Seroreactivity against lytic, latent and possible cross-reactive EBV antigens appears on average 10 years before MS induced preclinical neuroaxonal damage

Daniel Jons, Viktor Grut, Tomas Bergström, Henrik Zetterberg, Martin Biström, Martin Gunnarsson, Magnus Vrethem, Nicole Brenner, Julia Butt, Kaj Blennow, Staffan Nilsson, Ingrid Kockum, Tomas Olsson, Tim Waterboer, Peter Sundström, Oluf Andersen

ABSTRACT
Background Multiple sclerosis (MS) and presymptomatic axonal injury appear to develop only after an Epstein-Barr virus (EBV) infection. This association remains to be confirmed across a broad preclinical time range, for lytic and latent EBV seroreactivity, and for potential cross-reacting antigens.

Methods We performed a case-control study with 669 individual serum samples obtained before clinical MS onset, identified through cross-linkage with the Swedish MS register. We assayed antibodies against EBV nuclear antigen 1 (EBNA1), viral capsid antigen p18, glycoprotein 350 (gp350), the potential cross-reacting protein anocamtin 2 (ANO2) and the level of sNfL, a marker of axonal injury.

Results EBNA1 (latency) seroreactivity increased in the pre-MS group, at 15–20 years before clinical MS onset, followed by gp350 (lytic) seroreactivity (p=0.001–0.009), ANO2 seropositivity appeared shortly after EBNA1-seropositivity in 16.7% of pre-MS cases and 10.0% of controls (p=0.001). With an average lag of almost a decade after EBV, sNfL gradually increased, mainly in the increasing subgroup of seropositive pre-MS cases (p=8.10^-5 compared with non-MS controls). Seropositive pre-MS cases reached higher sNfL levels than seronegative pre-MS (p=0.038). In the EBNA1-seropositive pre-MS group, ANO2 seropositive cases had 26% higher sNfL level (p=0.0026).

Conclusions Seroreactivity against latent and lytic EBV antigens, and in a subset ANO2, was detectable on average a decade before the appearance of a gradually increasing axonal injury occurring in the last decade before the onset of clinical MS. These findings strengthen the hypothesis of latent EBV involvement in the pathogenesis of MS.

INTRODUCTION
Several pieces of evidence support the involvement of Epstein-Barr virus (EBV) in multiple sclerosis (MS) pathogenesis. Among them is that the risk of MS increases after infectious mononucleosis (IM). In addition, serological studies suggest that a primary EBV infection is a prerequisite for MS, and an increased EBV seroresponse, apparently mainly against EBV nuclear antigen 1 (EBNA1), is detectable several years before clinical onset of MS. Patients with MS may have experienced a severe and possibly atypical EBV primary infection. Thus, serological analyses of archived serum specimens from individuals who later developed MS have revealed an association of increased EBNA1 antibody titres and MS risk from the age of 25 onward. Investigators in Sweden reported that a primary EBV infection in
Children or adolescence was associated with relative protection against MS, whereas a later primary infection was associated with increased MS risk. In a pivotal US study, 766 of 801 (96%) of pre-MS patients were EBV seropositive in a baseline sample obtained at a median 3.8 (range 3–17) years before clinical MS onset. All but one of the 35 remaining EBV-seronegative individuals became EBV seropositive before onset. Moreover, in 30 cases and 30 controls randomly selected from the same cohort, elevation of serum neurofilament light (sNfL) protein, a marker of axonal damage, was observed at a median 6 (range 4–10) years before clinical MS onset. Similarly, in our previous report on sNfL in 30 cases and 30 controls randomly selected from the same cohort, elevation of serum neurofilament light (sNfL) protein, a marker of axonal damage, was observed at a median 6 (range 4–10) years before clinical MS onset. All but one of the 35 remaining EBV-seropositive individuals became EBV seropositive before onset. Moreover, in 30 cases and 30 controls randomly selected from the same cohort, elevation of serum neurofilament light (sNfL) protein, a marker of axonal damage, was observed at a median 6 (range 4–10) years before clinical MS onset. Combining their consecutive EBV and sNfL data the US investigators showed that EBV seroconversion events preceded incipient elevation of sNfL, implicating EBV infection as the leading cause of MS. The age distribution in these studies reflected that of active military personnel, and the crucial observation of sNfL elevation following EBV seroconversion depended on the absence of sNfL elevation in the random sample of the original cohort. The association between EBV conversion and sNfL elevation remains to be confirmed in a broader population with a wider age distribution.

Plasma antibody assays and absorption tests have uncovered an EBNA1 epitope that cross-reacts with the central nervous system protein anoctamin 2 (ANO2) in 14.6% of patients with MS and 7.8% of controls. Reactivity to a similar EBNA1 peptide was reported as part of polyspecific autoimmunity during acute IM. Here, we assessed whether the appearance of EBV seroreactivity in the presymptomatic phase of MS precedes axonal injury detectable with sNfL and whether it is associated with incipient autoimmunity against one reported possible MS autoantigen (ANO2). Leveraging a large repository of material from Swedish university hospital biobanks, we explored seroreactivity against a latency EBV antigen (EBNA1) and two lytic antigens, viral capsid antigen p18 (VCAp18) and glycoprotein 350 (gp350), the latter of which is the major neutralising antibody. We compare the preclinical distribution of sNfL elevation between pre-MS groups with negative and positive EBV and ANO2 seroreactivity.

**MATERIAL AND METHODS**

**Participants**

This nested case–control study used presymptomatically collected blood samples from 669 individuals (pre-MS group) who later received a diagnosis of relapsing-remitting MS and 669 matched controls, as described previously. These individuals were identified from the Swedish MS registry, and a local MS database in Umeå, Sweden. The MS register contained 11 146 patients (www.neuroreg.se), and the Umeå database contained 2887 patients. The register data were crosslinked with serum samples stored in six Swedish biobanks, containing aliquots remaining after clinical microbiological analyses performed at the university hospitals of Skåne, Gothenburg, Örebro, Linköping and Umeå, and at the Public Health Agency of Sweden in Stockholm. Individuals with relapsing-remitting MS whose presymptomatic blood sample was obtained when they were younger than 40 years were included. Controls were matched 1:1 for biobank, sex, date of blood sampling and date of birth, in decreasing priority. Controls were matched with a mean absolute difference of 6 days for date of sampling and 152 days for age at sampling. Because of the scarcity of sera, sNfL analyses were limited to 519 control–control pairs and gp350 analyses to 570 of the 669 case–control pairs. The median sampling age of the study population was 25 years (IQR 21–29 years; range: 2–39 years), and 84% of the participants were female.

<table>
<thead>
<tr>
<th>Pre-MS samples</th>
<th>Time from sampling until MS onset</th>
<th>&lt;5 years</th>
<th>5–10 years</th>
<th>10–15 years</th>
<th>15–33 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>669</td>
<td>222</td>
<td>188</td>
<td>147</td>
<td>112</td>
</tr>
<tr>
<td>Female sex, %</td>
<td>84</td>
<td>87</td>
<td>84</td>
<td>77</td>
<td>86</td>
</tr>
<tr>
<td>Median age (IQR) at sampling, years</td>
<td>25 (21–29)</td>
<td>26 (22–31)</td>
<td>25 (20–29)</td>
<td>25 (21–29)</td>
<td>22 (19–26)</td>
</tr>
<tr>
<td>Median age (IQR) at MS onset, years</td>
<td>33 (28–40)</td>
<td>29 (25–33)</td>
<td>32 (27–37)</td>
<td>37 (32–42)</td>
<td>43 (38–46)</td>
</tr>
</tbody>
</table>

**Samples analysed for sNfL**

<table>
<thead>
<tr>
<th>Time from sampling until MS onset</th>
<th>&lt;5 years</th>
<th>5–10 years</th>
<th>10–15 years</th>
<th>15–32 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>519</td>
<td>112</td>
<td>163</td>
<td>137</td>
</tr>
<tr>
<td>Female sex, %</td>
<td>82</td>
<td>82</td>
<td>85</td>
<td>77</td>
</tr>
<tr>
<td>Median age (IQR) at sampling, years</td>
<td>25 (21–29)</td>
<td>27 (23–32)</td>
<td>25 (20–29)</td>
<td>25 (21–29)</td>
</tr>
<tr>
<td>Median age (IQR) at MS onset, years</td>
<td>35 (29–41)</td>
<td>30 (26–34)</td>
<td>32 (28–37)</td>
<td>37 (32–42)</td>
</tr>
</tbody>
</table>

**Samples analysed for gp350**

<table>
<thead>
<tr>
<th>Time from sampling until MS onset</th>
<th>&lt;5 years</th>
<th>5–10 years</th>
<th>10–15 years</th>
<th>15–33 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>570</td>
<td>189</td>
<td>159</td>
<td>124</td>
</tr>
<tr>
<td>Female sex, %</td>
<td>84</td>
<td>88</td>
<td>84</td>
<td>77</td>
</tr>
<tr>
<td>Median age (IQR) at sampling, years</td>
<td>25 (21–29)</td>
<td>27 (23–31)</td>
<td>25 (20–29)</td>
<td>25 (21–28)</td>
</tr>
<tr>
<td>Median age (IQR) at MS onset, years</td>
<td>34 (28–40)</td>
<td>29 (25–33)</td>
<td>32 (28–37)</td>
<td>37 (33–42)</td>
</tr>
</tbody>
</table>

Controls were matched and had the same characteristics. MS onset means clinical MS onset.

MS, multiple sclerosis; sNfL, serum neurofilament light.
recorded onset of MS, defined as the first symptom suggestive of a demyelinating event, was often ascertained retrospectively. Participants gave informed consent, with an opt-out option, to participate in the study before taking part.

**Laboratory methods**

The German Cancer Research Centre in Heidelberg, Germany, performed assays of antibodies against EBV antigens EBNA1 trunc (aa 325–641), VCAmp18 (aa 1–175)\(^{18,19}\) and ANO2 (aa 79–167), using a bead-based multiplex assay as described previously.\(^{20}\) Samples were analysed in multiple batches, and interbatch controls were used to correct for batch-related variability, as previously described.\(^{21}\) EBV serostatus was determined using published cut-offs of 1800 median fluorescence intensity (MFI) for EBNA1 trunc and 2526 MFI for VCAmp18.\(^{22}\) Samples were classified as EBV seronegative when values for both EBNA1 and VCAmp18 fell below the published cut-offs. EBV seropositivity was defined as either EBNA1 or VCAmp18 above these cut-offs and ANO2 seropositivity as above a cut-off of 420 MFI.\(^{23}\)

The Department of Clinical Microbiology at Sahlgrenska University Hospital, Gothenburg, Sweden, performed the analyses of gp350 antibodies. These antibodies were analysed using ELISA with a DNA construct encoding the full 860-aa extracellular domain of gp350 as the antigen, as previously described.\(^{24}\) Plates were coated with antigen at a concentration of 2 μg/mL, and all samples were diluted 1:200 and analysed in duplicate. Case–control samples were analysed on the same plates.

The Clinical Neurochemistry Laboratory at Sahlgrenska University Hospital, Gothenburg, Sweden, measured sNfL levels using single-molecule array (Simoa) technology and the NF-LIGHT assay on an HD-X Analyzer, according to manufacturer instructions (Quanterix, Billerica, Massachusetts, USA). All samples were measured by board-certified laboratory technicians in a single round of experiments using a single batch of reagents. A quality-control sample with an sNfL concentration of 6.8 pg/mL resulted in 11.2% repeatability and 11.2% intermediate precision. In a quality-control sample with a concentration of 50.3 pg/mL, repeatability was 6.9%, and intermediate precision was 10.6%.

**Statistical methods**

Paired t-tests were used to compare sNfL levels between pre-MS samples and controls for EBNA1 seropositive and EBNA1 seronegative samples. To test the distributional difference of sNfL between those with pre-MS who were EBV seropositive and EBV seronegative, we counted the number X of the 504 EBV-seropositive samples with a higher sNfL level than the highest of the 39 EBV-seronegative sample. Age adjustment was based on the age dependency in the controls. P values were calculated using combinatorics. Assuming equal distributions this gives P(X≥k)=1/(n+m choose k) and the expected number of X is 504.1/(39+1).

Paired t-tests were used to compare seroreactivity and log sNfL between pre-MS cases and their matched controls for the whole sample and for 5-year time groups until clinical MS onset, and between subgroups with and without EBV seroreactivity. Individual delta values (pre-MS minus matched control) were calculated and plotted against time to MS onset, and relationships were estimated with smooth regression analysis using the locally estimated scatterplot smoothing (loess) regression function in R. Effect size (Cohen’s d) was calculated for each 5-year time group. Pearson’s correlations between each of the three antibodies and log sNfL were analysed for pre-MS cases.

**RESULTS**

**EBV serostatus**

Of the 669 pre-MS case–control pairs, 628 samples in the pre-MS group (94%) and 623 control samples (93%) were EBV seropositive. Approaching MS onset, a higher percentage of the pre-MS cases turned EBV seropositive than the matched controls. Of 222 pre-MS cases sampled in the last 5-year period before clinical onset, 2 were EBV seronegative, compared with 12 in the control group (p=0.01; table 2). Samples acquired earlier than 5 years before MS onset did not differ significantly (table 2).

**EBV serostatus and sNfL**

Among the EBV-seropositive samples, the sNfL levels were higher in the pre-MS group than in the control group (ratio: 1.14; 95% CI 1.07 to 1.22; p=8.10\(^{-7}\)) for the whole study time frame, and for the last 10-year period before clinical MS onset (ratio: 1.18; 95% CI 1.09 to 1.28; p=8.10\(^{-5}\)). Among the EBV-seronegative samples, which was a much smaller group, no difference was found, with a pre-MS:control sNfL ratio of 1.12 for the whole study timeframe (n=36; 95% CI: 0.93 to 1.34; p=0.22) and 1.02 (n=7; 95% CI: 0.52 to 1.99; p=0.95) for the last 10-year period before MS onset.

Within the pre-MS group, 39 of 543 pre-MS samples with sNfL available were EBV seropositive. The highest sNfL concentration in this subgroup was 13.9 pg/mL. However, 52 of the 504 EBV-seropositive pre-MS cases (10%) had sNfL values above the cut-off of 13.9 pg/mL. That proportion was significantly higher than the expected value calculated from the distribution in the EBV-seronegative group (2.5%=12.6, p=0.017; age-adjusted p=0.038) (figure 1). For controls, the highest sNfL level in the EBV-seronegative group was 14.7 pg/mL. Of the 495 EBV-seropositive controls, 23 (4.6%) had sNfL values>14.7 pg/mL.

**EBV antibody levels**

Among EBV-seropositive samples, EBNA1, VCAmp18 and gp350 seroreactivities were significantly higher in the pre-MS group.

**Table 2** Discordant pairs for EBV seropositivity

<table>
<thead>
<tr>
<th>Years to clinical MS onset</th>
<th>Pre-MS pos</th>
<th>Pre-MS neg</th>
<th>P value*</th>
<th>Both EBV pos</th>
<th>Both EBV neg</th>
<th>N (pairs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;5</td>
<td>38</td>
<td>33</td>
<td>0.64</td>
<td>590</td>
<td>8</td>
<td>669</td>
</tr>
<tr>
<td>5–10</td>
<td>12</td>
<td>6</td>
<td>0.24</td>
<td>167</td>
<td>3</td>
<td>188</td>
</tr>
<tr>
<td>&gt;10</td>
<td>14</td>
<td>25</td>
<td>0.11</td>
<td>215</td>
<td>5</td>
<td>259</td>
</tr>
</tbody>
</table>

Discordant pairs: one of the samples (case or control) was EBV seronegative and the other EBV seropositive, as defined in the Methods section.

*McNemar test.

EBV, Epstein-Barr virus; MS, multiple sclerosis; neg, negative; pos, positive.
than in the matched control group (p=0.005 to <0.0001; table 3).

EBNA1 seroreactivity was elevated in pre-MS cases from 15 to 20 years before MS onset and significantly higher than among control samples from 10 to 15 years before clinical MS onset. EBNA1 seroreactivity remained elevated at approximately the same higher level in the intervals of 5–10 and 0–5 years before MS onset (figure 2A). Gp350 seroreactivity displayed a similar pattern, with a possibly later increase. For VCAp18, we detected no significant increase in the corresponding 5-year intervals (figure 2B).

EBV antibody levels and sNfL levels
In the pre-MS group elevated sNfL was detectable 5–10 years before clinical MS onset, approximately a decade after the increase in EBV seroreactivity (figure 2).

We observed no correlation between the level of sNfL and EBV seroreactivity for EBV-seropositive pre-MS group with an sNfL concentration higher than the highest value in the EBV seronegative pre-MS group (13.9 pg/mL before age adjustment); n=52 before and n=42 after age adjustment. Compared with the expected value (504/(39+1)=12.6) under the null hypothesis of equal distribution, the difference for sNfL was significant both before and after age adjustment, indicating a higher percentage of samples with elevated sNfL in the EBV-seropositive versus EBV-seronegative pre-MS group. The same calculation comparing EBV seropositives and negatives in the matched control group showed no significant differences. EBV, Epstein-Barr virus; ns, not significant; pre-MS, before multiple sclerosis diagnosis; sNfL, serum neurofilament light.

Table 3 EBV and ANO2 antibodies and sNfL in EBNA1-positive case–control pairs

<table>
<thead>
<tr>
<th></th>
<th>Pre-MS</th>
<th>Control persons</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N (pairs)</td>
<td>Median IQR</td>
</tr>
<tr>
<td>EBNA1 (kMFI)</td>
<td>557</td>
<td>9.1 (7.6–11)</td>
</tr>
<tr>
<td>VCAp18 (kMFI)</td>
<td>557</td>
<td>7.7 (5.2–9.9)</td>
</tr>
<tr>
<td>Gp350 (OD)</td>
<td>477</td>
<td>1.8 (0.9–2.6)</td>
</tr>
<tr>
<td>sNfL (pg/mL)</td>
<td>427</td>
<td>6.6 (4.9–9.4)</td>
</tr>
<tr>
<td>ANO2 (kMFI)</td>
<td>557</td>
<td>9.2 (2.4–62)</td>
</tr>
</tbody>
</table>

Only case–control pairs in which both samples were EBNA1 seropositive are included. The ANO2 comparison includes values below the ANO2 cut-off.

ANO2 serostatus in relation to EBNA1 serology
Only one individual in the EBNA1-seronegative group was seropositive for ANO2, with an MFI value immediately above the cut-off. In the EBV-seropositive group, ANO2 seropositivity was observed in 16.7% of pre-MS cases and 10.0% of controls (p=0.001).

Focusing on sample pairs collected less than 15 years before clinical MS onset, we studied the role of ANO2 by counting all combinations of positivity for EBNA1 and ANO2 in cases and controls (table 4).

Of 87 EBNA1-seropositive case–control pairs, 39 were positive for ANO2, and 48 were negative for ANO2. This finding suggested that ANO2 seropositivity was linked to a further increased risk of subsequent MS in EBNA1-seronegative individuals. Furthermore, of 63 ANO2-seronegative pairs discordant for EBNA1, we found that EBNA1 was seropositive in 39 pre-MS cases compared with 24 controls (p=0.038), suggesting increased risk with EBNA1 seropositivity, independent of ANO2.

The age distribution was almost identical for ANO2-seropositive and ANO2-seronegative samples within EBV-seropositive pre-MS cases and within controls (p=0.68, online supplemental figure S1). If conversion to ANO2 seropositivity occurred well after EBNA1 seroconversion, a number of individuals would be ANO2 seropositive at a higher age, leading to a right-shifted age distribution. As we identified no difference in the age distribution (online supplemental figure S2), we can infer that ANO2 seropositivity developed shortly after seroconversion to EBV positivity.

For samples positive for both ANO2 and EBNA1, serum ANO2 levels were weakly correlated with EBNA1 (r=0.31, p=0.006). This correlation was attenuated and not significant if an outlier was removed (online supplemental figure S3). Among EBNA1-seropositive pre-MS cases, within 15 years from clinical

Figure 1  High sNfL values were predominately found in EBV-positive pre-MS samples. Number of samples in the EBV-seropositive pre-MS group with an sNfL concentration higher than the highest value in the EBV seronegative pre-MS group (13.9 pg/mL before age adjustment); n=52 before and n=42 after age adjustment. Compared with the expected value (504/ (39+1)=12.6) under the null hypothesis of equal distribution, the difference for sNfL was significant both before and after age adjustment, indicating a higher percentage of samples with elevated sNfL in the EBV-seropositive versus EBV-seronegative pre-MS group. The same calculation comparing EBV seropositivities and negatives in the matched control group showed no significant differences. EBV, Epstein-Barr virus; sNfL, serum neurofilament light.
MS onset, ANO2 seropositivity was associated with a 26% higher sNfL (p=0.0026).

**DISCUSSION**

Here, we analysed seroreactivity against lytic and latent EBV antigens and against the putative EBNA1-ANO2 cross-antigen, as well as sNfL levels in the presymptomatic phase of MS. We leveraged serum repositories representing a wide age range in a cross-matching procedure with a Swedish national MS register. Our findings show increased EBV antibody levels in seropositive pre-MS individuals from 15 to 20 years before clinical MS onset, followed by increased sNfL from approximately 5–10 years before onset, about a decade after EBV seroreactivity was detected. Antibodies against the putative autoantigen ANO2 appeared almost solely in the EBNA1-seropositive group. This seroreactivity developed soon after EBNA1 seroconversion, but only in a limited proportion of participants: 17% of pre-MS individuals and 10% of matched controls. As expected, the EBV-seronegative subgroup continuously diminished in size during the time leading up to clinical MS onset. Pre-MS individuals with the highest sNfL levels were concentrated in the EBV-seropositive subset.

We relied on one sampling date per individual, without longitudinal information. After an increased EBV antibody level was reached in pre-MS cases 10–15 years before clinical onset, similarly increased levels of antibodies were detected in individuals sampled in the subsequent periods leading up to MS onset. This observation supports the idea that presymptomatically increased
EBV antibodies reflect an acute preceding event, conceivably the primary EBV infection, rather than resulting from a proliferating EBV infection, which would be expected to generate increasing EBV seroreactivity along with increasing sNfL levels. A previous study showed an approximately 4-fold higher EBNA1 antibody titre at initial sampling in a small pre-MS group compared with EBV-seropositive controls (table 2 in 25) further supporting that increased EBV seroreactivity before clinical MS onset originates from a primary infection. How many of these primary infections manifested as IM is not known. EBNA1 is a latency antigen, described as a dominant antigen in serological studies in clinical26 and presymptomatic MS.18 27 VCA antibody reactivity, however, has been associated with other diseases, such as systemic lupus erythematosus,28 post-transplantation lymphoproliferative disorders29 and Hodgkin’s lymphoma.30

We also evaluated gp350 as a major pre-MS lytic antigen with an established neutralising capacity that elicits persistent immunoreactivity in convalescence 10 years after IM compared with asymptomatic primary infection.31 We found that neutralising gp350 antibodies did not correlate with antibody levels against latent EBNA1. This lack of correlation between lytic gp350 and latent EBNA1 seroreactivity also has been reported in patients diagnosed with MS.32 In the absence of clinically discernible EBV reactivations in preclinical MS, the increased gp350 seroreactivity demonstrated here may reflect the primary lytic infection or intermittent subclinical lytic reactivations of EBV32 perhaps representing an indirect indication of failing cellular immunity against EBV.33

Previous studies showed a lag of clinical MS after primary EBV infection. Serological studies from repositories showed EBV infection 1–2 decades before MS at the group level.8 10 Epidemiological studies showed an average lag of 10–20 years with wide distribution from IM to MS onset.34 35 We here show a decade of lag at the group level between the primary EBV infection and incipient neurodegeneration and a steady level of new primary infections during the preonset decade of increasing axonal injury. This indicates that MS pathology tends to evolve during the latency phase of an EBV infection. Pathology during the preclinical phase may result from secretion of inflammatory mediators (EBERS, several microRNAs), which occurs even in latency 0–1 of the EBV infectious cycle, when no EBV proteins are otherwise produced, or from non-proliferating reactivations. In subsequent MS pathology, such humoral factors may be relevant for cortical lesions without T-cell infiltrates and immunoglobulins at some distance from meningeal germinal centres.36 37 Furthermore, the lag between the primary EBV infection and MS-related axonal degeneration leaves time for possible influence by further exogenous risk factors for MS that also interact with EBV.20 23 27 38

According to contemporary opinion, MS is a central nervous system-specific autoimmune disease, although the search for an autoantigen has been unrewarding. Recently, cases with EBV-associated autoreactivity were identified in subsets of patients with MS. CSF serological analysis revealed cross-reactivity between a glial cell adhesion molecule and another EBNA1 epitope in 20%–25% of patients with MS.39 T-cell immunity depends on other EBV epitopes, and HLA-restricted CD4+ T cells might be primed against two EBV-specific peptides and the CNS antigen RASGRP2.40 ANO2, a chloride channel protein that is important in several cell types and expressed in glial cells and neurons, has recently been investigated as an autoantigen in MS. ANO2 autoimmune reactivity in MS was demonstrated by screening a large number of antigens, and ANO2 antigen showed increased presence in the MS brain.15 Another previous study16 used a reciprocal absorption test showing that EBV antigen was able to inhibit antibodies induced by ANO2 and vice versa. Also, the cross-reactivity was confirmed with different assays, using both short peptides and a long ANO2 1–365 segments as antigen.15 16 The carriage of an ANO2 reactivity associated to an increased risk of MS in all types of combinations with HLA MS risk genes and EBNA1 reactivity,16 supporting a pathogenic role of this particular mimicry antigen. ANO2 is an intracellular antigen, and it is likely that it does not directly confer CNS damage, however, it may denote a pathogenic T cell reactivity. In this study, increased ANO2 seroreactivity did not appear until after EBV seroconversion and was limited to a subset of EBV-seropositive participants. Preonset ANO2 seroreactivity had an independent association with MS and with preonset axonal injury beyond the association with EBV infection. This relationship may result from a cross-reactive mechanism between EBNA1 and ANO2 as described in previous work.15 16 However, the ANO2 reactivity may also be an aspect of altered EBNA1 reactivity,7 with possible epitope spreading of the EBNA1 IgG antibodies. The importance of the altered reactivity, due to epitope spreading or cross-reactivity, is its association with increased preclinical sNfL, suggesting that it contributes to pre-MS axonal injury. Our finding of virtually no ANO2-seropositive individuals in EBNA1-seronegative groups in either the pre-MS or control cohorts may indicate the presence of a basal autoimmune diathesis in certain individuals in the general population.18 EBV-infected memory B cells may constitute a ‘forbidden clone’, with low but long-term propensity for autoimmunity.11

The strength of this study is the large repository material, matched with a national MS register, and sNfL assays performed

### Table 4 Concordant and discordant pairs for EBNA1 and ANO2 seropositivity in samples collected <15 years before clinical onset

<table>
<thead>
<tr>
<th>Matched controls</th>
<th>EBNA1−, ANO2−</th>
<th>EBNA1−, ANO2+</th>
<th>EBNA1+, ANO2−</th>
<th>EBNA1+, ANO2+</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pre-MS</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>EBNA1−, ANO2−</td>
<td>9</td>
<td>0</td>
<td>24</td>
<td>2</td>
<td>35</td>
</tr>
<tr>
<td>EBNA1−, ANO2+</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>EBNA1+, ANO2−</td>
<td>39</td>
<td>0</td>
<td>387</td>
<td>29</td>
<td>431</td>
</tr>
<tr>
<td>EBNA1+, ANO2+</td>
<td>7</td>
<td>0</td>
<td>58</td>
<td>1</td>
<td>91</td>
</tr>
<tr>
<td>Total</td>
<td>55</td>
<td>0</td>
<td>449</td>
<td>54</td>
<td>558</td>
</tr>
</tbody>
</table>

Concordant (grey shading) and discordant matched pairs for EBNA1 and ANO2 seropositivity and seronegativity. Showing that in the last 15-year period before clinical MS onset there are more EBNA1 seropositive samples with also seropositive ANO2 in the pre-MS group (n=58) than in the control group (n=29, p=0.002). There are also more EBNA1 seropositive samples in the ANO2 seronegative pre-MS group (n=39) than in ANO2 seronegative control group (n=24, p=0.038). Numbers used in comparisons in bold. P values from McNemar test.

ANO2, anoctamin 2; EBNA1, Epstein-Barr virus nuclear antigen 1; MS, multiple sclerosis.
in aliquots from the same specimens as latency and neutralising EBV activity and ANO2 reactivity assays. Our material captures a lengthy retrospective period extending well before clinical MS onset, incorporating the long average intervals between IM and MS of 10 years to decades.44 45

A weakness of this study is the lack of individual longitudinal data, which precluded observation of the exact individual sequence of seroconversions and sNfL. Data in the Swedish MS register are generally recorded by neurologists and quality controlled. However, some pre-MS samples were likely drawn during the MS prodrome with undetected subtle initial focal symptoms.46 This limitation is common to all studies of preclinical events. Still, this was not crucial for this study, which focused on changes in EBV seroreactivity presenting at least a decade before an objective biochemical sign of MS.

A caveat is that the temporal relationships disclosed here do not exclude the presence of a confounder that could have been evoked by reduced function of natural killer, natural killer T or CD8+ cells. Low natural killer cell numbers also have been associated with MS.43 44 Increased and atypical EBV serology in pre-MS may trace to a relatively more severe primary infection, either because of a higher viral load, possibly dependent on the oral infection pathway, or an insufficient CD8+ T cell response.33 The subsequent control of the EBV survival also depends on cellular immunity, including CD8+ lymphocytes activated as cytotoxic T lymphocytes. The HLA dependence of IM and MS may influence the pre-MS course through infection control in these phases.45

Because the level of sNfL decreases with higher body mass index (BMI)46 and adolescent overweight is a risk factor for MS,47 BMI is a source of possible bias. We did not have access to BMI data, but any such bias would have led to an underestimation of sNfL levels in this study.

The main results of this study are that EBV seroreactivity appeared a decade prior to the first signs of neuroaxonal injury, which in turn concentrated in the EBV seropositive group during the last decade before the clinical onset of MS. Furthermore, ANO2 antibodies, appearing in a subgroup shortly after primary EBV infection, were associated with preclinical neuroaxonal damage. These relationships implicate latent EBV infection in the pathogenesis of MS, at least as an interactive agent.

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Contributors DI is the guarantor of the study and accepts full responsibility for the work and the conduct of the study, had access to the data, and controlled the decision to publish. DI, PS and OFA contributed to the conception and design of the study; all authors contributed to the acquisition and analysis of data; DI, SN, PS and OFA contributed to drafting the text and preparing the figures. SN and DI performed the statistical analysis. All authors discussed the results, contributed to, and approved the final manuscript.

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Competing interests HZ has served on scientific advisory boards and/or as a consultant for AbbVie, Alexion, ALZPath, Annexon, Apellis, Artery Therapeutics, AZTherapeutics, CogBi, Denali, Eisai, NervGen, Nano Nordisk, Pint Therapeutics, Red Abbey Labs, remYND, Passage Bio, Roche, Samumed, Siemens Healthineers, Triplet Therapeutics, and Wave; has given lectures in symposia sponsored by Cellectircon, Fujirebio, AlzeCure, Biogen in the Swedish Government and County Councils, the JPMCC, and is a cofounder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is part of the GU Ventures Incubator Program, outside the work presented in this paper. TO has received advisory board/lecture honoraria from Biogen, Novartis, Merck, and Sanofi. The same companies have provided unrestricted MS research grants. MB has received a speaker fee from Biogen. PS will serve as an unpaid consultant for Moderna. KB has served as a consultant, on advisory boards, or on data monitoring committees for Biogen, Astex, BioArctic, Biogen, JOMDSD/SHimadzu, Julius Clinical, Lilly, MagQu, Novartis, Ono Pharma, Pharmatrophix, Prothena, Roche Diagnostics, and Siemens Healthineers, and also is a cofounder of Brain Biomarker Solutions in Gothenburg AB (BBS).

Patient consent for publication Not applicable.

Ethics approval This study involves human participants and the study was performed in accordance with the Declaration of Helsinki and approved by the research ethics board of Umeå, Sweden (2011-198/31, approved 3 August 2011; addendum 2017-484-32M, approved 18 January 2018, addendum 2019-03402, approved 19 August 2019). Participants gave informed consent to participate in the study before taking part.

Provenance and peer review Not commissioned; externally peer reviewed.

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Multiple sclerosis

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Figure S1. Intercorrelations between EBV antibodies and correlation with sNfL in pre-MS samples. Black circles indicate double (i.e., EBNA1 and VCAp18) seropositive pairs included in the correlation analyses in a–f; gray circles indicate pairs singly or not seropositive and not used for the present correlations; and dashed lines indicate cut-off for seropositivity as defined. (A and B) VCAp18 correlated weakly with EBNA1 and gp350. (C) No correlation was seen between EBNA1 and gp350 or between (D and F) sNfL and EBV serologies.

Abbreviations: EBNA1, Epstein–Barr virus nuclear antigen 1; EBV, Epstein–Barr virus; gp350, glycoprotein 350; OD, optical density; pre-MS, before multiple sclerosis diagnosis; sNfL, serum neurofilament light; MFI, median fluorescent intensity; VCAp18, viral capsid antigen protein 18.
Figure S2. Age distribution of ANO2-seropositive and -negative samples within the EBV-seropositive group. (A) Controls. (B) Pre-MS cases. Abbreviations: ANO2, anoctamin 2; EBV, Epstein–Barr virus; pre-MS, before multiple sclerosis diagnosis.

Figure S3. Correlation between ANO2 and EBNA1 antibodies and correlation with sNfL in pre-MS samples. Correlation coefficients were calculated for pre-MS samples seropositive for both ANO2 and EBNA1 (black circles). Correlation given in (A) included an outlier (far top right) and decreased to $r=0.20$ ($p=0.09$) after this data point was removed. Dotted lines indicate cut-off for ANO2 seropositivity as defined, and unbroken lines indicate regression on EBNA1 kMFI and sNfL with ANO2 kMFI as the dependent variable. ANO2, anoctamin 2; EBNA1, Epstein–Barr virus nuclear antigen 1; MFI, median fluorescent intensity; pre-MS, before multiple sclerosis diagnosis; sNfL, serum neurofilament light.