross-react with a calpain-calpastatin complex. In Sydenham’s chorea, we have described three dominant basal ganglia autoantigens of molecular weights 40, 45, and 60 kDa. The consistent finding of a 60 kDa antigen from this study may suggest that the antigen could be important in Sydenham’s chorea, PANDAS, and Tourette’s syndrome.

Not all of the studies reported have had similar findings to ours. Two studies using a neuroblastoma cell line rather than basal ganglia as the antigen source found no discriminate antibody responses in Tourette’s syndrome. It would appear from these studies that human or mammalian brain homogenates should be the preferential source of antigen. The methods for detecting antibodies may also influence the results from different studies; enhanced chemiluminescence is a very sensitive method but produces a multitude of responses in western immunoblotting of patients and controls, most of which are of doubtful significance. We propose that colorimetric analysis of western blots in this setting is preferable and improves specificity, although arguably at the cost of reduced sensitivity.

Using the methods described we found a correlation between ABGA and positive ASOT. Serological evidence of recent group A streptococcal infection was detected in 91% of patients with positive ABGA and in 57% with negative ABGA. This may suggest that ABGA and recent streptococcal infection are related in some patients. Interestingly the prevalence of raised ASOT in the ABGA negative patients was significantly more common than in the control groups. This finding suggests that post-streptococcal autoimmunity could be important in a larger proportion of patients with Tourette’s syndrome. It is possible that ABGA reactivity is be a phenomenon that waxes and wanes with the clinical course. Indeed three of four positive antibodies we followed up with raised ASOT but negative ABGA had positive ABGA and decreased ASOT three months after the initial sample was taken. A longitudinal study is required to investigate the temporal association between ABGA, streptococcal serology, and clinical course.

An alternative explanation for ABGA in Sydenham’s chorea, PANDAS, and Tourette’s syndrome is that it occurs as an epiphenomenon secondary to basal ganglia damage. We believe this is unlikely, for three reasons. First, control patients with inflammatory, metabolic, and ischemic basal ganglia diseases uncommonly develop these antibodies. Second, in a provocative study by Hallett and coworkers, the microinjection of sera or IgG from patients with Tourette’s syndrome into the striatum of rats induced stereotypes analogous to the involuntary movements seen in Tourette’s syndrome, suggesting that ABGA may play a pathogenic role. Finally, a randomized trial of therapeutic plasma exchange and intravenous immunoglobulin for PANDAS showed a therapeutic response in favour of the treated patients.

Conclusions

We have shown that a significant proportion of patients with Tourette’s syndrome have evidence of recent streptococcal infection and anti-basal ganglia antibodies. The studies published so far on these antibodies have used immuno-fluorescence, western immunoblotting, and enzyme linked immunosorbent assay methods, which have different sensitivities and specificities. This may help explain the differences between studies in the reported associations between ABGA and disease. Many investigators have proposed that the presence of a 60 kDa autoantigen in Sydenham’s chorea, PANDAS, and Tourette’s syndrome is significant. The identification of this antigen is crucial in determining the pathogenicity of ABGA in post-streptococcal CNS syndromes.

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REFERENCES

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