Biological activity of interferon betas in patients with multiple sclerosis is affected by treatment regimen and neutralising antibodies

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The efficacy of interferon (IFN) beta in the treatment of multiple sclerosis (MS) has been demonstrated in several trials. However, in some patients with MS this therapy is less effective, and they are considered non-responders. In some of these patients the development of anti-IFN beta neutralising antibodies (NAbs) has been observed. NAbs can interfere with the receptor binding sequence of IFN beta, inhibiting biological activity and therapeutic action.

Biological activity can be estimated by measuring markers of IFN beta activity, which are specific IFN induced proteins that have been demonstrated to peak after IFN beta injections. Of the usual biological markers of IFN beta activity, which are specific IFN induced proteins, MxA has a dose dependent specificity for type I IFNs, but not for IFN gamma. MxA has a shorter half-life than the protein, the measurement of a specific transcript offers a better measure of the biological activity of IFN beta. We measured MxA mRNA using a quantitative-competitive polymerase chain reaction (qPCR) technique shown to be precise, accurate, and sensitive to a level of 1 fgMxA/pgGAPDH. This method of quantification allowed us to determine and analyse fluctuations of MxA expression during five days of treatment, to evaluate the biological activity of IFN beta in parallel with the development of NAbs, and to compare the biological response to IFN beta in patients with MS treated once a week with Avonex or three times a week with either Betainterferon or Rebif.

PATIENTS AND METHODS

Patients

Blood samples were obtained from a total of 62 patients with MS (22 men and 40 women) who received treatment with recombinant IFN beta. Of these, 16 received IFN beta-1a (Avonex; Biogen, Cambridge, USA) 30 micrograms intramuscularly (IM) once a week, 36 received IFN beta-1a (Rebif; Serono, Basel, Switzerland), either 22 micrograms (n = 24) or 44 micrograms (n = 12) subcutaneously (SC) three times a week, and 10 received IFN beta-1b (Betaferon; Schering, Berlin, Germany) 250 micrograms SC three times a week. The mean duration of therapy was 20 (SD 17) months (range 3–60 months) (table 1).

Patients were not randomised and enrolled retrospectively. Eligibility criteria included a diagnosis of MS according to the McDonald criteria, relapsing–remitting (RR) clinical course, Expanded Disability Status Scale (EDSS) score 0–6.5, and informed consent. All patients included were clinically inactive and steroid free in the three months preceding the enrolment. Exclusion criteria included significant other medical illnesses, previous switch in type of IFN beta treatment, and prior immunosuppressive therapy with cytotoxic activity.

Study design

Eligible patients had been under treatment for at least three months prior to the study and had been screened for the

Abbreviations: AUC, area under the concentration time curve; EDSS, Expanded Disability Status Scale; IFN, interferon; MS, multiple sclerosis; MxA, myxovirus resistance protein A; NAbs, neutralising antibodies; OAS, oligoadenylate synthetase; PBMC, peripheral blood mononuclear cell; TRU, tenfold reduction unit
presence of IFN beta induced NAbs before and every three months, as previously described.\textsuperscript{23,24}

MxA mRNA was assessed every morning from day 1 to day 5. All patients had their injections synchronised for the study; as a consequence Avonex was always administered once a week between 8 pm and 10 pm of day 1, whereas Betaferon, Rebif 22, and Rebif 44 were injected between 8 pm and 10 pm of day 1 and day 3. Blood samples were taken every morning between 8 am and 10 am. On day 1 NAbs were also measured.

Quantification of MxA mRNA
MxA mRNA was quantified according to our previously published protocol.\textsuperscript{19,21} Briefly, peripheral blood mononuclear cells (PBMCs) were separated on a Ficoll-Hypaque (Pharmacia, Uppsala, Sweden) density gradient and total RNA was extracted using RNAzol reagent, following the manufacturer’s instructions (Ambion, Austin, TX). Complementary DNA (cDNA) was then prepared, using 10 mM of random hexamer primers (Perkin Elmer, Norwalk, CT) and 100 U of Moloney murine leukaemia virus reverse transcriptase (Gibco Laboratories, Grand Island, NY). For the qc-PCR reaction two competitor cDNA fragments (co-MxA and co-glyceraldehyde phosphate dehydrogenase (GAPDH)) were generated and co-amplified with target cDNA.\textsuperscript{20} PCR amplification products were then resolved following separation by 2% agarose gel electrophoresis. Bands were visualised by EtBr staining and quantified by densitometric scanning of the gel using a GelDoc 1000 UV fluorescent system (Bio-Rad, Richmond, CA).

The ratios between competitors and target cDNA were evaluated as ratios between bands values, taking as ratio 1 an amount of starting targets (MxA or GAPDH) equal to the amount of each competitor.\textsuperscript{19} The MxA mRNA levels, expressed as fgMxA/pgGAPDH, were normalised using GAPDH as housekeeping gene, to avoid differences due to possible RNA degradation/contamination or different reverse transcription efficiency.

Evaluation of neutralising antibodies
NAbs were measured with a bioassay based on the cytopathic effect (CPE) of encephalomyocarditis virus (EMC) (CPE assay) on human lung carcinoma cells (A549).\textsuperscript{23,24} Serum samples were diluted, mixed with one of the three IFN beta preparations at a final concentration of 10 IU/ml, and added overnight to monolayers of A549 cells in 96 well plates. Cells were then infected with EMC murine virus and viable cells were quantified 24 hours later by staining with crystal violet in 20% ethanol. The dye taken up by the cells was eluted with 33% acetic acid and its absorbance was measured in a densitometer at 620 nm. The neutralisation titre of a serum sample was calculated according to Kawade’s formula\textsuperscript{25} and expressed in tenfold reduction unit (TRU).\textsuperscript{26} A level of \textsuperscript{26}TRU was considered as the threshold for positivity.

Three categories of patients were identified based on NAbs: NAB negative (NAB−) patients had no positive samples during follow up; persistent NAB positive (NAB+) patients had at least two consecutive samples positive for NAbs; and isolated NAB+ patients had a single NAB+ sample or had sporadic positivity during follow up.

Statistical analysis
The data was statistically analysed using GraphPad Prism program version 4.0 (GraphPad Software Inc., San Diego, CA). The area under the concentration time curve (AUC) was calculated using the trapezoidal summation after adjusting for baseline values. Non-parametric statistical tests were used according to the parameters analysed (Mann–Whitney test and Wilcoxon’s test). Differences were considered significant when \( p < 0.05 \).

RESULTS

Patients
Table 1 shows the baseline demographic and clinical characteristics of the study patients. There were no significant differences between the groups with regard to demographic or clinical characteristics, except for the duration of treatment with Rebif 44, which was shorter compared with the other three treatment regimens (Rebif 22 vs Rebif 44, \( v = 1.724 (0.17) \), 1.94 vs 1.95, 1.50 vs 1.0, 1.26 vs 1.26, 1.67 vs 1.18, 8.0 vs 5.2). There were no significant differences in disease duration or sex distribution between the groups. The patients were not randomised, but retrospectively included, since the present study was not conducted to determine the incidence and prevalence of NAbs in serum samples from patients with MS who were treated with Betaferon, Avonex, Rebif 22, or Rebif 44. Hence a direct comparison of the percentage of NABs and Nabs- patients reported in this study is not possible.
IFN beta biological responders and non-responders
In a previous report, we examined the MxA gene expression in PBMCs from 99 untreated patients with MS and we calculated an upper threshold of normal as mean baseline expression + 3 SD = 132 fgMxA/pgGAPDH.11,12 Two categories of patients treated with IFN beta were identified based on MxA mRNA levels after IFN beta administration: IFN beta biological responders had at least one MxA mRNA value higher than the established threshold (>132 fgMxA/pgGAPDH), and IFN beta biological non-responders had MxA mRNA values lower than 132 fgMxA/pgGAPDH during the whole study.

Based on the above threshold, 49/62 (79%) patients were IFN beta biological responders, whereas the remaining 13/62 (21%) patients were IFN beta biological non-responders as MxA expression was unaffected by IFN beta administration. Among the biological responders, 13 were treated with Avonex (81% of the patients treated with Avonex), 7 were treated with Betaferon (70%), 19 were treated with Rebif 22 (79%), and 10 were treated with Rebif 44 (83%). Moreover, of the 49 IFN beta biological responders, 45 (92%) patients were NAb–, three patients were isolated NAb+, and one patient was persistent NAb+. However, the three isolated NAb+ and the single persistent NAb+ patient presented NAb titres <45 TRU during their follow up and were negative during the study.

Among the 13 IFN beta biological non-responders, 10 were found to be persistent NAb+ and positive during the study, one was persistent NAb+ and negative during the study, and two patients were NAb–.

MxA expression and neutralising antibody status
A comparison of MxA expression and NAb status showed that changes in MxA mRNA levels were greater in NAb– than in persistent NAb+ patients (fig 1). Such analysis was not possible for isolated NAb+, because they were too few in number.

Not unexpectedly, abolished MxA gene expression was more commonly found in patients with high NAb titres (>45 TRU) than in patients with low (<45 TRU) NAb titres: all patients (100%) with NAb titres >45 TRU showed no biological activity, as indicated by MxA mRNA levels <132 fgMxA/pgGAPDH during the whole study (fig 1). Of the three remaining persistent NAb+ patients with NAb titres >45 TRU, two subjects showed no biological activity, but one patient showed significant increases in MxA mRNA.

When the AUCs were compared, the total augmentation of MxA mRNA was fourfold greater in the NAb– group than in persistent NAb+ (p<0.0001).

Comparison of the biological activity of the four interferon beta preparations
Following the administration of IFN beta in biological responders, MxA mRNA concentrations peaked at 12 hours and then declined to baseline levels.11,12 As expected, MxA mRNA levels in patients given Avonex were lowest on day 1, peaked on day 2 (+12 hours after injection) and then decreased in the following days (fig 2A). No such decrease was seen with Betaferon or Rebif (Rebif 22 and Rebif 44) because, for both preparations, a booster was given three times a week rather than once a week. In particular, the average profiles of MxA expression, in patients treated with both Betaferon and Rebif, showed a second peak of expression on day 4, +12 hours after the second IFN beta injection (fig 2B–D).

MxA expression for both Betaferon and Rebif groups was statistically greater than that for the Avonex group on day 1 (all p < 0.038), 4 (all p < 0.045), and 5 (all p < 0.044) (table 2). On the other hand, differences of MxA mRNA levels among the four preparations of IFN beta were not significant on day 2 and 3 (+12 and +36 hours after the first IFN beta injection) (all p > 0.077) (table 2). Interestingly, no differences in the MxA expression were found between Rebif 22 and Rebif 44 during the five days of treatment (all p > 0.24) (table 2).

There were no statistical differences among the MxA mRNA levels induced at each time point in persistent NAb+ patients treated with Avonex versus Betaferon, Rebif 22, and Rebif 44 (all p > 0.12) (fig 2).

Although this study analysed only five treatment days and did not consider the effect of the third weekly injection of Betaferon and both Rebif preparations, the MxA mRNA AUC values approached statistical significance for both Rebif preparations and Betaferon versus Avonex (0.058 > p > 0.071).

Biologically effective injections
The profiles of the MxA concentration time curve were similar in all biological responders under treatment with Avonex. On the other hand, some subjects treated three times a week with either Betaferon or Rebif presented an unexpected profile, as MxA expression did not increase after one of the two injections considered. To examine this phenomenon, we evaluated the biological efficacy of every injection separately: injections were considered “biologically effective” when they induced MxA mRNA levels higher than the established threshold (132 fgMxA/pgGAPDH).

Among the 49 biological responders, 13 were treated once a week, and 36 were treated three times a week (7, Betaferon; 19, Rebif 22; 10, Rebif 44). Of these 36 patients, 13 (36%) presented a single “biologically effective” injection, instead of two: 2/7 (29%) were treated with Betaferon, 7/19 (37%) were treated with Rebif 22, and 4/10 (40%) were treated with Rebif 44. Effective injections were detected indiscriminately after both the first and second injection. As a whole, in biological responders treated three times a week, only 82% injections (59/72) were “biologically effective”, compared with 100% (13/13) of Avonex injections.

DISCUSSION

Four IFN beta preparations are presently used in the treatment of MS: Avonex, Betaferon, Rebif 22, and Rebif 44. The differences among the four preparations are in their biochemical structure, dose, dosing frequency, route of administration, and vehicle. Despite these differences,
significant therapeutic effects were observed with each preparation. However, the optimal dosing regimen for IFN beta therapy in this indication is still under debate. Moreover, several studies have demonstrated different degrees of immunogenicity, probably due to one or more of the above-mentioned differences.

To compare the degree and duration of modulation of the biological response induced by the four IFN beta preparations, in the present study we evaluated in vivo changes in MxA gene expression in patients with MS during five days of treatment. As far as we know, such temporal characterisation of biological activity has not been described previously. In the other investigations in which IFN beta biological activity was studied for several consecutive days, the following topics were analysed: (a) healthy volunteers treated with IFN beta rather than treated patients with MS, (b) study of only one or two preparations instead of four, and (c) data obtained ex vivo instead of in vivo. Moreover, the authors measured protein markers such as β2-microglobulin, OAS, and MxA. Although these proteins are considered as classic markers of the biological activity of IFN beta, they are characterised by slow decay. Therefore, although the protein quantification using methods such as enzyme-linked immunosorbent assay (ELISA) is simpler than the measurement of the specific transcript, we preferred mRNA quantification because mRNA has a shorter half-life than the protein, and its level reflects biological activity of every single injection allowing the detection of small fluctuations in expression.

In the present study, two categories of patients were identified based on MxA mRNA induction: IFN beta biological non-responders and IFN beta biological responders. As previously demonstrated, the lack of biological activity correlated with both higher NAb titres and persistent presence of NAb. Under these conditions, IFN beta injections always failed to increase MxA mRNA levels independently of dose or dosing frequency of the treatment. On the other hand, the profiles of MxA AUCs in patients presenting persistently low NAb titres (<45 TRU) show a MxA induction, although at significantly lower levels (fig 1). This could imply that low NAb titres can be overcome by increasing the dose of IFN beta, whereas in the presence of higher NAb titres, the biological response to IFN beta is always abolished, even by higher doses and dosing frequency of the treatment.

Analysis of MxA mRNA levels in IFN beta biological responders clearly demonstrated higher biological responses in patients treated three times a week instead of once a week. The cumulative biological activities, as measured by AUC, approached statistical significance, although only two out of the three weekly injections were considered in this study. The third weekly injection influenced the level of MxA mRNA on day 1 (+156 hours for patients treated with Avonex and +60 hours for patients treated with Betaseron and the two Rebifs) as it was significantly higher in patients treated with Betaseron and the Rebifs compared with those treated with Avonex.

Table 2 MxA mRNA expression in patients negative for neutralising antibodies during five days of treatment

<table>
<thead>
<tr>
<th></th>
<th>Avonex (n = 14)</th>
<th>Betaseron (n = 6)</th>
<th>Rebif 22 (n = 17)</th>
<th>Rebif 44 (n = 10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 1</td>
<td>69 (43)</td>
<td>284 (243)</td>
<td>142 (88)</td>
<td>109 (57)</td>
</tr>
<tr>
<td>Day 2</td>
<td>351 (193)</td>
<td>429 (352)</td>
<td>397 (331)</td>
<td>258 (112)</td>
</tr>
<tr>
<td>Day 3</td>
<td>20 (102)</td>
<td>189 (76)</td>
<td>235 (213)</td>
<td>147 (74)</td>
</tr>
<tr>
<td>Day 4</td>
<td>159 (122)</td>
<td>498 (242)</td>
<td>321 (264)</td>
<td>360 (431)</td>
</tr>
<tr>
<td>Day 5</td>
<td>95 (83)</td>
<td>261 (272)</td>
<td>191 (160)</td>
<td>201 (110)</td>
</tr>
</tbody>
</table>

Values are mean (SD) fgMxA/pgGAPDH.

*Statistical difference was found between the Mxa mRNA levels induced by Avonex versus Betaseron, Rebif 22, and Rebif 44 on day 1, 4, and 5. No difference was found on day 2 (+12 hours after the first injection) and day 3 (+36 hours).
showed increase of MxA mRNA level after their single injection. On the other hand, the absence, or greatly reduced, MxA induction could have a biological basis. Therefore, it seems to be more likely that PBMCs of patients treated with IFN beta, undergo a process of desensitisation in response to repeated exposure to the cytokine. Accordingly, recently it has been observed that in vitro T cells become desensitised as a result of persistent IFN beta-1a stimulation, regaining full responsiveness to treatment by 168 hours.29

Despite clear evidence of higher biological response in patients treated three times a week, it is unclear whether this difference is clinically relevant, as the lower biological activity of Avonex may be counterbalanced by its lower incidence of NAB induction observed in all therapeutic trials27 30 and in longitudinal comparisons between the different types of IFN beta.27 30 Moreover, it should be taken into consideration that a higher cumulative biological activity does not necessarily induce greater clinical efficacy. Indeed, clinical efficacy relates to half-life of induced molecules and/or modifications of cell populations with therapeutic action. Several lines of evidence point to the important role of the up/down-regulation of cytokines and chemokines,28 31–33 with fast induction and short half-life.29 Conversely, IFN beta also induces longlasting effects such as up/downregulation of other cytokines,28 increases in NK cells functional activity,27 34 development of different cell subsets,35 or downregulation of matrix metalloproteinases.36 Consequently, these effects, which are maintained for a week or more, require less frequent administrations of the drug. Anyway, therapeutic differences among the four IFN beta regimens can only be demonstrated by head to head clinical trials that have to last some years to take into consideration the delayed negative effects of NABs.13

In conclusion the results of the present study demonstrate that an optimal IFN beta regimen is not yet available: Avonex, given once a week, shows significantly lower cumulative biological activity, but significantly lower incidence of NABs, compared with both Betaferon and Rebif.27 29 30 On the other hand both Betaferon and Rebif (Rebif 22 and 44) given three times a week, show greater cumulative biological activity but higher risk of development of NABs,13 29 30 which abrogate IFN beta therapeutic action.12 29 30 Hence, to tailor the best treatment in both patients with newly diagnosed MS and those already receiving treatment, the neurologist must carefully consider the results of clinical trials, the pharmacokinetic data, the risk of loss of therapeutic efficacy due to the development of NABs, and the peculiar clinical and prognostic characteristics of each patient.

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References

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HISTORICAL NOTE

Cotugno and cerebrospinal fluid

In 1761 Cotugno gave the first reliable account of ventricular and subarachnoid fluid. Until Cotugno, anatomists had found empty spaces around the brain and cord and thought that in life they were filled by vapour. Willis said the ventricles were empty spaces, or served the "vile duty of a sewer. In the dead they may be filled with water...if the serous fluid in the blood is too abundant." Albrecht von Haller's famous textbook (at the same time as Cotugno's studies) describes:

"As in the pericardium...a thin humour constantly exhales from the arteries into the ventricles of the brain and is constantly drawn back through the veins...so often the collected moisture turns into water and even distends the ventricles...A great abundance of water has been found in the ventricles of apoplexies, the sorporose, convulsive, paralytics, and victims of epidemic fevers; hydrocephalus even more."

Cotugno studied 20 adult male bodies. He established the free circulation between the cranial and spinal dura of cerebrospinal fluid (sometimes referred to as liquor Cotunnii). His lucid description indicating its formation and absorption from blood vessels is contained in his work on sciatica.

"Not only does this water contained in the tube of dura mater ensheathing the spinal marrow [cord] from the occiput to the os sacrum, surround the marrow constantly, but it also abounds in the hollow of the skull and fills all the spaces found between the brain and the encompassing dura mater...It seems to be a human law that the space around the spinal marrow that is filled with water increases with man's age...Hitherto anatomists have not observed this large collection of water in the spine and around the brain because of the ridiculous method usually employed for the dissection of bodies...they cut off the head with the neck...all the fluid collected around the brain and spinal marrow is at once lost...and the anatomist is misled by the appearance of empty spaces...It seems beyond all doubt that the spinal fluid, as well as that which lubricates all other cavities of the body, constantly oozes from the extremities of the smallest arteries and, finally is absorbed through very small inhaling veins, so that there is a continual state of renovation."

Further, he noted the incoagulability of CSF in health, but like urine in nephritis, which he observed some 50 years before Bright, it clouded on boiling, only in disease. This work was overlooked until Magendie reprinted it in 1827.

In this crucial work, Cotugno, an astute observer and clinician, differentiated sciatic nerve pain from arthritis of the hip, probably for the first time. The eponym Cotugno's syndrome was subsequently applied to unilateral sciatic neuralgia. He also wrote about typhus and gave a fine description of the pathology of smallpox pustules.

Domenico Felice Antonio Cotugno (1736–1822)

Near the heel of Italy lies the town of Ruvo Pugliese, the birthplace (29 Jan 1736) of Cotugno. Most of his life he spent in Naples. His family were poor and hardship was his constant companion in his formative years. After medical training in Salerno, he worked in the University of Naples and the Ospedale degli Incurabili. Cotugno surmounted serious illness while resident at the hospital. He became an assistant at the Ospedale degli Incurabili. In 1766 he became professor of anatomy, the leading physician in Naples, and director of the Ospedale. By the age of 31 he was widely acclaimed for his excellent publications, including two books.

When he was only 25, in 1761, his dissertation, Aquaeductus auris humane internae, predated the work of Hermann von Helmholtz. In it he described the vestibule, semicircular canals, and cochlea. He demonstrated the labyrinthine fluid, and considered mechanisms of resonance, sound transmission, and hearing. He depicted the columns in the bony spiral lamina of the cochlea known as Cotunnius' columns. His description of the nasopalatine nerve, and its role in sneezing anticipated Antonio Scarpa's work. In 1765 he visited Rome and northern Italy, and was befriended by Morgagni. Notable success in practice led to appointment as physician to Ferdinand IV, King of Naples, accompanying his travels to Austria and Germany.

He was a dedicated doctor, but was also a student of art, architecture, Latin, and antiquities. A greatly esteemed physician, local lore was that nobody in Naples could die without a passport from him. He was renowned for his devotion to medicine and scientific investigation. He stopped teaching in 1814 but continued to attend his hospital daily. In 1818 he had a cerebral embolism that eventually caused his death on 6 October 1822.

The generous son of a poor father, he left 100 000 ducats to the Ospedale degli Incurabili.