Objectives: The most common familial early onset dementia mutations are found in the genes involved in Alzheimer’s disease; the amyloid precursor protein (APP) and the presenilin 1 and 2 (PSEN1 and 2) genes; the prion protein gene (PRNP) may be involved.

Methods: Following identification of a two-octapeptide repeat insertion in PRNP, we conducted a meta-analysis to investigate the relation of number of PRNP octapeptide repeats with age at disease onset and duration of illness; identifying 55 patients with PRNP octapeptide repeat insertions. We used a linear mixed effects model to assess the relation of number of repeats with age at disease onset, and studied the effect of the number of inserted octapeptide repeats on disease duration with a Cox proportional hazards regression analysis.

Results: We found an increasing number of repeats associated with younger age at onset (p<0.001). Duration of the disease decreased significantly with the length of the octapeptide repeat (p<0.001) when adjusting for age at onset.

Conclusions: Our findings show significant inverse associations of the length of the PRNP octapeptide repeat with age at disease onset and disease duration in the spongiform encephalopathies.

Several genes are known in familial early onset dementias. The most common mutations are found in the genes involved in Alzheimer’s disease. This concerns mutations in the amyloid precursor protein (APP) and the presenilin 1 and 2 (PSEN1 and 2) genes. Further, mutations in the prion protein gene (PRNP) may be common in early onset dementia.1 Mutations in PRNP may lead to different clinical phenotypes, including familial Creutzfeldt–Jakob disease (CJD), Gerstmann–Sträussler–Scheinker disease (GSS), and fatal familial insomnia.2,3 These mutations can be missense mutations, deletions, or insertions.2,3 Insertions are only found in the octapeptide repeat region located between codons 51 and 91 of PRNP, which normally contains one nonapeptide and four octapeptide repeats.3 The insertions identified so far comprised one, two, and four to nine extra octapeptide repeats.4 Expansion of an octapeptide repeat or 24-nucleotide repeat region is rare, but expansion of trinucleotide repeats are found in other neurodegenerative disorders, including Huntington’s disease and the spinal cerebellar ataxias.10-12 The expansion of the polyglutamine tract in these disorders is associated with accumulation and higher toxicity of the protein.13 For several disorders with an expanded polyglutamine repeat an inverse correlation was found between repeat length and age at onset.14 15 The relation between the numbers of octapeptide repeats and onset of disease in human spongiform encephalopathies has not been studied.

We sequenced the coding region of APP, PSEN1, PSEN2, and PRNP in a population based sample of 17 patients with early onset dementia and identified in one patient an insertion of two-octapeptide repeats. We present a meta-analysis in which we studied the relation of the number of inserted repeats with age at onset and duration of disease in all patients with PRNP octapeptide repeat insertions reported in the literature to date.

Methods
Since 1998 we have been studying patients with early onset dementia in a defined geographical region in the northern part of The Netherlands. A medical doctor visited the patients and a member of the research team and a neurologist, using a standardised protocol based on the DSM III-R criteria, independently confirmed the diagnosis of dementia.16 After obtaining written informed consent from a relative, blood was drawn for DNA collection. In this study, 17 patients with early onset dementia were extensively studied for mutations in genes known to be involved in dementia. Exons 16 and 17 of the APP gene, and exons 3–12 of the PSEN1 and 2 genes were analysed by direct sequencing.17 The single coding exon (exon 2) of PRNP was analysed by direct sequencing of three overlapping PCR fragments (A, B, and C) spanning the coding region.18 In one patient, visualisation of the PCR fragments on a 1.5% agarose gel stained with ethidium bromide demonstrated an insertion as an extra band of about 100 bp. To further characterise the insertion we cloned the PCR fragments in the pCR4-TOPO vector (Invitrogen, Carlsbad CA). Clones carrying the insertion were selected by colony PCR and the insert was sequenced using vector primers.

Meta-analysis
We used a Medline search with a combination of the words “mutation”, “insertion”, “octapeptide”, or “repeats”, and “prion”, “PRNP”, “Creutzfeldt-Jakob disease”, “Gerstmann–Sträussler–Scheinker disease”, or “spongiform encephalopathy” to identify all patients with octapeptide repeat insertions in the PRNP gene. We included patients suspected of a prion disease with an octapeptide repeat extension disclosed by DNA analysis, patients with postmortem examination supporting the diagnosis of spongiform encephalopathy and a relative with an octapeptide repeat insertion, and patients with both postmortem examination and DNA analysis. Patients from PRNP insertion families

Abbreviations: APP, amyloid precursor protein; CJD, Creutzfeldt-Jakob disease; GSS, Gerstmann–Sträussler–Scheinker disease; PRNP, prion protein gene; PSEN1 and 2, presenilin 1 and 2; REML, restricted maximum likelihood.
suffering from an unidentified neurodegenerative disease, without PRNP analysis or postmortem examination, were not included.

For statistical analyses, we used the log transformed age at onset to obtain a normally distributed outcome. A linear mixed effects model fitted by restricted maximum likelihood (REML) was used to assess the relation between number of inserted octapeptide repeats and age at disease onset, and to estimate the part of the between family variance of age at onset explained by PRNP codon 129 and number of inserted octapeptide repeats. Analyses were performed adjusted and unadjusted for the PRNP codon 129 genotype. To study the effect of the number of inserted octapeptide repeats on disease duration we performed a Cox proportional hazards regression analysis, adjusted for age at onset. PRNP codon 129 was not included in this model because of the limited number of independent observations. Because we observed an outlier in the first 6 months, analyses were performed by conditioning on surviving the first 6 months. Robust standard errors were used that take into account dependence of the data.

RESULTS
Mutation screening and clinical phenotype
In this sample of 17 patients with early onset dementia, we found two mutations in genes known to be involved in dementia. Firstly, we observed a single base change predicting an amino acid change at codon 301 (T301M) in the PSEN2 gene. The patient carrying this mutation experienced a lack in short and long term memory just after 60 years of age. Within a couple of years the condition of the patient deteriorated and the patient died more than 10 years after onset of the symptoms, with the clinical diagnosis of Alzheimer’s dementia. The family history revealed early and late onset dementia of unknown type in several family members. However, in the cell transfection experiments there is no evidence up until now that this PSEN2 mutation is functional. The β amyloid 42/40 ratio was not altered in cells carrying the T301M mutation compared to cells carrying the wild type allele. Although this does not exclude the possibility that the mutation is functional, it is unlikely, as all functional PSEN1 and 2 mutations known to date have an effect on the β amyloid 42/40 ratio.

The second mutation found was a two-octapeptide repeat insert in the PRNP gene. The patient carrying this mutation developed an atypical dementia, the age of first symptoms being in the late 50s. In the following 10 years, the condition of the patient slowly deteriorated into a mute state. Myoclonus, cerebellar, and visual disturbances were absent. Electroencephalography did not show the periodic sharp wave complexes typical for CJD. Atypical dementia of late onset was reported in one first degree relative.

The octapeptide repeat insertion in PRNP contained an identical double repeated 24-nucleotide sequence located between the fourth (normally R3) and fifth (R4) repeat and included a silent nucleotide substitution from GGG to GGA in the seventh triplet (coding for glycine in both cases). The nucleotide sequence in the third between the fourth (R3) and fifth (R4) repeats is identical to that described by van Harten et al in a patient of Dutch origin (R1 R2 R2a R2 R2a R2a R4). The normal order is R1 (the nonapetide repeat) R2 R3 R4. The patient was heterozygous for the polymorphic PRNP codon 129, carrying the valine (V) allele on the chromosome with the octapeptide repeat insertion, which is identical to the disease haplotype in the patient described by Van Harten.

Genealogical data from our patient and the patient described by Van Harten et al were collected using the municipal registers. We were able to trace back the ancestors of both patients for six generations, until 1780, but it was not possible to link both patients to a common ancestor within these six generations. Genealogical data were complete, except for one branch in the fifth generation of the patient described here, for which no information was available. The dead end in one branch in the fifth generation, non-paternity, or a common ancestor before 1780 may explain our failure to establish a genealogical link.

Meta-analysis of studies of PRNP octapeptide repeat insertions
Twenty four articles reporting PRNP octapeptide repeat insertions in 22 different families were identified. In total 55 patients fulfilled our inclusion criteria and had sufficient information on clinical characteristics. Key clinical features were a progressive dementia, ataxia, or other cerebellar disturbances, and abnormal behaviour, mood disorders, or other psychiatric symptoms. Table 1 shows the age at onset, disease duration and PRNP codon 129 genotype as described in the papers. Fig 1 gives the findings of the meta-analysis on the association between the number of inserted octapeptide repeats and the age at disease onset. In this analysis, 38 patients from 17 families with information on the PRNP codon 129 genotype were included. We found a significant inverse relation between the number of inserted octapeptide repeats in PRNP and age at disease onset (p=0.0002 in the model unadjusted for PRNP codon 129 genotype and p<0.0001 after adjusting for this genotype). Overall, the codon 129 genotype contributed significantly to the model (p=0.0003). Compared with carriers of the PRNP codon 129 genotype MV, patients with genotype MM tended to have an earlier onset and carriers of genotype VV a later onset. The PRNP codon 129 genotype and the number of inserted octapeptide repeats in PRNP explain together 93% of the between family variance in age at onset, leaving only 7% unexplained.

The relation between number of repeats and survival after disease onset was assessed in all 55 patients. Adjusted for age at onset, survival was significantly prolonged in carriers of the shorter repeats (p<0.001).

Finally, we extended the analyses to affected family members (n=40) of confirmed cases, who were suspected to carry a PRNP insert mutation on clinical grounds, but for whom genetic analysis or a neuropathological confirmation of CJD or GSS were not available. In this analysis, the relation between number of PRNP repeats and age at onset remained statistically significant (p<0.001). However, the relation between number of PRNP repeats and survival was no longer significant.

DISCUSSION
In this study of 17 patients with early onset dementia we identified two carriers of mutations in genes known to be involved in presenile dementia. One of the mutations, the PSEN2 T301M mutation, is unlikely to be a causal mutation given the absence of an effect on the β amyloid 42/40 levels in cells transfected with the mutation. Further, we detected a mutation in PRNP in one of 17 patients with early onset dementia. The PRNP two-octapeptide repeat insertion we observed in this patient led to an atypical form of dementia with disease duration of more than 10 years. Repeat insertions of two extra octapeptide repeats have been described previously. Van Harten et al also described a patient with a two-octapeptide repeat insertion with atypical dementia in The Netherlands. We could not find a link between the two Dutch patients within six generations.
However, the rarity of the PRNP insertion mutations, the presence of the V allele on the PRNP insertion haplotype, and particularly the identical nucleotide sequence of the inserted octapeptide repeats in both patients, suggest that the patients are related and developed the disease as a result of the same mutation.

Goldfarb et al described the second patient. This young patient (58 years) showed a severe neurological deterioration leading to mutism and comatose state within weeks. However, the mother of the patient, carrying the same mutation.

<table>
<thead>
<tr>
<th>Article</th>
<th>n Repeats</th>
<th>n Patients*</th>
<th>PRNP codon 129†</th>
<th>Age at onset (years)</th>
<th>Disease duration (months)‡</th>
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<tr>
<td>Laplanche⁸⁹</td>
<td>1</td>
<td>1</td>
<td>MM</td>
<td>73</td>
<td>4</td>
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<tr>
<td>Goldfarb⁹⁰</td>
<td>2</td>
<td>2</td>
<td>MM</td>
<td>58/75</td>
<td>3/156+</td>
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<tr>
<td>Van Harten⁹¹</td>
<td>2</td>
<td>1</td>
<td>VV</td>
<td>61</td>
<td>84</td>
</tr>
<tr>
<td>This article</td>
<td>2</td>
<td>1</td>
<td>MV</td>
<td>59</td>
<td>120+</td>
</tr>
<tr>
<td>Laplanche⁹²</td>
<td>4</td>
<td>1</td>
<td>MM</td>
<td>82</td>
<td>4</td>
</tr>
<tr>
<td>Rossi⁹³</td>
<td>4</td>
<td>1</td>
<td>MM</td>
<td>65</td>
<td>6</td>
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<td>1</td>
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<td>MM</td>
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<td>5</td>
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<td>5</td>
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<td>n.a.</td>
<td>31/45</td>
<td>60/180</td>
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<td>3</td>
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<td>26 to 44</td>
<td>84 to 168+</td>
</tr>
<tr>
<td>Shwarz³⁸</td>
<td>5</td>
<td>3</td>
<td>MV</td>
<td>35 to 54</td>
<td>3 to 24</td>
</tr>
<tr>
<td>Obi³⁹</td>
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<td>3</td>
<td>MM</td>
<td>33 to 34</td>
<td>60 to 84+</td>
</tr>
<tr>
<td>Capellani³⁴</td>
<td>6</td>
<td>2</td>
<td>MV</td>
<td>38</td>
<td>48/120</td>
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<tr>
<td>Nicholl³⁶</td>
<td>6</td>
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<td>n.a.</td>
<td>34/46</td>
<td>3/60</td>
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<tr>
<td>Collinge, Poulter³³</td>
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<td>13</td>
<td>MM³/MM³/MV³⁵</td>
<td>22 to 47</td>
<td>24 to 156</td>
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<td>7</td>
<td>3</td>
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<td>23 to 35</td>
<td>120 to 156</td>
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<tr>
<td>Dermaut⁴⁰</td>
<td>7</td>
<td>2</td>
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<td>24/31</td>
<td>132</td>
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<tr>
<td>Goldfarb³⁶</td>
<td>8</td>
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<td>35 to 54</td>
<td>3 to 24</td>
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<tr>
<td>Laplanche³⁶</td>
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<td>3</td>
<td>MV³/MM³</td>
<td>36 to 144+</td>
<td>12± to 72</td>
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<tr>
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<td>3</td>
<td>MV³/MM³</td>
<td>21 to 32</td>
<td>36 to 144+</td>
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<td>Moore, Xiang⁴²</td>
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<td>n.a.</td>
<td>23 to 41</td>
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<td>Owen⁴³</td>
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<td>53</td>
<td>30</td>
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<tr>
<td>Krasemann⁴⁰</td>
<td>9</td>
<td>1</td>
<td>MM</td>
<td>32</td>
<td>72+</td>
</tr>
</tbody>
</table>

*Patients with mutation analysis, or patients with autopsy and a blood relative with mutation analysis, or patients with mutation analysis as well as autopsy, are reported. †M, methionine; V, valine; the numbers in superscript denote the number of carriers. ‡For patients alive at time of publication, the disease duration is indicated with a †.§Not included in the analyses because of missing age at onset or duration for each patient. n.a., not available.

![Figure 1](http://jnnp.bmj.com/content/10.1136/jnnp.2003.020198)

Figure 1 Relation between the number of insertions in the PRNP octapeptide region and age at onset (on a log scale), adjusted for PRNP codon 129 genotype. The best fit line is shown.
Animal studies have shown that prion protein molecules containing a larger octapeptide repeat region aggregate more readily and have a higher protease resistance. In line with these findings, our study suggests that longer insertions in the octapeptide repeat region in PRNP may lead to the inhibition of turnover resulting in a faster accumulation of the protein, earlier onset of disease, and reduced survival.

In conclusion, in a Dutch population-based sample of 17 patients with early onset dementia, we identified one patient in whom the disease was most likely explained by a two-octapeptide repeat insertion in PRNP. We further showed in a meta-analysis that the number of inserted octapeptide repeats in PRNP is inversely related to age at onset and duration of disease. Our meta-analysis shows that the length of the octapeptide repeat insertion in combination with the PRNP codon 129 genotype explains a large part of the variation in age at onset observed. The challenge in the future will be to determine the biological mechanism underlying this process.

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Competing interests: none declared

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REFERENCES


6 Palmer MS, Collinge J. Mutations and polymorphisms in the prion protein gene. Hum Mutat 1993;2:168–73


13 Risodaa SS. Nuclear inclusions in glutamine repeat disorders: are they perinuclear, coincidental, or beneficial? Cell 1998;95:1–4


Unusual differential diagnosis of leptomeningeal enhancement: moyamoya disease

We report on a 13 year old boy who presented with a focal left sided motoric seizure and a history of progressive headaches since 6 years of age. On neurological examination, motor function of the limbs, deep tendon reflexes, and sensory examination were normal, but there was slight left sided bradydysdiadochokinesis. Routine blood tests and anti-immune antibody screening as well as cerebrospinal fluid (CSF) analysis were normal.

Magnetic resonance imaging (MRI) showed multiple curvilinear areas of high signal intensity in leptomeninges (fig IA) and diffuse enhancement along leptomeningeal surfaces (fig IB). Digital subtraction angiography demonstrated severe stenoses of both distal internal carotid artery (ICA) segments (fig IC) as well as extensive leptomeningeal collaterals (fig ID).

Leptomeningeal enhancement on post-contrast T1 weighted MR images has been reported in association with meningitis, meningeal carcinomatosis, and angiitis. Common among these diseases is pathologic inflammation or carcinomatous infiltration of the leptomeninges, with or without involvement of the small leptomeningeal vessels.

In 1995 Ohta et al reported diffuse leptomeningeal enhancement on post-contrast T1 weighted MRI in children with moyamoya disease and named these finding the “ivy-sign”, because it resembles ivy creeping on stones. The ivy-sign can be found on fluid attenuation inversion recovery (FLAIR) images, as well as on contrast enhanced MR images.

The characteristic enhancement is caused by an engorged pial network via leptomeningeal anastomosis. The source of leptomeningeal high signal intensity on FLAIR images is not clear but it seems largely due to slow flowing vessels of the engorged pial network.

Thus, the mechanism of the leptomeningeal ivy-sign can be explained by an engorged pial network and possibly the congested meninges.

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References

Figure 1 (A) Axial FLAIR MRI reveals multiple areas of high signal intensity (arrows) in leptomeninges. (B) Axial contrast enhanced T1 weighted image shows diffuse enhancement along leptomeningeal surfaces (arrows). (C) Lateral view of right carotid angiogram and (D) right vertebral angiogram demonstrate severe stenoses at the distal ICA (arrow) and wide terminal segments of the posterior and middle cerebral arteries due to extensive leptomeningeal collateral flow (arrowheads).

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