SHORT REPORT

Profilin1 E117G is a moderate risk factor for amyotrophic lateral sclerosis

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ABSTRACT

Background Amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) are progressive neurodegenerative disorders that share significant clinical, pathological and genetic overlap and are considered to represent different ends of a common disease spectrum. Mutations in Profilin1 have recently been described as a rare cause of familial ALS. The PFN1 E117G missense variant has been described in familial and sporadic cases, and also found in controls, casting doubt on its pathogenicity. Interpretation of such variants represents a significant clinical-genetics challenge.

Objective and results Here, we combine a screen of a new cohort of 383 ALS patients with multiple-sequence datasets to refine estimates of the ALS and FTD risk associated with PFN1 E117G. Together, our cohorts add up to 5118 ALS and FTD cases and 13 089 controls. We estimate a frequency of E117G of 0.11% in controls and 0.25% in cases. Estimated odds after population stratification is 2.44 (95% CI 1.048 to 5.78), Mantel-Haenszel test p=0.036.

Conclusions Our results show an association between E117G and ALS, with a moderate effect size.

INTRODUCTION

Amyotrophic lateral sclerosis (ALS) is a devastating neurodegenerative disorder in which the progressive loss of motor neurons causes weakness and paralysis leading to death typically 3–5 years after onset.1 Around 5–10% of ALS cases are familial, which has allowed the identification of a handful of mutant genes that cause classical dominant adult-onset ALS, and are occasionally found in sporadic patients with ALS, who report no family history for the disease.2 Frontotemporal dementia (FTD) co-occurs in 5–10% of patients with ALS and the finding that a number of mutations can cause either disorder, and that ALS and FTD post-mortem pathology observations share numerous similarities, has led to these disorders being considered part of the same disease spectrum.3

Recently, Wu et al.4 have identified missense mutations in Profilin 1 (PFN1), a three-exon gene encoding a protein involved in actin filament formation, as causative of ALS. With the exception of the E117G mutation, all mutant PFN1 alleles were found in familial ALS cases. One successive study identified a novel missense mutation, while a number of screening studies have failed to identify mutations, suggesting these are a rare cause for ALS. By contrast, the E117G mutation was identified in control and familial cases, and was also found at a lower frequency in controls in other populations, thus casting a doubt on the significance of this variant.5

Here we report the screening of a UK population for PFN1 mutations. We identify cases carrying the E117G variant and analyse the frequency of this variant in 4700 UK controls. We then meta-analyse all the published and publicly available data for this variant, to conclude this may represent a rare risk factor for ALS.

METHODS

Study population

Three hundred and eighty-three sporadic and familial ALS cases, from specialist motor neuron disease clinics in London, UK, (UCL Partners motor neuron disease clinics), provided written informed consent. The familial cases were 23 and represent 6% of the cohort. DNA was extracted from whole blood.

Sequencing analysis

PFN1 exons 1, 2 and 3 were subjected to PCR amplification by REDtaq Polymerase (Sigma), using the primers and PCR conditions as described in Wu et al.4 Reaction mixtures were made up in accordance with the manufacturer’s instructions. The PCR product was purified from the amplification mixture using microCLEAN PCR clean-up kit (Microzone), and the repeat region of interest was sequenced Ready Reaction kit V1.1. 15 μL sequencing reactions comprised of 5–20 ng of DNA template, 1 μL BigDye mix, 5 μL Better Buffer (Microzone), 5 pmol of the forward and reverse sequencing primer (same as PCR primers), 1 μL betaine and were made up with an appropriate volume of nuclease-free water. Cycling parameters for sequencing were set to 300 s at 95°C, 30 s denaturing at 96°C, 15 s annealing at 55°C, 240 s extension at 60°C and repeated for 35 cycles. Sequences were precipitated in ethanol, EDTA and sodium acetate solution and further washed in 70% ethanol. Sequences were determined on a 3730xl DNA Analyser capillary sequencer (Applied Biosciences) and analysed with Geneious V6.1 sequencing software (Biomatters).
Search of public databases and published data
We searched for published sequencing data on PFN1 in ALS, FTD and control samples from prevalently Caucasian cohorts, updated to 10 May 2013. In particular, we received from the UK10K, (http://www.uk10k.org, a large sequencing study of UK individuals with conditions not related to ALS/FTD) the allele frequency for E117G in 4732 exome sequences.

Statistical methods
Association p values were based on Fisher exact tests using the allele count data. Stratified population analysis was performed using the stratified Mantel-Haenszel test. All tests were performed using the R statistical software.

RESULTS
Sequence screening of PFN1 in a UK ALS cohort
Screening of the three PFN1 exons in our samples revealed no novel variant. We identified two ALS samples carrying the dinucleotide substitution leading to the previously described E117G coding change. Both patients had no ALS family history. The first patient was a male with bulbar onset ALS, age was 65 years at onset, and a very slowly progressive form of disease. The second patient was a male with limb onset ALS, age was 54 years at onset, and a disease duration of 3 years. Both patients were negative for screening of C9orf72 expansions and mutations in TARDBP, FUS and VCP.

Meta-analysis of PFN1 E117G variant
To estimate the frequency of the E117G alleles in controls, we combined several publicly available cohorts of exome-sequenced individuals, whose conditions do not overlap with the ALS-FTD spectrum. The largest control cohorts are: NHLBI Exome-sequencing dataset (4300 European–American samples), and the UK10K dataset (4732 UK samples).

We then collected all the published and publicly available data on the E117G PFN1 variant in ALS and FTD. Combining all cohorts, we obtained information on 5118 cases and 13 089 controls of prevalently European descent.

We initially performed our statistical analysis using the original discovery set from Wu et al., with the following modifications to the control group: (A) we excluded the 1K Genome Project data due to low coverage and potential shortfall in detecting rare variants; (B) we excluded the African–American samples from the Exome Variant Server, since the E117G mutation appears to be specific to European-descent populations; (C) we updated the European-descent samples as on June 2013. This analysis shows an OR=3.71 (Fisher exact test p=0.098) (table 1).

We then analysed all available datasets which included cases and controls (total of 4024 cases and 8777 controls) and took into account population stratification by using a Mantel-Haenszel test. This analysis showed a significant difference between groups (p=0.036) and an OR of 2.22 (1.1 to ∞ 95% CI) (table 1).

By combining all available data, also from non-case-control studies, the variant was present in 13 out of 5118 cases and 15 out of 13 089 control samples conferring similar results to the Mantel-Haenszel test, and showing an OR=2.22 (95% CI 1.1 to ∞ Fisher exact test p=0.036) (table 1).

DISCUSSION
PFN1 mutations have been reported to be a rare cause of familial ALS. The PFN1 E117G variant has been also found in sporadic ALS patients, but its presence in numerous control individuals has raised concerns about its role in disease. With the recent advances in DNA sequencing technologies, information on one patient’s whole exome or multiple candidate genes is quickly obtainable—in this context, the interpretation of variants, such as PFN1 E117G, becomes an important clinical-genetics challenge.

In a new screen of 383 UK patients with ALS, we found two E117G carriers. No novel variant was found. In order to gain insight as to how to interpret the role of this variant, we

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<th>E117G</th>
<th>FTD</th>
<th>E117G</th>
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<th>E117G</th>
<th>CONTROL</th>
<th>E117G</th>
<th>p Value</th>
<th>OR (one-tailed 95% CI)</th>
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<td>0.038</td>
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<td>4024</td>
<td>10</td>
<td>8777</td>
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<td>0.038</td>
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*Indicates case-control datasets used for the Mantel–Haenszel test.
ALS, amyotrophic lateral sclerosis; FTD, frontotemporal dementia. 
performed an analysis on all publicly available data for the E117G variant, and we find consistent evidence for the association between E117G and ALS and FTD (p=0.036), with an estimated OR of 2.44 (95% CI 1.048 to 6.0 p=0.038).

Wu et al1 performed functional studies on five PFN1 mutations, including the E117G variant, demonstrating an effect on protein aggregation, actin binding and axon outgrowth. The E117G variant generally showed intermediate results between the wild-type and the other mutations, reaching significant differences from the wild-type in protein aggregate formation and in increased levels in insoluble protein fractions. Axon outgrowth inhibition was observed, but did not reach statistical significance, and actin binding appeared to be normal.4 In summary, functional characterisation of the E117G variant shows lesser changes than clear PFN1 pathogenic mutations, but supports a possible role as risk factor for ALS.

The possibility of multiple mutations contributing to ALS in one individual,1,2 and the role of rare variants in complex diseases such as ALS and FTD, remains to be elucidated and will require very large cohorts in order to obtain sufficiently convincing results. Recently, Geschwind and collaborators have shown that a rare variant in the MAPT gene, where mutations are known to be causative for FTD, can act as a risk factor for FTD and Alzheimer’s disease.3,4 Similarly, we here suggest that a rare variant in PFN1, a gene in which mutations are known to be causative for ALS, can act as a risk factor for disease.

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Contributors PF contributed to the design and conceptualisation of the study, analysis and interpretation of the data and drafted and revised the manuscript. JC, TC and AD contributed to the analysis and interpretation of the data, and revising the manuscript. RH, AM, RO, KS, JC, MS and CL contributed to the analysis of the data and revising the manuscript. JH and EMCF contributed to the analysis and interpretation of the data and revised the manuscript.

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Ethics approval Ethics approval for this project was obtained by the local University College Hospital Ethics committee.

Provenance and peer review Not commissioned; externally peer reviewed.

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REFERENCES